The Microtubule-Stabilizing Agent Discodermolide Competitively Inhibits the Binding of Paclitaxel (Taxol) to Tubulin Polymers, Enhances Tubulin Nucleation Reactions More Potently than Paclitaxel, and Inhibits the Growth of Paclitaxel-Resistant Cells

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SUMMARY

The lactone-bearing polyhydroxylated alkatetraene (+)-discodermolide, which was isolated from the sponge Discodermia dissoluta, induces the polymerization of purified tubulin with and without microtubule-associated proteins or GTP, and the polymers formed are stable to cold and calcium. These effects are similar to those of paclitaxel (Taxol), but discodermolide is more potent. We confirmed that these properties represent hypernucleation phenomena; we obtained lower tubulin critical concentrations and shorter polymers with discodermolide than paclitaxel under a variety of reaction conditions. Furthermore, we demonstrated that discodermolide is a competitive inhibitor with [3H]paclitaxel in binding to tubulin polymer, with an apparent Ki value of 0.4 μM. Multidrug-resistant human colon and ovarian carcinoma cells overexpressing P-glycoprotein, which are 900- and 2800-fold resistant to paclitaxel, respectively, relative to the parental lines, retained significant sensitivity to discodermolide (25- and 89-fold more resistant relative to the parental lines). Ovarian carcinoma cells that are 20–30-fold more resistant to paclitaxel than the parental line on the basis of expression of altered β-tubulin polypeptides retained nearly complete sensitivity to discodermolide. The effects of discodermolide on the reorganization of the microtubules of Pororous tridactylis kidney epithelial cells were examined at different times. Intracellular microtubules were reorganized into bundles in interphase cells much more rapidly after discodermolide treatment compared with paclitaxel treatment. A variety of spindle aberrations were observed after treatment with both drugs. The proportions of the different types of aberration were different for the two drugs and changed with the length of drug treatment.

Recently, an ever-increasing number of potently cytotoxic natural products that interact with tubulin have been isolated (see Ref. 1 for a review). Cells treated with these compounds arrest in the mitotic phase of the cell cycle. At medium-to-higher drug concentrations, this occurs because the microtubule spindle fails to form, whereas, at lower concentrations, these agents seem to interfere with spindle microtubule dynamics (2), apparently prolonging metaphase. Most of these antimitotic agents also cause the disappearance of intracellular microtubules. Such compounds invariably inhibit the assembly of microtubule protein or purified tubulin, often at concentrations substantially below the tubulin concentration in the reaction mixture.

A few antimitotic drugs, however, cause massive reorganization of intracellular microtubules and promote biochemical assembly reactions of both microtubule protein and purified tubulin. For more than 15 years, paclitaxel and structurally related taxoids were the only compounds known to have this mechanism of action (3). Recently, two additional drug classes have been shown to act by a similar mechanism. A computer-assisted search for novel compounds with structural analogy to colchicine site inhibitors suggested that the marine natural product (+)-discodermolide (4, 5), which was isolated from the sponge Discodermia dissoluta, could be an antimitotic agent (6), and initial studies with the compound had demonstrated potent antiproliferative activity (low

ABBREVIATIONS: MDR, multidrug resistant (resistance); MAP, microtubule-associated protein; MES, 2-(N-morpholino)ethanesulfonic acid; FCS, fetal calf serum; PBS, phosphate-buffered saline; PtK2, Pororous tridactylis kidney epithelial.
nanomolar IC₅₀ values), with cell cycle arrest at G2/M (7). However, the treatment of breast carcinoma cells with nanomolar concentrations of discodermolide resulted in spectacular microtubule bundle formation, even more extensive than that observed with micromolar concentrations of paclitaxel. Studies with purified tubulin confirmed that discodermolide was significantly more potent than paclitaxel in inducing polymerization under a variety of reaction conditions (8). The activity of epothilones A and B, isolated from the soil bacterium Sorangium cellulosum, was discovered through a screen of almost 8000 natural product extracts for compounds that would polymerize microtubule protein, and the purified compounds, like paclitaxel, caused extensive rearrangement of microtubules in cells (9). The structures of discodermolide and the epothilones are compared with that of paclitaxel in Fig. 1.

