Protective Effects of the Antiparkinsonian Drugs Talipexole and Pramipexole against 1-Methyl-4-phenylpyridinium-Induced Apoptotic Death in Human Neuroblastoma SH-SY5Y Cells

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ABSTRACT

Treatment of human neuroblastoma SH-SY5Y cells with 1 mM 1-methyl-4-phenylpyridinium (MPP+) for 3 days induced production of reactive oxygen species (ROS), followed by caspase-3 activation, cleavage of poly(ADP-ribose) polymerase (PARP), and apoptotic cell death with DNA fragmentation and characteristic morphological changes (condensed chromatin and fragmented nuclei). Simultaneous treatment with 1 mM talipexole slightly inhibited the MPP+-induced ROS production and apoptotic cell death. In contrast, pretreatment with 1 mM talipexole for 4 days markedly protected the cells against MPP+-induced apoptosis. However, this protective effect might not be mediated by dopamine receptors. The talipexole pretreatment induced an increase in antiapoptotic Bcl-2 protein levels but had no effect on levels of proapoptotic Bax, Bak, and Bad. It also inhibited MPP+-induced ROS production, p53 expression, and cleavages of caspase-3 and PARP. Similarly, pramipexole pretreatment increased Bcl-2 and inhibited MPP+-induced apoptosis. Although pretreatment with bromocriptine also had a protective effect against MPP+-induced apoptosis, it had no effect on the protein levels of Bcl-2 family members. On the other hand, N6,2’-O-dibutyryl cAMP or calphostin C induced a decreased Bcl-2 level and enhanced MPP+-induced cell death. These results suggest that talipexole has dual actions: (1) it directly scavenges ROS, affording slight protection against MPP+-induced apoptosis, and (2) it induces Bcl-2 expression, thereby affording more potent protection, if it is administrated before MPP+. Pramipexole has similar effects, whereas bromocriptine seems to exhibit the former but not the latter effect.

MPTP produces an irreversible and severe parkinsonian-like syndrome that causes selective degeneration of the nigrostriatal dopaminergic neurons in humans (Davis et al., 1979; Langston et al., 1983). This neurotoxin has been used to create animal models of Parkinson’s disease (Langston and Irwin, 1986). MPTP is converted by monoamine oxidase B in glial cells such as astrocytes to MPP+, which is accumulated intracellularly in neurons via a dopaminergic transporter (Snyder and D’Amato, 1986). Thus, MPP+ is an active metabolite of MPTP and a neurotoxin for dopaminergic neurons. The MPP+-induced neuronal death is caused by apoptosis in rat mesencephalic and striatal neurons (Mochizuki et al., 1994) and human neuroblastoma SH-SY5Y cells (Itano and Nomura, 1995). It also has been shown that apoptotic death of neurons occurs in the brain of patients with Parkinson’s disease (Mochizuki et al., 1996). The characteristic morphological and biological features of apoptosis are cellular shrinkage, membrane blebbing, chromatin condensation, and fragmentation of chromatin DNA into nucleosomal fragments of ~180 bp.

In recent animal studies, dopamine receptor agonists such as bromocriptine and pergolide, which are used for symptomatic therapy of Parkinson’s disease, showed possible neuroprotective effects under a variety of neurodegenerative conditions (Lange et al., 1994; Liu et al. 1995). Great interest, therefore, is focused on the neuroprotective effects of dopamine receptor agonists as candidates for the current and future treatment of Parkinson’s disease. The human neuroblastoma cell line SH-SY5Y, which was subcloned from the SK-N-SH cell line, often is used as a model of human dopaminergic neurons (Itano and Nomura, 1995; Ross and Biedler, 1985). The azepine

ABBREVIATIONS: MPTP, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; diBu-cAMP, N6,2’-O-dibutyryl cAMP; CDDHF-DA, 6-carboxy-2’-7’- dichlorodihydrofluorescein diacetate di(acetoxyethyl) ester; MPP+, 1-methyl-4-phenylpyridinium; MTT, 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide; PARP, poly(ADP-ribose) polymerase; PKA, protein kinase A; PKC, protein kinase C; PMA, phorbol-12-myristate-13-acetate; ROS, reactive oxygen species; ANOVA, analysis of variance.

