Transcriptional Induction of Heme Oxygenase-1 Gene Expression by Okadaic Acid in Primary Rat Hepatocyte Cultures

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Received May 13, 1999; accepted November 19, 1999

ABSTRACT

Heme oxygenase (HO) catalyzes the rate-limiting enzymatic step of heme degradation and regulates the cellular heme content. The gene expression of the inducible isozyme of HO, HO-1, is up-regulated in response to various agents causing oxidative stress. To investigate the regulatory role of protein phosphatases in the hepatic regulation of HO-1 gene expression, primary cultures of rat hepatocytes were treated with okadaic acid (OA), which specifically inhibits the serine threonine protein phosphatases 1 and 2A. Both protein synthesis and mRNA expression of HO-1 were induced by OA in cultured hepatocytes, but not in cultured tissue macrophages of rat liver. The HO-1 mRNA induction by OA occurred in a time- and concentration-dependent manner. Simultaneous treatment with OA plus dibutyryl cAMP caused a synergistic up-regulation of steady-state levels of HO-1 mRNA, and the specific protein kinase A inhibitor KT5720 markedly reduced the OA-dependent HO-1 mRNA induction. In contrast, the dibutyryl cAMP-dependent induction of the phosphoenolpyruvate carboxykinase mRNA expression and enzyme activity was inhibited by simultaneous treatment with OA in hepatocytes. The induction of the HO-1 gene expression by OA was transcriptional as determined by studies with actinomycin D, nuclear run-off assay, and measurement of the half-life of HO-1 mRNA. Luciferase reporter constructs containing DNA sequences of the rat HO-1 promoter 5′-flanking region were up-regulated by OA in transiently transfected hepatocytes. Mutation of the cAMP response element/activator protein-1 (−665/−654) site obliterated the OA-dependent induction, suggesting that this element is involved in the transcriptional induction of the rat HO-1 gene by OA.

Heme oxygenase (HO) catalyzes the first and rate-limiting step of heme degradation and controls the cellular heme availability (Tenhunen et al., 1968). HO enzymatically breaks down the pro-oxidant heme, producing equimolar amounts of carbon monoxide, iron, and biliverdin, which are converted by biliverdin reductase into the antioxidant bilirubin (Stocker et al., 1987). At least two distinct isoforms of HO are known that are the products of different genes. In contrast to the constitutive isozyme HO-2 (Maines et al., 1986), HO-1 is the inducible isozyme, which is highly up-regulated by various stress stimuli including its substrate heme, heavy metals, UV light, lipopolysaccharide, heat shock, and hyperoxia (Shibahara et al., 1987; Applegate et al., 1991; for reviews see Maines, 1988, and Choi and Alam, 1996). Although the exact functional role of HO-1 induction is not fully understood, various researchers have shown that HO-1 provides protection against oxidative stress in various cell culture and in in vivo models (Abraham et al., 1995; Lee et al., 1996). Overexpression of the HO-1 gene attenuates the toxic effects of heme proteins in coronary endothelial cells (Abraham et al., 1995) and protects pulmonary epithelial cells against hyperoxia (Lee et al., 1996). Poss and Tonegawa (1997a) have shown that HO-1-deficient mice develop an anemia with abnormally low serum iron levels, along with an overload of iron in liver and kidney, causing oxidative damage and chronic inflammation. In addition, HO-1-deficient mice were highly susceptible to endotoxin-mediated hepatic damage, resulting in a higher mortality rate from endotoxic shock in these animals (Poss and Tonegawa, 1997b).

OA is a polyether fatty acid isolated from marine sponges.
that initially has been shown to be a tumor promoter (Holmes and Boland, 1993). Instead of activating protein kinase (PK) C as do phorbol ester tumor promoters, OA is a specific inhibitor of protein phosphatase (PP)1 and PP2A (Holmes and Boland, 1993). PP1 and PP2A dephosphorylate serine and threonine residues in cellular target proteins that are involved in the regulation of multiple signaling pathways (for review, see Wera and Hemmings, 1995). Induction of HO-1 gene expression by activation of PKC (Muraoasa and Shiba-
hará, 1993), cAMP-dependent PK (PKA) (Durante et al., 1997; Immenschuh et al., 1998b), or cGMP-dependent PK (PKG) (Immenschuh et al., 1998a) has been demonstrated previously; however, little is known about the role of PPs in the gene regulation of HO-1. Because it has become increasingly obvious that PPs play a major role in maintaining the intracellular balance of gene expression (Hunter, 1995), we investigated the effects of OA on the expression of the HO-1 gene in cultures of primary rat hepatocytes.