The diverse, seemingly unrelated, structures of these molecules raised the question of whether they bind to the same

![Discodermolide](image1)

Discodermolide

![Paclitaxel](image2)

Paclitaxel

![Epothilone A](image3)

Epothilone A: \( R = H \)

![Epothilone B](image4)

Epothilone B: \( R = CH_3 \)

Fig. 1. Structures of (+)-discodermolide, paclitaxel, and epothilones A and B. The configurations of the chiral centers of discodermolide are those shown in Ref. 38. The configurations of the chiral centers of epothilones A and B are those shown in Ref. 39.

or different sites on tubulin. Radiolabeled taxoids seem to bind with high affinity only to polymerized forms of tubulin, and inhibitors of assembly usually simultaneously prevent paclitaxel-driven polymerization and binding of taxoid to tubulin (10–12). The epothilones (9, 13) and discodermolide (14, 15) were shown to inhibit the binding of \(^{3}H\)paclitaxel to tubulin polymers. Recently, we reported that epothilones A and B displayed kinetic inhibitory patterns observed with competitive inhibitors (13), indicating they bind to the paclitaxel site on polymer. A similar kinetic characterization of discodermolide binding was one of the goals of the current study, as was further characterization of the potent activity of discodermolide in nucleating microtubule assembly (8).

Paclitaxel and its semisynthetic analog docetaxel (Taxotere) have become increasingly important in the treatment of neoplastic diseases (16, 17), and it is to be hoped that discodermolide and the epothilones will have similar value. Paclitaxel is a substrate for P-glycoprotein, whose overexpression can be responsible for the MDR phenomenon. Epothilones A and B do not seem to bind to P-glycoprotein; a number of MDR cell lines that have lost their sensitivity to paclitaxel retain nearly complete sensitivity to both epothilones (9, 13, 18, 19). In addition, we found that the paclitaxel-resistant ovarian carcinoma cell lines 1A9PTX10 and 1A9PTX22, which express altered \( \beta \)-tubulins\(^1\) (20), retained full sensitivity to epothilone B, whereas only 1A9PTX22 cells retained full sensitivity to epothilone A (13, 19). We evaluated the activity of discodermolide in two MDR carcinoma cell lines and in the paclitaxel-resistant 1A9PTX10 and 1A9PTX22 ovarian carcinoma lines. Further, microtubule bundles appear more rapidly in cells treated with discodermolide than in cells treated with paclitaxel. This complements the earlier observation that bundles appear after treatment of cells with significantly lower concentrations of discodermolide than of paclitaxel (8).

**Experimental Procedures**

**Materials.** Discodermolide was isolated from *D. dissoluta* as previously described (21). Paclitaxel and \(^{3}H\)paclitaxel (19–23 mCi/mmol) were generously provided by the Drug Synthesis and Chemistry Branch, National Cancer Institute (Rockville, MD). Drugs were dissolved in dimethylsulfoxide, and control samples contained solvent equivalent to that in drug-treated cultures or reaction mixtures. Electrophoretically homogenous tubulin and heat-treated MAPs were prepared from bovine brain (22). Tubulin was freed from unbound nucleotide by gel filtration chromatography and concentrated as previously described (23). For the polymerization studies, 1.0 mM glutamate (used to stabilize the tubulin; can replace MAPs in inducing tubulin assembly) was removed through dialysis against 0.1 M MES, pH 6.9. The tubulin was centrifuged to remove aggregates and stored in liquid nitrogen in 0.2-ml aliquots. PtK cells were from American Type Culture Collection (Rockville, MD). The ovarian carcinoma line 1A9, a clone of line A2780 (24), was used to generate paclitaxel-resistant lines by incubating the cells with increasing concentrations of paclitaxel in the presence of verapamil (20). The ovarian MDR line A2780AD was also derived from line A2780 (25). The colon carcinoma line 1A9, a clone of line A2780 (24), was used to generate paclitaxel-resistant lines by incubating the cells with increasing concentrations of paclitaxel in the presence of verapamil (20). The ovarian MDR line A2780AD was also derived from line A2780 (25).