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derivatives talipexole (Arnt et al., 1986) and pramipexole (Mierau et al., 1995) are agonists for D2/D3 receptors and D2/D3 receptors, respectively, both lacking the ability to stimulate the D1 receptor. In the current study, we examined the protective effects of talipexole and pramipexole, which are novel drugs for the treatment of Parkinson's disease, along with bromocriptine, which is the most widely used ergot-derivative dopamine receptor agonist, against MPP⁺-induced apoptotic death in human neuroblastoma SH-SY5Y cells.

**Experimental Procedures**

**Materials.** Talipexole (B-HT 920CL₂; 6-allyl-2-amino-5,6,7,8-tetrahydro-4H-thiazolo[4,5-d]azepine dihydrochloride) and pramipexole (SND 919CL₁,₂; (-)-2-amino-4,5,6,7-tetrahydro-6-(propylamino)benzothiazole dihydrochloride) were obtained from Boehringer-Ingelheim (Ingelheim, Germany). Bromocriptine mesilate was kindly donated by Sandoz Pharma A.G. (Basel, Switzerland). MPP⁺ and dopamine receptor antagonists were from Research Biochemicals International (Natick, MA). diBu-cAMP was from Wako (Osaka, Japan). Calphostin C was from Kyowa Hakko Kogyo (Tokyo, Japan). PMA was from Sigma Chemical (St. Louis, MO). MTT was from Dojindo Laboratories (Kumamoto, Japan). Hoechst 33258 bis-benzimide (H-1398) and C-DC-DHF-DA (C-2938) were from Molecular Probes (Eugene, OR).

Primary antibodies included mouse monoclonal antibodies to human Bcl-2 (clone 124, M887) from DAKO (Copenhagen, Denmark), human Bak (clone TC100, AM03) from Oncogene Research Products (Cambridge, MA), mouse Bad (clone 48, B36420) and human caspase-3 (clone 19, C31720) from Transduction Laboratories (Lexington, KY), human p53 (DO-1, sc-126) from Santa Cruz Biotechnology (Santa Cruz, CA), rabbit polyclonal antibodies to human Bel-x (B22630) from Transduction Laboratories (Lexington, KY), human p53 (DO-1, sc-126) from Santa Cruz Biotechnology, proenzyme/active fragments of human caspase-3 (65906E) from PharMingen (San Diego, CA), and the caspase-3 cleavage site of human PARP (06–557) from Upstate Biotechnology (Lake Placid, NY). An enhanced chemiluminescent detection system (ECL kit) from Amersham (Buckinghamshire, England) was used for immunodetection.

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**Fig. 1.** Effects of simultaneous treatment and pretreatment with talipexole on MPP⁺-induced cell death (A) and DNA fragmentation (B). SH-SY5Y cells were treated without (100% activity, lane 1) or with 1 mM MPP⁺ for 3 days in the absence (black column, lane 2) or presence (hatched column, lane 3) of talipexole. Alternatively, cells were pretreated with 1 mM talipexole for 1–4 days before treatment for 3 days with 1 mM MPP⁺ in the absence of talipexole (B, lanes 4–7, respectively).

**Fig. 2.** Protective effects of pretreatment with talipexole, pramipexole, and bromocriptine on MPP⁺-induced cell death and DNA fragmentation. SH-SY5Y cells were pretreated for 4 days with vehicle (black column), talipexole (at 0.3, 1, or 3 mM; open column), pramipexole (at 0.3, 1, or 3 mM; gray column), or bromocriptine (at 0.01 or 0.03 mM; hatched column) and then further treated with 1 mM MPP⁺ for 3 days. A, Cell survival was measured by MTT assay. Each value is the mean ± standard error (%) of six determinations, based on the vehicle control as 100%. The F value in ANOVA was F(5,24) = 129.589 (p < 0.001). In addition, the Bonferroni/Dunn test was performed for post hoc comparisons. **, p < 0.01; ***, p < 0.001 versus no pretreatment with talipexole (–) (Bonferroni/Dunn test). B, DNA fragmentation in ~2 × 10⁶ cells was assessed. M, DNA size markers. Simultaneous treatment with talipexole afforded slight but significant protection against cell death (A, hatched column) and inhibition of DNA fragmentation (B, lane 3). Pretreatment with talipexole markedly inhibited MPP⁺-induced cell death and DNA fragmentation.
Cell culture and treatment with drugs. The human neuroblastoma cell line SH-SY5Y (Ross and Biedler, 1985) was used in the current experiments. The cells were cultured in Dulbecco's modified Eagle's medium supplemented with 10% (v/v) fetal calf serum, 50 μg/ml penicillin, and 100 μg/ml streptomycin and were kept at 37° in humidified 5% CO₂/95% air. SH-SY5Y cells were treated with drugs as follows. (1) For simultaneous treatment, SH-SY5Y cells were simultaneously treated for 24 hr or 3 days with 1 mM MPP⁺ and 1 mM talipexole. (2) For pretreatment, the cells were pretreated for 1–4 days with vehicle, talipexole, pramipexole, bromocriptine, diBu-cAMP, calphostin C, or PMA in the absence or presence of a dopaminergic antagonist. The cells then were washed with fetal calf serum-free Dulbecco's modified Eagle's medium and further treated with 1 mM MPP⁺ for 3 days in the absence of drugs. Treated cells were subjected to cell survival assay, DNA fragmentation analysis, immunoblotting, and fluorescence measurement.