In this study, it is shown that OA induces HO-1 gene expression on the protein and mRNA level in a time- and dose-dependent manner. This induction of HO-1 by OA is regulated on the transcriptional level and appears to be mediated by the cAMP response element (CRE)/AP-1 site of the rat HO-1 gene promotor 5'—flanking region.

**Experimental Procedures**

**Animals.** Male Wistar rats (2 months old, body weight 170–200 g) were used throughout the study.

**Materials.** Media M199 and RPMI were obtained from Life Technologies (Karlsruhe, Germany), nitrocellulose filters were from Schleicher and Schuell (Dassel, Germany), and radioisotopes and the chemiluminescent detection system for Western blotting were from Amershamsch- Buchler (Braunschweig, Germany). The multiprime labeling kit and restriction endonucleases were from New England Biolabs (Cambridge, MA). Falcon tissue culture dishes were from Becton Dickinson (Heidelberg, Germany). OA, calcyquin A, and KT5720 were from Calbiochem (San Diego, CA). The polyclonal rabbit anti-rat HO-1 antibody was obtained from Stress Gene (Victoria, Canada). All other chemicals were obtained from Sigma (Deisenhofen, Germany) and Boehhringer Mannheim (Mannheim, Germany) unless indicated otherwise.

**Cell Isolation and Culture.** Hepatocytes were isolated from male Wistar rats by circulating perfusion with collagenase under sterile conditions as described previously (Immenschuh et al., 1998b). The cells were cultured under air/CO₂ (19/1) in medium 199 with Earle's salts containing 2 g/l BSA, 20 mM NaHCO₃, 10 mM HEPES, 117 mg/l streptomycin sulfate, 60 mg/l penicillin, 1 mM insulin, and 10 nM dexamethasone. Fetal calf serum (5%) was added to the culture medium. Hepa 1–6 and NIH3T3 cells were from the American Type Culture Collection (Manassas, VA). Hepa 1–6 cells were cultured in RPMI 1640 medium containing 2% fetal calf serum, and NIH3T3 cells were cultured in DMEM with 10% fetal calf serum until confluency of cell monolayers was reached. Confluent monolayers were incubated in serum-free medium 18 h before treatment.

Tissue macrophages of rat liver (Kupffer cells) were isolated as described (Immenschuh et al., 1999). In brief, the liver was digested with pronase/collagenase solutions, and nonparenchymal cells were separated by density gradient centrifugation. Kupffer cells were purified by counterflow elutriation (J2–21, JE-B6 rotor; Beckmann Instruments, Fullerton, CA), and the obtained Kupffer cells were resuspended in M199 containing 15% fetal calf serum, 100 U of penicillin/ml, and 100 µg of streptomycin/ml. Cell viability was as-

sessed by trypan blue staining, and cells were plated on six-well plates (3 × 10⁶ cells/well). After 2 h, cells were washed for elimination of nonadherent cells, and cell culture was continued with serum-
free medium.

**Bioisynthetic Labeling, Immunoprecipitation, and SDS-Polyacrylamide Gel Electrophoresis (PAGE) of Synthesized Proteins.** Hepatocytes were washed with methionine-free M199 and were pulsed for 2 h with M199 containing [³⁵S]methionine (600 µCi/ml). Cell layers were washed with ice-cold PBS, covered with lysis buffer (PBS, 0.5%; deoxycholic acid, 1%; SDS, 7.4%) containing 1 mM phenylmethylsulfonyl fluoride (PMSF) and were frozen at −70°C. After two freezing/thawing cycles in lysis buffer, lysates were centrifuged (10,000 g, 30 min, 4°C) and diluted with lysis buffer (1:1). For immunoprecipitation, samples adjusted to contain equal amounts of radioactivity, as determined by trichloroacetic acid precipitation and β-ray counting, were incubated overnight with an excess of antisera at 4°C. Subsequently, samples were incubated with Pansorbin for 1 h, and the precipitates were washed with lysis buffer and analyzed using SDS-PAGE (15% acrylamide).