1 Both paclitaxel-resistant lines bear mutations in the M40 gene and express large amounts of its product, the \( \beta \)-tubulin isotype. In 1A9PTX10 the mutation is at residue 270 (phenylalanine to valine; TTT to GTT) and in 1A9PTX22 at residue 364 (alanine to threonine; GCA to ACA).
Mixtures of discodermolide with \[^3H\]paclitaxel in varying assembly of tubulin into polymer (data not presented; see Refs. justed to pH 6.6 with HCl). This condition induced nearly complete PBS. Attached cells were fixed in incubation times, medium was removed, and cells were rinsed with attached cells were incubated in supplemented minimal medium containing 10 \(\mu\)M concentration of drug at 37°. After various incubation times, medium was removed, and cells were rinsed with 37° PBS. Attached cells were fixed in −20° methanol for 10 min, rinsed three times with PBS, and incubated in PBS containing 0.1% Triton X-100 and 10% FCS for 30 min at room temperature to block nonspecific antibody binding sites. The blocking buffer was removed, and the cells were incubated for 1 hr at 37° with antibody to \(\beta\)-tubulin (200 \(\mu\)g/well, 1:200 dilution blocking buffer). Cells were washed three times with PBS and incubated for 45 min with Texas Red-conjugated goat anti-mouse IgG (150 \(\mu\)g/well, 1:50 dilution in PBS). After removal of the secondary antibody by two washes with PBS, diaminopropidium iodide (2 mg/ml) dissolved in PBS was added to stain DNA. Slides were wet-mounted in ProLong according to the manufacturer's procedures. Images were obtained with a Nikon Opo...

**Inhibition of human colon and human ovarian carcinoma cell growth.** Cells were continuously incubated in RPMI medium containing 10% FCS, 12 mg/ml gentamicin sulfate, and 2 mM glutamine. The medium for the paclitaxel-resistant ovarian cells also contained 5 mg/ml verapamil (to inhibit expression of P-glycoprotein) and 15 ng/ml paclitaxel. Before drug treatment, the paclitaxel-resistant ovarian cells were removed from paclitaxel containing medium for 5–7 days. Drug effects were determined in 96-well microtiter plates, with cells fixed and stained for protein (27).

**Indirect immunofluorescence.** Confluent monolayers of Ptk2 cells, grown on 25-cm\(^2\) plates, were suspended by incubation with 7 ml/plate of 0.05% trypsin and 0.53 mM EDTA in Hanks’ balanced salt solution. Suspended cells were diluted 1:40 with minimal essential medium containing 1 mM sodium pyruvate, 0.1 mM nonessential amino acids, 10% FCS, 2 mM glutamine, and 10 \(\mu\)g/ml gentamicin sulfate and allowed to attach and grow for 24 hr at 37° in a humid 5% CO\(_2\) atmosphere on sterile, eight-well Permanox or glass cell-culture slides (300 \(\mu\)g/well, 100 cells/well). Medium was removed, and attached cells were incubated in supplemented minimal essential medium containing 10 \(\mu\)M concentration of docetaxel at 37°. After various incubation times, medium was removed, and cells were rinsed with 37° PBS. Attached cells were fixed in −20° methanol for 10 min, rinsed three times with PBS, and incubated in PBS containing 0.1% Triton X-100 and 10% FCS for 30 min at room temperature to block nonspecific antibody binding sites. The blocking buffer was removed, and the cells were incubated for 1 hr at 37° with antibody to \(\beta\)-tubulin (200 \(\mu\)g/well, 1:200 dilution blocking buffer). Cells were washed three times with PBS and incubated for 45 min with Texas Red-conjugated goat anti-mouse IgG (150 \(\mu\)g/well, 1:50 dilution in PBS). After removal of the secondary antibody by two washes with PBS, diaminopropidium iodide (2 mg/ml) dissolved in PBS was added to stain DNA. Slides were wet-mounted in ProLong according to the manufacturer's procedures. Images were obtained with a Nikon Opo...