MTT assay. MTT is converted in living cells to formazan, which has a specific absorption maximum. After pre- and post-treatments of SH-SY5Y cells as described, the culture medium was changed to the medium containing 5 mg/ml MTT, and the cells were incubated further for 4 hr. Then, they were mixed thoroughly with an equal volume of isopropanol/0.04 M HCl. After centrifugation at 10,000 g for 20 min, the absorbance of the supernatant was measured at 570 nm.

Assay of DNA fragmentation. After treatment of −2 × 10⁶ cells with vehicle or MPP⁺ in 60-mm-diameter dishes, the cells were scraped from the dishes using a rubber policeman, centrifuged at 800 × g for 10 min, and resuspended in 100 μl of lysis buffer of 10 mM Tris-HCl, pH 7.4, containing 10 mM EDTA and 0.5% Triton X-100. After incubation for 10 min at 4°, lysates were centrifuged at 15,000 × g for 30 min. The supernatants thus obtained were supplemented with 2 μl of RNase A (20 mg/ml) and then incubated at 37° for 1 hr. Next, 2 μl of proteinase K (20 mg/ml) was added, and incubation was continued at 37° for 1 hr. After these incubations, 20 μl of 5 mM NaCl and 120 μl of isopropanol were added, and the mixture was held overnight at −20°. After centrifugation at 15,000 × g for 20 min, DNA pellets were resuspended in 10 mM Tris-HCl, pH 7.4, containing 1 mM EDTA. DNA fragments thus obtained were electrophoretically separated on a 2% agarose gel for 90 min at 50 V. The gel was stained with ethidium bromide and photographed under UV transillumination. DNA size markers (pX174 RF DNA/HaeIII fragments) are 1353, 1078, 872, 603, 310, 271/281, 234, 194, 118, and 72 bp.

Hoechst 33258 staining. Chromosomal condensation and DNA fragmentation were determined using the chromatin dye Hoechst 33258. After pre- and post-treatments of SH-SY5Y cells, the cells were harvested and fixed with 4% paraformaldehyde in PBS for 30 min at 4°. After three rinses with the same buffer, cells were stained with 1 μM Hoechst 33258 for 5 min and analyzed under a fluorescence microscope (Axioplan; Carl Zeiss, Jena, Germany) with excitation at 352 nm. Fluorescence micrographs were scanned with a high resolution camera (ProRes 3008, Carl Zeiss).

C-DCHDF-DA staining. To detect ROS production, we used the redox-sensitive dye C-DCHDF-DA, which is readily taken up by cells. After simultaneous treatment or pretreatment of SH-SY5Y cells in uncoated glass-bottomed microwells (inner diameter, 18 mm; MatTek, Asland, MA), C-DCHDF-DA was added to the cell culture to a final concentration at 2 μM for 10 min at 37°. After two rinses with serum-free medium, samples were scanned under a confocal microscope (LSM410, Carl Zeiss). The optimal vertical position at the middle of the cells was set, and then the field was rapidly scanned. Because illumination at the excitation wavelength of 488 nm caused increased fluorescence because of oxidation of this dye (Greenlund et al., 1995), each field was exposed to light for exactly the same time, and the laser intensity was set at only 3% of maximum. After scanning (excitation, 488 nm; emission, over 515 nm), the average relative fluorescence intensity for every cell in each field was evaluated. Fluorescence micrographs were printed with a full-color digital photo printer (Pictography 3000; Fuji Film, Tokyo, Japan).