**Western Blot Analysis.** Total protein was prepared from whole liver or cultured hepatocytes by the addition of 1 ml of boiling lysis buffer (0.1% SDS, 10 mM Tris, pH 7.4) and subsequent sonication of liver or scraping of the cells. Cells then were boiled for 5 min and homogenized by passing through a 25-gauge needle. The homogenate was centrifuged for 5 min at 4°C, and the protein content was determined in the supernatant using the Bradford method. Total protein (40 µg) was loaded onto a 10% SDS-polyacrylamide gel and blotted onto nitrocellulose membranes by electrophoresis. Membranes were blocked with Tris-buffered saline containing 1% BSA, 10 mM Tris/HC1 (pH 7.5), and 0.1% Tween 20, 1 h at room temperature. The primary antibody for HO-1 was added in a 1:1000 dilution, and the blot was incubated for 12 h at 4°C. The enhanced chemiluminescent detection system was used for detection.

**RNA Isolation, Northern Blot Analysis, and Hybridization.** Total RNA for Northern blotting from hepatocytes, Kupffer cells, or whole liver was isolated as described (Immenschuh et al., 1998b, 1999). Equal quantities of RNA were separated on 1.2% agarose, 2.2 M formaldehyde gels. After electrophoresis, RNA was blotted onto nitrocellulose membranes and baked at 80°C for 4 h. After prehybridization for 4 h at 42°C, blots were hybridized overnight with 10⁻³⁻[^3²P]dCTP-radiolabeled cDNA probes at 42°C or a 28S rRNA oli-
gonucleotide as described previously (Immenschuh et al., 1999). The hybridization solution contained 6× standard saline citrate; 5× Den-
hardt's solution (0.2% Ficoll 400, 0.2% polyvinyl pyrrolidone, and 0.2% BSA); 0.5% SDS; 50% formamide; and 100 µg/ml denatured salmon sperm DNA. Blots were washed subsequently with 2× SSC/ 0.1% SDS (once) and 0.1× SSC/0.1% SDS (twice) at 65°C. Filters were autoradiographed with X-ray films (X-OMAT RP, Kodak; Rochester, NY) at −70°C for up to 48 h or stored on a phosphorimager screen for 4 to 8 h. Autoradiograms were quantified with phospho-
rimage running Imagequant software (Molecular Dynamics, Sunnyvale, CA). When nitrocellulose filters were sequentially hybridized with different cDNA probes, the [³²P]-labeled cDNA was removed after autoradiography by two washing steps with boiling 0.05× SSC/ 0.1% SDS for 15 min before rehybridization.

**cDNA Probes.** The probes were the cDNAs of HO-1, phosphoenolpyruvate carboxykinase (PKC), and glyceraldehyde-3-phosphate de-
hydrogenase (GAPDH) of rat (Immenschuh et al., 1998b). The cD-
NAs were labeled by the oligomer method with [³²P]dCTP using the multiprime DNA labeling kit according to the manufacturer's instructions.

**Isolation of Nuclei from Rat Hepatocyte Cultures.** Approximately 1 × 10⁶ cells from primary rat hepatocyte cultures were washed twice with ice-cold 320 mM sucrose, 3 mM CaCl₂, 2 mM magnesium acetate, 100 µM EDTA, 100 µM PMSF, 150 µM sperm-
mine, 500 µM spermidine, 1 mM dithioerythritol, and 10 mM Tris/HC1, pH 8.0 (buffer A). The cells were scraped off the dishes into buffer A and homogenized in a 2-m1 Dounce homogenizer at 4°C. After addi-

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tion of 4 ml of buffer A, the nuclei were pelleted by centrifugation at 300g for 5 min. The pellets were resuspended in 0.4 ml of buffer A, and the suspension was mixed with 1.6 ml 2 M sucrose, 5 mM magnesium acetate, 100 μM PMSF, 150 μM spermine, 500 μM spermidine, 1 mM dithiothreitol, and 10 mM Tris/HCl, pH 8.0 (buffer B). This suspension was layered onto a cushion of 2 ml of buffer B and pelleted for 1 h in a Beckman SW60 rotor at 20,000 rpm at 4°C. The pelleted nuclei were suspended in 25 ml of 25% buffer B and pelleted for 1 h in a Beckman SW60 rotor at 20,000 rpm at 4°C. The pelleted nuclei were suspended in 25 ml of 25% glycerol, 5 mM magnesium acetate, 100 μM EDTA, 100 μM PMSF, 5 mM dithiothreitol, and 50 mM Tris/HCl, pH 8.0 (buffer C).