**Electron microscopy.** During polymerization reactions, 10-\(\mu\)l aliquots were removed at various times and placed onto 200-mesh carbon-coated, Formvar-treated copper grids. Samples were stained with several drops of 0.5% uranyl acetate and examined with a Zeiss model 10CA electron microscope.

**Results**

**Competitive binding of discodermolide and paclitaxel to microtubules.** We showed that discodermolide and paclitaxel caused Burkitt lymphoma cells to accumulate in mitotic arrest and led to rearrangement of the microtubule network of breast carcinoma cells. Both agents caused hypernucleation of tubulin polymerization in reactions with reduced requirements for GTP, MAPs, and higher incubation temperatures, and the polymers formed had increased stability to cold and calcium (8). These properties had been often described for paclitaxel (3, 12, 23, 31–34). Finally, discodermolide, like taxoids and the epothilones, inhibited the binding of \[^3H\]paclitaxel to tubulin polymer (14). In most respects, however, discodermolide seemed to be significantly more potent than paclitaxel, with more extensive reactions occurring at lower temperatures and drug concentrations (8), and discodermolide was particularly potent in inhibiting the binding of \[^3H\]paclitaxel to tubulin polymer (14). Hung et al. (15), using synthetic discodermolide, reported similar observations and also noted that paclitaxel only weakly inhibited the binding of \[^3H\]discodermolide to polymer.

The disparate structures of discodermolide and paclitaxel caused us to undertake studies to gain more specific information about their binding sites. Success in these experiments required that virtually all tubulin in the reactions be driven into polymer before the addition of \[^3H\]paclitaxel and inhibiting drugs. We accomplished this by using ddGTP to induce tubulin assembly (28–30). In the kinetic experiments summarized in Fig. 2, we demonstrated that discodermolide apparently is a competitive inhibitor of the binding of \[^3H\]paclitaxel to the polymer. (A similar finding, as a control experiment, was made with docetaxel, generously provided by Dr. D. G. I. Kingston.) Fig. 2A presents our binding data in the Hanes format, in which a competitive inhibitor generates a family of parallel curves at different inhibitor concentrations (35). Dixon analysis (35) of these data yielded an apparent \(K_i\) value of 0.4 \(\mu\)M for discodermolide (Fig. 2B); the value obtained for docetaxel was 0.3 \(\mu\)M.

**Discodermolide nucleates microtubule assembly more potently than paclitaxel.** Although many of the effects of paclitaxel on microtubule assembly (polymerization at low temperatures and without GTP and/or MAPs) probably are manifestations of the ability of paclitaxel to hypernucleate tubulin assembly, this property is directly demonstrated by the more extensive polymerization, shorter microtubules, and lower tubulin critical concentration observed with the drug. With paclitaxel analogs, we found excellent correlation of enhanced assembly effects with relative tubulin critical concentrations (23); we have now also found that among paclitaxel, docetaxel, and 2-debenzoyl-2-meta-azidobenzoylpaclitaxel, relative microtubule lengths decrease as taxod potency increases. Paclitaxel is the least active among the three compounds and yields the longest...
microtubules, whereas 2-debenzoyl-2-meta-azidobenzylopaclitaxel is the most active and yields the shortest microtubules.\(^2\) Similarly, epothilone A had activity in inducing tubulin assembly and effects on tubulin critical concentration and microtubule lengths comparable to results obtained with paclitaxel, whereas epothilone B was distinctly more active (13).

In our initial studies with discodermolide (8), we found that its potency in inducing microtubule assembly reactions was significantly greater than that of paclitaxel. We have now determined the tubulin critical concentrations with discodermolide at 37\(^\circ\) under a variety of reaction conditions in comparison with paclitaxel (Table 1). The experimental data were analyzed by linear regression to generate the values shown in Table 1. The turbidity values obtained below 1 \(\mu\)M tubulin were too low to be reliable (versus instrument noise).