**Fig. 3.** Protective effects of pretreatment with talipexole, pramipexole, and bromocriptine against MPP⁺-induced chromatin destruction. SH-SY5Y cells were pretreated for 4 days with vehicle (A and B), 1 mM talipexole (C), 1 mM pramipexole (D), or 0.03 mM bromocriptine (E) and then further treated with vehicle (A) or 1 mM MPP⁺ (B–E) for 3 days. Nuclei were stained with Hoechst 33258. Numerous cells treated with MPP⁺ after vehicle pretreatment showed apoptotic features (chromatin condensation and DNA fragmentation) (B). Many nuclei appeared normal in cells pretreated with talipexole (C), pramipexole (D), and bromocriptine (E). Bar (in A) = 20 μm.

**Fig. 4.** Lack of effect of dopaminergic antagonists on talipexole pretreatment-induced protection. SH-SY5Y cells were pretreated for 4 days with 0.5 mM talipexole in the presence of 0.1 mM SCH23390 (SCH), 0.3 mM sulpiride (Sul), 0.01 mM spiperone (Spi), 0.01 mM domperidone (Dom), or 0.01 mM clozapine (Clo) and then further treated with 1 mM MPP⁺ for 3 days. Subsequently, cell survival was measured by MTT assay. Each value is the mean ± standard error (%) of four determinations, based on the vehicle control as 100%. The F value in ANOVA was F(6,21) = 137.141 (p < 0.0001). When the Bonferroni/Dunn test was performed for post hoc comparisons, dopaminergic antagonists had no significant effect.
Immunoblotting assay for Bcl-2 family members, p53, caspase-3, and PARP. Cell lysates were dissolved in Laemmli’s sample buffer and subjected to sodium dodecyl sulfate-polyacrylamide gel electrophoresis (15%, 10%, or 8% polyacrylamide gels). Immunoblotting was performed by transferring proteins from a slab gel to a sheet of polyvinylidene difluoride membrane (BioRad Laboratories, Richmond, CA) by electroelution at a constant voltage of 50 V for 2 hr at 4°. The PVDF membrane was incubated with Tris-buffered saline, pH 8.0, containing 0.3% Triton X-100 (TBS-T) and 5% dehydrated skim milk (Difco Laboratories, Detroit, MI) to block nonspecific protein binding. The membrane then was incubated with primary antibodies, including mouse monoclonal antibodies to Bcl-2 (diluted 1:300), Bak (1:400), Bad (1:300), p53 (1:2000), and pro-caspase-3 (1:1000) or rabbit polyclonal antibodies to Bcl-x (1:1000), Bax (1:300), active caspase-3 (1:1000), and PARP (1:600), followed by horseradish peroxidase-linked antibodies against either rabbit or mouse immunoglobulins (each diluted 1:100) as secondary antibodies. Bound HRP-labeled antibodies were detected by chemiluminescence assay (ECL kit, Amersham). The protein bands that reacted with the antibodies were detected on radiographic film (X-Omat JB-1; Kodak, Rochester, NY) after exposure for 5–60 sec. The bands of Bcl-2, Bcl-x, p53, and caspase-3 on radiographic films were scanned and densitometrically analyzed by a dual-wavelength flying-spot scanner (CS-9000; Shimadzu, Kyoto, Japan). Pre-stained sodium dodecyl sulfate-polyacrylamide gel electrophoresis standards (BioRad) were used as molecular mass markers. Apparent molecular masses of phosphorylase B, bovine serum albumin, ovalbumin, carbonic anhydrase, soybean trypsin inhibitor, and lysozyme were 112, 84, 53.2, 34.9, 28.7, and 20.5 kDa, respectively, according to the manufacturer’s labeling.

Statistical evaluation. Results in MTT assay and densitometric analysis of the immunoblots are given as mean ± standard error values. Statistical significance of differences was determined by mean values of the ANOVA. Further statistical analysis for post hoc comparisons was done by using the Bonferroni/Dunn test.

Results

MPP⁺-induced apoptotic death in SH-SY5Y cells. Treatment with MPP⁺ caused a time- and concentration-dependent increase in cell death, and delayed DNA fragmentation was observed (data not shown). In particular, SH-SY5Y cells treated with 1 mM MPP⁺ for >3 days exhibited marked cell death and DNA fragmentation (Fig. 1). To confirm the morphological change, we further examined cells under a fluorescence microscope using a chromatin dye, Hoechst 33258. At 3 days after MPP⁺ treatment, the cell body and nucleus were shrunk and compacted, with condensed chromatin and fragmented nuclei (Fig. 3B). In addition, MPP⁺ induced cleavages of 32-kDa caspase-3 and 116-kDa PARP to 20/17-kDa active fragments and an 85-kDa fragment, respectively (Fig. 10). These morphological and biochemical changes are characteristic of apoptosis.