**Nuclear Run-off Transcription Assay.** The nuclear run-off assay was performed with 2 × 10⁶ nuclei in a volume of 20 μl as described (Immenschuh et al., 1998b). In the in vitro transcription reaction was started by the addition of 30 ml of 58% glycerol, 150 mM NaCl, 8.3 mM MgCl₂, 830 μM MnCl₂, 70 μM EDTA, 25 U of ribonuclease inhibitor, 830 μM ATP, 830 μM CTP, 830 μM GTP, and 100 μCi [³²P]UTP, and 33 mM NHEPES, pH 8.0 (solution D). After incubation of nuclei for 30 min at 37°C, the reaction was stopped by the addition of EDTA.

RNA extraction, prehybridization, and hybridization were performed as described previously (Immenschuh et al., 1998b). In brief, prehybridization was performed in hybridization solution for 12 h at 42°C, followed by hybridization for 72 h at 42°C, using the rat HO-1 and GAPDH cDNAs immobilized on nitrocellulose membrane. As a control for the hybridization specificity, linearized pBR322 plasmid DNA was used. Posthybridization washes were performed in decreasing concentrations of NaCl/sodium citrate solution.

**Plasmid Constructs.** The rat HO-1 promoter 5'-flanking region from −1338 to +71 was amplified by PCR from rat genomic DNA by using the oligonucleotide 5’-CTCAGGATTAACAAAGACACAAACAGG-3’ (−1338/−1309) as forward and 5’-GAGATGGCTCTGCTGGCAGGCCTCACA-3’ (+42/+71) as reverse primer, respectively. The resulting PCR product was blunted by Klenow enzyme and phosphorylated with T4 polynucleotide kinase and ligated into the SmaI site of pUC18. The insert was excised with KpnI/BamHI and cloned into the KpnI/BglII site of pG3basic (Promega, Madison, WI) (pHO-1338 Luc; see Fig. 7, construct 1). Construction of plasmid pHO-754 Luc (see Fig. 7, construct 2), mutated rat HO-1 gene promoter constructs (see Fig. 7, pHO-754del, construct 3; and pHOACRE/AP-1, construct 4), and chloramphenicol acetyltransferase (CAT) construct pPCK-2500 CAT has been described previously in detail (Immenschuh et al., 1998a, Bratke et al., 1999). All constructs were verified by sequencing in both directions.

**Cell Transfection, Luciferase, and CAT Assay.** Rat hepatocyte cultures (~1 × 10⁶ cells per dish) were transfected transiently with 2.5 μg of plasmid DNA containing 500 ng of pRL-SV40 (Promega) to control transfection efficiency and 2 μg of the HO-1 promoter luciferase construct (Immenschuh et al., 1998a). Luciferase and CAT activity were determined as described previously (Immenschuh et al., 1998a; Bratke et al., 1999).

**PCK Enzyme Activity.** PCK enzyme activity was determined in duplicate as described previously (Bratke et al., 1999).

**Results**

**OA-Dependent Induction of HO-1 Gene Expression in Cultures of Primary Rat Hepatocytes.** To study the effect of the PP inhibitor OA on the synthesis of HO-1, newly synthesized proteins were pulse-labeled with [³⁵S]methionine in primary rat hepatocytes treated with OA at various concentrations, and HO-1 protein was immunoprecipitated from cell lysates. As shown in Fig. 1A, HO-1 protein was dose dependently up-regulated in the presence of OA. Next, we determined the effect of OA on steady-state levels of HO-1 mRNA. HO-1 message was markedly induced in hepatocyte cultures after 6 h (Fig. 1B). For comparison, no up-regulation of HO-1 mRNA expression by OA was observed in cultured rat liver tissue macrophages (Kupffer cells; Fig. 1C). The induction of HO-1 mRNA expression by heme, which is one of the most effective inducers of this enzyme, is shown as a positive control in Kupffer cells (Fig. 1C, lane 4). In two hepatoma cell lines (H35 and Hepa 1-6) and NIH3T3 fibroblasts, treatment with OA had no effect on HO-1 mRNA steady-state levels (data not shown). Because HO-1 activity has been shown to be increased during the first days of cell culture of primary rat hepatocytes (Schuetz et al., 1988), we compared HO-1 gene expression in our system of hepatocyte cultures with that in whole liver. Both the expression of HO-1 protein and mRNA were higher in 24-h cultured rat hepatocytes...
cytes compared with that in whole liver (Fig. 1D). Thereafter, HO-1 mRNA and protein declined and reached approximately the level of whole liver after 72 h of cell culture. To exclude the possibility that OA augments the effect of a stimulating factor, which may be generated during the isolation of rat hepatocytes, rather than stimulating the HO-1 gene expression per se, hepatocytes also were treated with OA after 120 h of cell culture. The OA-dependent induction of HO-1 gene expression in these long-term cultured hepatocytes was $14 \pm 1.5$-fold ($n = 3$) on the mRNA level and $4 \pm 0.6$-fold ($n = 3$) on the protein level (data not shown).