Several repetitions of experiments with 1 \(\mu\)M tubulin that yielded higher turbidity readings compared with reaction mixtures without drug convinced us that the critical concentrations for paclitaxel with MAPs plus GTP and for discodermolide with MAPs plus GTP, with MAPs only, and with GTP only were truly \(<1 \mu\)M.

With the tubulin and MAPs preparations used here, the tubulin critical concentration with 100 \(\mu\)M GTP was 3 \(\mu\)M (0.3 mg/ml). In previous experiments (23) without drug, MAPs or GTP, there was no assembly at tubulin concentrations up to 80 \(\mu\)M. With 10 \(\mu\)M paclitaxel, the tubulin critical concentration was \(<1 \mu\)M with both GTP and MAPs. The tubulin critical concentration increased to \(\sim 2 \mu\)M when GTP was not included in the reaction mixture, to \(\sim 4 \mu\)M without MAPs, and to \(\sim 20 \mu\)M without GTP and MAPs. This progressive increase in the critical concentration correlates well with the relative ability of tubulin to assemble with paclitaxel, particularly at low temperatures, under these different reaction conditions (8, 13, 23). With 10 \(\mu\)M discodermolide, the tubulin critical concentration was \(<1 \mu\)M under all reaction conditions except when neither GTP nor MAPs was in the reaction mixture, when a value of 3 \(\mu\)M was obtained. This was approximately one seventh of the value obtained with paclitaxel.

Polymer lengths were measured under all reaction conditions in which assembly occurred with 10 \(\mu\)M tubulin and 10 \(\mu\)M concentration of drug (Table 2). Data previously obtained (8) with both MAPs and GTP are also included in Table 2. In the MAPs-plus-GTP system, average polymer length without drug in both experiments was approximately twice that obtained with paclitaxel and 5–6-fold that obtained with discodermolide. With paclitaxel, there was a 50% increase in average microtubule length with MAPs only and a 3-fold increase with GTP only compared with the complete system. With discodermolide, compared with the complete system, there was a 50% increase in average length with MAPs only, a 2-fold increase with GTP only, and a 3-fold increase with neither MAPs nor GTP in the reaction.

Fig. 3 provides high-power views of the polymer formed with discodermolide under these four reaction conditions. With MAPs plus GTP (Fig. 3A), MAPs only (Fig. 3B), and no MAPs/no GTP (Fig. 3D), significant amounts of microtubule forms, mixed with ribbon polymers, were observed. With GTP only (Fig. 3C), most of the polymer was in the form of ribbons. The simultaneously obtained paclitaxel polymers were primarily of microtubule morphology (data not presented). Although the GTP-only sample with paclitaxel had a predominant microtubule morphology in the current study, we previously observed abundant ribbons under this reaction condition (23). The nuance in experimental conditions that results in this variability is not known.

Effects of discodermolide and paclitaxel on cultured human carcinoma cells. Table 3 summarizes the inhibitory effects of discodermolide and paclitaxel on the growth of several human tumor cell lines; these include two MDR lines to evaluate discodermolide as a potential substrate of P-

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\(^2\) R. J. Kowalski and E. Hamel, unpublished observations.
glycoprotein and two lines resistant to paclitaxel as a consequence of genetic alterations in a β-tubulin gene (20).

The parental colon carcinoma line SW620 was 10-fold more sensitive to paclitaxel than to discodermolide, but the MDR line derived from it was almost 4-fold more sensitive to discodermolide. The 930-fold increased resistance of the SW620AD-300 line to paclitaxel relative to the parental line was reduced to a 25-fold increased resistance to discodermolide. The parental ovarian line 1A9 was 4-fold less sensitive to discodermolide than to paclitaxel, but the related MDR line was almost 7-fold more sensitive to discodermolide. The 2800-fold increased resistance of the A2780AD line to paclitaxel relative to the 1A9 line was reduced to 89-fold increased resistance to discodermolide.

The ovarian lines 1A9PTX10 and 1A9PTX22 with altered β-tubulin gene products were >20-fold resistant to paclitaxel relative to the parental line. Both lines retained nearly complete sensitivity to discodermolide. As a consequence, these cells were significantly more sensitive to discodermolide than to paclitaxel.