Protective effects of talipexole and pramipexole against MPP⁺-induced apoptotic death of SH-SY5Y cells. Preliminary treatment of SH-SY5Y cells for 4 days with talipexole or pramipexole at 0.01–1 mM did not markedly alter cell growth, but bromocriptine treatment at >0.1 mM decreased the cell viability (data not shown). Bromocriptine has ~10-fold higher affinity for the D2-receptor than...
talipexole and pramipexole, but the agonistic potency of bromocriptine is lower than that of talipexole. Therefore, we used talipexole or pramipexole at 0.01–3 mM and bromocriptine at 0.01–0.03 mM.

Simultaneous treatment with 1 mM MPP⁺ and 1 mM talipexole for 3 days (without talipexole pretreatment) resulted in slight but significant protection against cell death and DNA fragmentation compared with 1 mM MPP⁺ alone (Fig. 1). In addition, talipexole pretreatment at the same concentration inhibited MPP⁺-induced cell death and DNA fragmentation in a pretreatment time- and concentration-dependent manner (Figs. 1 and 2). Pretreatment with other D₂ receptor agonists, pramipexole and bromocriptine, also inhibited MPP⁺-induced cell death and DNA fragmentation (Fig. 2). Although MPP⁺ treatment alone induced many apoptotic features such as condensed chromatin and fragmented nuclei (Fig. 3B), pretreatment with talipexole, pramipexole, and bromocriptine markedly decreased these features (Fig. 3, C–E, respectively).

Because the D₂ receptor is expressed in SH-SY5Y cells (Farooqui, 1994), we examined whether the protective effect of talipexole is mediated by dopaminergic receptors. However, various selective or nonselective antagonists for dopaminergic receptor subtypes, SCH23390 (D₁/D₅), sulpiride (D₂/D₃), spiperone (D₂/D₄), domperidone (D₂/D₃/D₄), and clozapine (D₄), did not inhibit the talipexole-induced protection (Fig. 4).

**Effects of talipexole and pramipexole on MPP⁺-induced ROS production in SH-SY5Y cells.** We further performed confocal analysis with a ROS-specific fluorogen, C-DCDHF-DA, to examine whether MPP⁺ induces ROS production and, if so, what is the effect of talipexole. The fluorescence intensity of SH-SY5Y cells showed an early increase when the cells were treated for 30 min with H₂O₂ as a positive control (Fig. 5D). In contrast, a delayed increase of the intensity was observed after 12 and 24 hr in the cells treated with 1 mM MPP⁺ (Fig. 5, B and C, respectively).

Compared with the fluorescence intensity 24 hr after treatment with 1 mM MPP⁺ alone, that with simultaneous treatment with 1 mM talipexole (Fig. 5E) was much lower, being close to the basal level (Fig. 5A). Similarly, simultaneous treatment with pramipexole or bromocriptine inhibited MPP⁺-induced fluorescence (ROS production; data not shown). Pretreatment with 1 mM talipexole for 4 days also markedly inhibited MPP⁺-induced ROS production (Fig. 5F).

**Effects of talipexole and pramipexole on protein levels of Bcl-2 family members in SH-SY5Y cells.** Bcl-2 family members regulate apoptotic cell death, and SH-SY5Y cells natively express antiapoptotic members such as Bcl-2 and Bcl-x (Dole et al., 1995; Reed et al., 1991) and proapoptotic members such as Bax, Bak, and Bad (Kitamura et al., 1998b). Therefore, we examined whether the protein levels of Bcl-2 family members are influenced by treatment with talipexole, pramipexole, and bromocriptine. For this purpose, we used each specific antibody for immunoblotting of 27-kDa Bcl-2, 20-kDa Bcl-xL, 21-kDa Bax, 30-kDa Bak, or 23-kDa Bad (Figs. 6 and 8). Treatment with talipexole or pramipexole for 4 days induced significant enhancement of the antiapoptotic Bcl-2a protein level but did not change the Bcl-xL level (Figs. 6C and 7A, respectively), whereas treatment with bromocriptine for 4 days did not markedly change the levels of either antiapoptotic protein (Fig. 7B). On the other hand, protein levels of proapoptotic members such as Bax, Bak, and Bad in SH-SY5Y cells treated with talipexole, pramipexole or bromocriptine. SH-SY5Y cells were treated for 4 days with vehicle (lane 1), 1 mM talipexole (lane 2), 1 mM pramipexole (lane 3), and 0.03 mM bromocriptine (lane 4). Samples (10 μg protein/lane) were subjected to immunoblotting with antibodies against Bax, Bak, and Bad. These proapoptotic protein levels were not markedly changed in cells treated with talipexole, pramipexole, or bromocriptine.