**Indirect immunofluorescence studies.** Kangaroo rat PtK2 cells were used to compare the effects of discodermolide with those of paclitaxel on intracellular microtubules (Figs. 4 and 5). IC₅₀ values for this line, which were determined by counting both adherent and detached cells, were 250 and 120 nM for discodermolide and paclitaxel, respectively. To minimize concentration effects, cells were studied after treatment with 10 μM concentration of drug. We wanted to determine whether there was a difference in the time at which microtubule bundles appeared in drug-treated cells and to compare

<table>
<thead>
<tr>
<th>Drug</th>
<th>Average polymer length (relative to lengths with paclitaxel)</th>
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<tbody>
<tr>
<td></td>
<td>MAPs and GTP</td>
</tr>
<tr>
<td>None</td>
<td>4.3 ± 2 (2.3)</td>
</tr>
<tr>
<td>Paclitaxel</td>
<td>1.9 ± 1 (1.0)</td>
</tr>
<tr>
<td>Discodermolide</td>
<td>0.83 ± 0.4 (0.44)</td>
</tr>
</tbody>
</table>

* Data from Ref. 8, a separate experiment.

**TABLE 2**

<table>
<thead>
<tr>
<th>Drug</th>
<th>Average polymer length (relative to lengths with paclitaxel)</th>
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<tr>
<td></td>
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<tr>
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* Data from Ref. 8, a separate experiment.
the appearance of microtubules in mitotic cells after treatment with the two drugs. Cells were examined after drug treatment for 2, 6, 12, 24, and 48 hr.

Fig. 4 demonstrates that microtubule bundles appeared more rapidly after discodermolide treatment than after paclitaxel treatment, although both drugs ultimately caused microtubules in interphase cells to become nearly completely reorganized into bundles with multiple regions of nucleation.

TABLE 3
Inhibitory effects of paclitaxel and discodermolide on growth of human cancer cells

<table>
<thead>
<tr>
<th>IC_{50} (relative resistance)</th>
<th>Colon carcinoma</th>
<th></th>
<th>Ovarian carcinoma</th>
<th></th>
</tr>
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<tbody>
<tr>
<td></td>
<td>Parental</td>
<td>Paclitaxel resistant MDR</td>
<td>Parental</td>
<td>Paclitaxel resistant</td>
</tr>
<tr>
<td></td>
<td>SW620</td>
<td>SW620AD-300</td>
<td>1A9</td>
<td>A2780AD</td>
</tr>
<tr>
<td>Paclitaxel</td>
<td>0.28 ± 0.06/2</td>
<td>260 ± 10/2 (930)</td>
<td>1.4 ± 0.2/11</td>
<td>3900 ± 400/4 (2800)</td>
</tr>
<tr>
<td>Discodermolide</td>
<td>2.8 ± 0.4/3</td>
<td>70 ± 10/3 (25)</td>
<td>6.5 ± 1/7</td>
<td>580 ± 200/3 (89)</td>
</tr>
</tbody>
</table>

Fig. 4. Microtubule bundles form more rapidly in PtK2 cells treated with discodermolide than with paclitaxel. A, No drug. B and C, Discodermolide. D–F, Paclitaxel. Cells were grown for 2 (A, B, and D), 6 (C and E), or 12 (F) hr at 37° before fixation and staining. See text for further details. Magnification, 630×.
A control metaphase cell is shown in Fig. 5A, whereas aberrant mitotic figures after discodermolide treatment are shown in Fig. 5, B–H. Increased numbers of mitotic cells (those with condensed chromosomes and no nuclear membrane) were observed with both paclitaxel and discodermolide as early as 2 hr and, of the three time points examined, were maximal at 6 hr. All mitotic aberrations were present after treatment with both drugs, but their relative proportions varied between paclitaxel- and discodermolide-treated cells. Although the morphologies of the different types of aberrant “spindles” tended to blend into each other, making our classification scheme somewhat arbitrary, some mitotic aberrations occurred with sufficient frequency to permit the quantification of subtypes shown in Table 4. Paclitaxel produced more punctate-stained spindle aberrations, probably representing multiple microtubule organizing centers or spindle poles. Short filaments resembling astral microtubules were attached to the multiple spindle poles; these were called “aster spindles” (Fig. 5B but after discodermolide treatment). With discodermolide, the most common type of mitotic cell had a complex, spiral-like microtubule array, which is best appreciated by examining the cell at several focal planes. The microtubules in these cells were relatively long, and there was usually only one apparent or two poorly separated microtubule organizing centers. Such a “spiral spindle” is shown in Fig. 5C. Sometimes, however, the microtubule organizing centers were well separated, yielding a double spiral (Fig. 5D). Cells with spiral spindles, but in a lower proportion, were also observed after paclitaxel treatment. At 12 hr, a third type of mitotic aberration was ob-