![Fig. 7](image7.png)  
**Fig. 7.** Changes in protein levels of antiapoptotic Bcl-2a and Bcl-x induced by pretreatment with pramipexole and bromocriptine in SH-SY5Y cells. SH-SY5Y cells were treated for 4 days with vehicle, pramipexole (A), or bromocriptine (B). Samples (10 μg protein/lane) were subjected to immunoblotting with antibodies against Bcl-2a and Bcl-x. Protein levels of Bcl-2a (open column) and Bcl-xL (hatched column) were measured by densitometric analysis of the immunoblots. Each value is the mean ± standard error (%) of three determinations, based on the density of the band in the vehicle control as 100%. In pramipexole treatment, F values for protein levels of Bcl-2a and Bcl-xL in ANOVA were F(3,8) = 135.326 (p < 0.0001) and 1.035 (p = 0.4275), respectively. In bromocriptine treatment, F values for protein levels of Bcl-2a and Bcl-xL in ANOVA were F(3,8) = 1.763 (p = 0.2318) and 1.450 (p = 0.2992), respectively. In addition, the Bonferroni/Dunn test was performed for post hoc comparisons. ***, p < 0.001 versus the level in the corresponding vehicle control (Bonferroni/Dunn test).

![Fig. 8](image8.png)  
**Fig. 8.** No change in protein levels of proapoptotic Bax, Bak, and Bad in SH-SY5Y cells treated with talipexole, pramipexole or bromocriptine. SH-SY5Y cells were treated for 4 days with vehicle (lane 1), 1 mM talipexole (lane 2), 1 mM pramipexole (lane 3), and 0.03 mM bromocriptine (lane 4). Samples (10 μg protein/lane) were subjected to immunoblotting with antibodies against Bax, Bak, and Bad. These proapoptotic protein levels were not markedly changed in cells treated with talipexole, pramipexole, or bromocriptine.
Bax, Bak, and Bad were not markedly changed by treatment for 4 days with talipexole, pramipexole, or bromocriptine (Fig. 8). Although treatment for 3 days with MPP⁺ alone did not significantly change the Bel-2-a level, the protein level was significantly increased even 3 days after MPP⁺ treatment by pretreatment with talipexole for 4 days (Fig. 9).

Inhibitory effects of talipexole against MPP⁺-induced p53 expression and cleavages of caspase-3 and PARP in SH-SY5Y cells. p53 protein and caspase-3 (CPP32) act as a transcription factor and a key enzyme for apoptosis, respectively (Ko and Prives, 1996; Nicholson et al., 1995; Tewari et al., 1995). In the case of caspase-3, the 32-kDa proenzyme is cleaved by other proteases to generate the active form, which is composed of 12-kDa and active 20/17-kDa fragments. Subsequently, 116-kDa PARP is cleaved by active caspase-3 in cells undergoing apoptosis (Nicholson et al., 1995; Tewari et al., 1995). Therefore, we further examined p53 expression and cleavages of caspase-3 and PARP.

Although treatment with MPP⁺ for 3 days markedly induced expression of p53 protein, pretreatment with talipexole for 4 days inhibited the MPP⁺-induced p53 expression (Fig. 9). Using mouse monoclonal anti-caspase-3 antibody (C31729; Transduction Laboratories), the 32-kDa proenzyme of caspase-3 was decreased by MPP⁺ treatment, but 20/17-kDa active fragments were undetectable. Talipexole pretreatment inhibited the MPP⁺-induced degradation of pro-caspase-3, and the level of the protein was similar to that in the control (Fig. 9). Using rabbit polyclonal anti-caspase-3 antibody (65906E; PharMingen), 20/17-kDa active fragments cleaved from 32-kDa proenzyme were detected. MPP⁺-induced cleavage of caspase-3 was inhibited by pretreatment with talipexole or pramipexole (Fig. 10A). In addition, MPP⁺-induced PARP cleavage to an 85-kDa fragment also was detected using polyclonal antibody against the caspase-3 cleavage site of human PARP (06–557; Upstate Biotechnology). This PARP cleavage also was inhibited by pretreatment with talipexole or pramipexole (Fig. 10B).