TABLE 4

<table>
<thead>
<tr>
<th>Drug</th>
<th>Incubation time</th>
<th>Mitotic index</th>
<th>Aster spindles</th>
<th>Spiral spindles</th>
<th>Spike spindles</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>hours</td>
<td>%</td>
<td>%</td>
<td>%</td>
<td>%</td>
</tr>
<tr>
<td>Paclitaxel</td>
<td>2</td>
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<td>47</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>6</td>
<td>23</td>
<td>67</td>
<td>2</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>12</td>
<td>17</td>
<td>31</td>
<td>41</td>
<td>21</td>
</tr>
<tr>
<td>Discodermolide</td>
<td>2</td>
<td>8</td>
<td>21</td>
<td>37</td>
<td>0</td>
</tr>
<tr>
<td></td>
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<td>85</td>
<td>2</td>
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<tr>
<td></td>
<td>12</td>
<td>17</td>
<td>11</td>
<td>68</td>
<td>21</td>
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</table>

Fig. 5. Mitotic aberrations in PtK2 cells after treatment with discodermolide. A, Control cell in metaphase (no drug) observed after growth for 6 hr. B, Aster spindle, observed after 2 hr in discodermolide. C and D, Spiral spindles, observed after 6 hr in discodermolide. E, Spike spindle, observed after 24 hr in discodermolide. F–H, Unclassified spindle aberrations, observed after 6 hr in discodermolide. See text for further details. All photographs were taken under oil with a 100× objective (numerical aperture, 1.30). Magnification, 1300×.
served with both drugs ("spike spindles"). Cells with these structures had clusters of thick microtubule bundles, with the microtubules of intermediate length between those of the shorter aster spindle microtubules and the longer spiral spindle microtubules. In most cases, cells with spike spindles had one or two microtubule organizing centers (Fig. 5E). Separation between spike spindle microtubule organizing centers was highly variable. In addition, a significant number of mitotic cells could not be readily classified into one of these categories (Figs. 5, F–I). It is unknown whether there is any progression in these spindle variants or whether the proportions of specific spindle morphologies reflect different lengths of drug treatment relative to stage in the cell cycle for specific cells.

Discussion

Interaction of discodermolide with tubulin. The tubulin-based experiments were undertaken to determine whether the strong binding of discodermolide to tubulin polymers (8, 14, 15) occurred at the same site as paclitaxel binding and to confirm the hypothesis that the potent induction of tubulin assembly observed with discodermolide compared with paclitaxel represented superior activity in hypernucleating polymerization reactions.

With discodermolide, we obtained a competitive pattern of inhibition of binding of [3H]paclitaxel to tubulin polymer (Fig. 2), and a similar pattern was obtained with the taxoid docetaxel. Classically, such results are interpreted as indicating that "substrate" (paclitaxel) and "inhibitor" (discodermolide) bind in the same site, but overlapping sites and/or allosteric effects cannot be readily eliminated as possible alternative explanations. The apparent Ki values obtained for discodermolide (0.4 μM) and docetaxel (0.3 μM) were essentially identical, even though discodermolide is the more potent compound in inducing tubulin assembly.