Decrease of Bcl-2 level and enhancement of MPP⁺-induced apoptosis by diBu-cAMP or calphostin C. We previously found that activation of cAMP-dependent PKA or inhibition of PKC induced a decrease in Bcl-2 level and then enhanced NO-induced apoptosis (Kitamura et al., 1989a). In the current study, treatment for 4 days with 1 mM diBu-cAMP or 30 nM calphostin C induced a decrease in antiapoptotic Bcl-2 and Bcl-x but did not markedly change the levels of proapoptotic Bax, Bak, and Bad (Fig. 11A). These pretreatments enhanced MPP⁺-induced cell death (Fig. 11B) and cleavages of caspase-3 and PARP (data not shown). On the other hand, treatment with 100 nM PMA induced an increase in Bcl-2 and Bcl-x but did not change Bax, Bak, and Bad (Fig. 11A). In addition, MPP⁺-induced cell death (Fig. 11B) and cleavages of caspase-3 and PARP were inhibited by PMA pretreatment (data not shown).

Discussion

The brains of patients with Parkinson’s disease show evidence of inhibition of mitochondrial complex I (Schapira et al., 1989), generation of oxidative stress (Castellani et al., 1996), and induction of apoptosis (Mochizuki et al., 1996). Similar phenomena are observed in MPTP-treated animal...
models (Tatton and Kish, 1997; Sriram et al., 1997). It is known that levodopa and dopamine exhibit neurotoxicity (Ogawa, 1994), whereas D2 receptor agonists such as bromocriptine have neuroprotective effects against MPTP-induced and ischemic insults (Lange et al., 1994; Liu et al., 1995). We recently found that the administration of talipexole or pramipexole inhibited MPTP-induced degeneration of dopaminergic neurons in C57BL/6 mouse, but talipexole did not inhibit the uptake of MPP⁺ into striatal synaptosomes (Kitamura et al., 1997). Thus, the mechanisms of these neuroprotective effects remain unclear. In the current study, we therefore focused on intracellular events related to MPP⁺ cytotoxicity, and we examined the protective effects of talipexole and pramipexole on the MPP⁺-induced changes.

In human neuroblastoma SH-SY5Y cells, MPP⁺ treatment induced delayed ROS production in comparison with the immediate production induced by H₂O₂ (Fig. 5). This was followed by apoptotic cell death with the characteristic DNA fragmentation and morphological changes (condensed chromatin and fragmented nuclei). It is known that MPP⁺ is concentrated in mitochondria (Ramsay and Singer, 1986; Mizuno et al., 1995), where it inhibits mitochondrial complex I (Mizuno et al., 1995; Nicklas et al., 1987) and that mitochondrial dysfunction causes apoptosis (Kroemer et al., 1997). Therefore, delayed ROS production by MPP⁺ in SH-SY5Y cells may be mediated through inhibition of mitochondrial complex I, leading to apoptotic death.

Because simultaneous treatment with talipexole inhibited MPP⁺-induced ROS production (Fig. 5E) and slightly inhibited MPP⁺-induced apoptosis (Fig. 1), talipexole may have direct scavenging effects against ROS. On the other hand, pretreatment with talipexole markedly protected the cells against MPP⁺-induced apoptosis in a treatment time- and concentration-dependent manner (Figs. 1 and 2), and this protective effect might not be mediated by dopamine receptors (Fig. 4). This pretreatment also inhibited H₂O₂-induced DNA fragmentation (data not shown). Furthermore, talipexole treatment induced an increase in antiapoptotic Bcl-2 protein level but did not change the Bcl-x level (Fig. 6), whereas protein levels of proapoptotic Bax, Bak, and Bad were not markedly changed by talipexole treatment (Fig. 8). This pretreatment also inhibited MPP⁺-induced ROS production (Fig. 5F). In this case, MPP⁺ treatment was conducted after talipexole had been washed out, so the protection was not mediated by direct ROS scavenging but probably via another pathway, such as induction of Bcl-2 expression. Pramipexole pretreatment also enhanced the Bcl-2 level (Fig. 7A) and inhibited MPP⁺-induced apoptosis (Fig. 2). Furthermore, talipexole pretreatment inhibited the MPP⁺-induced p53 expression and decrease in a 32-kDa proenzyme of caspase-3 (Fig. 9). Induction of p53 protein is usually considered one of the primary events in the apoptotic cascade after DNA damage (Ko and Prides, 1996). It is also known that 32-kDa caspase-3 is activated through cleavage to 12-kDa and active 20/17-kDa fragments by apoptotic signals, and subsequently, PARP is cleaved to the 85-kDa fragment (Nicholson et al., 1995; Tewari et al., 1995). Talipexole pretreatment seems to inhibit events leading to apoptosis, such as p53 expression and cleavages of caspase-3 and PARP (Fig. 10). Pramipexole also has similar effects.

PKA activator (diBu-cAMP) or PKC inhibitor (calphostin C) induced a decrease in protein levels of antiapoptotic Bcl-2 and Bcl-x but did not change the levels of proapoptotic Bax, Bak, and Bad (Fig. 11). With this pretreatment, MPP⁺-induced apoptotic changes (cleavages of caspase-3 and PARP) and cell death were enhanced. Recent studies suggest that Bcl-2 protein inhibits ROS production and apoptosis (Hockenbery et al., 1993; Kane et al., 1993) and that Bcl-2 inhibits the loss of mitochondrial membrane potential and activation of caspase-3 (Shimizu et al., 1996). In view of these observations, we consider that (1) talipexole (or pramipexole) has a direct scavenging effect on ROS but only a small protective effect and (2) treatment with talipexole (or pramipexole) induces an increase in Bcl-2, which affords more potent protection against MPP⁺-induced apoptosis. In addition, PKA inhibition or PKC activation may participate with neuroprotection by talipexole (or pramipexole). However, because dopamine receptor antagonists did not inhibit talipexole-induced protection, the detail mechanisms of this neuroprotection are still unclear. Talipexole (or pramipexole) may affect other receptors regulating PKA, PKC, or both, which are not still identified as these binding receptors.
whereas talipexole (or pramipexole) may be uptaken into neuronal cells through monoaamine transporters (dopamine, noradrenergic, and so on) or other mechanisms, and then these drugs directly or indirectly may regulate intracellular events to Bcl-2 expression. Further studies of neuroprotective mechanisms of talipexole (or pramipexole) in detail are necessary before definite conclusions can be drawn.

The Bcl-2 protein level is known to be up-regulated in brains of patients with Parkinson’s disease (Mogi et al., 1996), as well as those with Alzheimer’s disease (Kitamura et al., 1998b). Such up-regulation of Bcl-2 protein may represent a compensatory response of remaining neurons to protect them from oxidative stress and subsequent apoptosis. Therefore, neurons with lower Bcl-2 expression may be more vulnerable to apoptotic death. Because an increase of Bcl-2 was induced by treatment for ≥4 days with talipexole and pramipexole (but not bromocriptine), these drugs may be useful for long term treatment of patients with Parkinson’s disease. On the other hand, pretreatment with bromocriptine also inhibited MPP⁺-induced apoptotic cell death (Fig. 2), although it had no effect on the protein levels of Bcl-2 family members (Figs. 7B and 8). Recent studies have suggested that bromocriptine directly scavenges superoxide (Yoshikawa et al., 1994) and NO radical (Nishibayashi et al., 1997), inhibits lipid peroxidation (Tanaka et al., 1995), inhibits glutamate uptake into synaptic vesicles (Carlson et al., 1989), and enhances glutamate uptake through human glutamate transporter hGLuT-1 (Yamashita et al., 1995). These effects may contribute to the neuroprotective action of bromocriptine in contrast to talipexole and pramipexole.

In conclusion, MPP⁺ treatment induced delayed ROS production in SH-SY5Y cells in comparison with the immediate production induced by H₂O₂. This was followed by apoptotic cell death. Simultaneous treatment with talipexole markedly inhibited MPP⁺-induced ROS production and slightly inhibited MPP⁺-induced apoptosis. On the other hand, pretreatment with talipexole markedly protected the cells against MPP⁺-induced apoptosis, and this protective effect might not be mediated by dopamine receptors. This pretreatment induced an increase in antiapoptotic Bcl-2 level and inhibited MPP⁺-induced ROS production. Pramipexole pretreatment exhibited similar effects. Although pretreatment with bromocriptine also had a protective effect, there was no change in the level of Bcl-2 family members. On the other hand, pretreatment with diBu-AMP or calphostin C, which caused a decrease in Bcl-2 level, enhanced MPP⁺-induced apoptosis. These results suggest that talipexole directly scavenges ROS, but this effect offers only slight protection; however, pre-treatment markedly increases Bcl-2, which results in more potent protection against MPP⁺-induced apoptosis. Pramipexole has similar effects, but bromocriptine may lack the ability to induce Bcl-2 expression.

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