Both tubulin critical concentrations and polymer lengths were lower with discodermolide than with paclitaxel under every reaction condition examined in which reliable quantitative measurements could be made (Tables 1 and 2). Thus, with discodermolide, as with taxoids and the epothilones, polymer length data and tubulin critical concentrations support the concept that the reduced requirements for MAPs and GTP result from the ability of this broad class of drugs to hypernucleate tubulin assembly. The four systems we studied seem to have the following potency order for supporting nucleation: MAPs plus GTP > MAPs only > GTP only > no MAPs/no GTP.

The most reliable quantitative comparison between paclitaxel and discodermolide was probably the 7-fold-lower critical concentration obtained with discodermolide in the absence of both MAPs and GTP. This was in excellent agreement with another assay we performed to obtain a quantitative measure of the relative potency of the two drugs, in which we found that 3.2 μM discodermolide and 23 μM paclitaxel induced 50% tubulin assembly in 0.4 μM glutamate without GTP (8), also a ∼7-fold difference. The two measures together perhaps indicate the relative affinities of the two drugs for tubulin. Because no polymer exists in either 0.1 M MES or 0.4 M glutamate without GTP or MAPs, these relative affinities may apply to the α,β-heterodimer rather than to polymerized forms of tubulin, but transient polymeric forms that bind the drugs and are in turn stabilized by them cannot be excluded as an alternate explanation.

In summary, the biochemical analysis is most consistent with the two drugs acting by a similar mechanism of action, with discodermolide having a higher affinity than paclitaxel for tubulin (probably 5–10-fold greater).

Effects of discodermolide on cellular microtubule morphology. Previous work with breast carcinoma cells showed that discodermolide more potently induced microtubule bundle formation than did paclitaxel (8). Despite nearly equivalent IC50 values of the two drugs in the cell lines studied, bundle formation observed with 10 nM discodermolide required 1 μM paclitaxel. Few mitotic cells were observed in the breast carcinoma cultures, but both interphase and mitotic Swiss 3T3 cells were reported to have multiple bundles of microtubules after discodermolide treatment (15). In our study with PtK2 cells (Fig. 4), we found that microtubule bundling occurred much more rapidly with discodermolide than with paclitaxel treatment, but that after 12 hr, the appearance of interphase cells treated with the two drugs was identical. We also observed that mitotic PtK2 cells (no nucleus; condensed chromosomes) differed dramatically in appearance from interphase cells (with nuclei) after treatment with the two drugs. A variety of microtubule patterns was observed in the mitotic cells, and their distribution changed as a function of incubation time in the presence of drug (Table 4, Fig. 5). Precise proportions of different aberrant spindles differed with the two drugs and changed over time, but we could detect no discodermolide- or paclitaxel-specific spindle variant.

The enhanced bundling observed with discodermolide compared with paclitaxel is readily understood in terms of the apparent increased affinity of discodermolide for tubulin relative to paclitaxel. The differences in the proportions of mitotic aberrations seen with the two drugs and, indeed, the changing distribution of these aberrations as a function of time are not readily explained by the biochemical properties we examined. However, it seems probable that paclitaxel and discodermolide will differ either quantitatively or qualitatively in their effects on microtubule dynamics, and dynamic effects may be more important in mitotic cells than in interphase cells (36, 37). We should also note that the differing mitotic aberrations may also ultimately derive from relative affinities for tubulin. Epothilone B has a higher affinity for tubulin than epothilone A on the basis of apparent Ki values for inhibition of paclitaxel binding to polymer and its more potent enhancement of nucleation. After epothilone B treatment, PtK2 cells had large numbers of spiral spindles, whereas after epothilone A treatment, aster spindles were more prevalent (13). Thus, comparable distributions of mitotic aberrations were observed with the more potent agents (discodermolide and epothilone B), on the one hand, and with the less potent agents (paclitaxel and epothilone A), on the other. In support of this idea, the morphological effects of these four agents on the PtK2 cells was more closely related to their apparent affinities for tubulin (discodermolide > epothilone B > epothilone A > paclitaxel) than on their effects on growth of the cell line [epothilone A > epothilone B > paclitaxel > discodermolide (40, 44, 120, and 250 nM IC50 values, respectively)].

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Microtubule-Stabilizing Agent Discodermolide

References


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