The increase of HO-1 mRNA expression after exposure to OA was time-dependent, reaching a transient maximum after 6 h (Fig. 2A). Moreover, the induction of HO-1 by OA was dose-dependent, with a peak at 10 nM. For normalization, the mRNA expression of GAPDH and that of the 28S rRNA were determined. Because GAPDH mRNA was not appreciably affected by the treatment with OA (Fig. 2, A and B; see also Fig. 1, B–D), in the following, GAPDH was used for normalization of HO-1 mRNA expression. The range of OA concentrations regulating HO-1 gene expression was similar to that observed for the mRNA regulation of the cytochrome P450 (CYP) isozymes 2B1 and 2B2 in primary rat hepatocyte cultures (Sidhu and Omiecinski, 1997). At OA concentrations $>50$ nM, hepatocytes were damaged severely as observed by light microscopy and lactate dehydrogenase leakage (data not shown). For a quantitative comparison, the effects of heme ($10 \mu$M), CoCl$_2$ (100 $\mu$M), and dibutyryl cAMP

**Fig. 2.** Time- and dose-dependent induction of HO-1 mRNA expression by OA in rat hepatocyte cultures. Primary rat hepatocytes were isolated and cultured as described in Experimental Procedures. Hepatocytes were treated for 18 h with serum-free medium before cell culture was continued (A) in the presence of OA (10 nM; lanes 2–7) for the times indicated, or (B) for 6 h in the absence (lane 1) or presence of increasing concentrations of OA (0.1–100 nM; lanes 2–6). Total cellular RNA (15 $\mu$g) was assessed by Northern blot analysis and sequentially probed with the $^{32}$P-labeled cDNAs of HO-1 and GAPDH or a 28S rRNA oligonucleotide. The size marker was the 18S ribosomal RNA band. Autoradiograms were quantitated by densitometry, and values represent the fold induction rate of HO-1 mRNA normalized to the 28S rRNA signal or that of the GAPDH mRNA normalized to the 28S rRNA signal from at least three independent experiments (mean ± S.E.).
(Bt2cAMP; 250 μM) on HO-1 mRNA expression in rat hepatocyte cultures are shown in Table 1. We also examined HO-1 mRNA expression in the presence of calyculin A, which has a lower inhibitory effect on PP2A, but a higher inhibitory effect on PP1 compared with OA (Holmes and Boland, 1993). In contrast to OA, calyculin A had no effect on steady-state levels of HO-1 mRNA (Table 1); however, it exhibited cell toxicity similar to that in OA. Comparable with these findings, a diverging effect of OA and calyculin A has been demonstrated previously for the regulation of nerve growth factor expression in primary rat astrocyte cultures (Pshenichkin and Wise, 1995).

From the data, we conclude that HO-1 gene expression is induced in a time- and dose-dependent manner by OA in primary cultures of rat hepatocytes.

**Differential Effects of OA on the Bt2cAMP-Dependent Induction of HO-1 and PCK Gene Expression.** Because HO-1 gene expression is induced by activation of PKA in primary rat hepatocyte cultures (Immenschuh et al., 1998b) and OA has been shown to augment the transcriptional response to cAMP (Hagiwara et al., 1992), hepatocytes were treated with the specific inhibitor of PKA, KT5720, before OA was added for another 6 h. As shown in Fig. 3A, pretreatment with KT5720 reduced the OA-dependent HO-1 mRNA induction by >50%. Moreover, simultaneous treatment of hepatocytes with Bt2cAMP and OA at submaximal doses caused a synergistic induction of HO-1 mRNA expression (Fig. 3B). To investigate the putative cross-talk of OA with the PKA signaling pathway, we also examined the effect of OA on the expression of the PCK gene, which is a liver-specific, cAMP-induced gene. PCK catalyzes the rate-controlling step of the gluconeogenic pathway and is induced by a variety of stimuli enhancing intracellular cAMP levels (for review, see Hanson and Reshef, 1997). In contrast to the OA-dependent regulation of HO-1 gene expression, OA on its own did not affect basal PCK mRNA expression or enzyme activity (Fig. 4). Simultaneous treatment of Bt2cAMP-treated hepatocytes with OA reduced the PCK mRNA expression and enzyme activity elicited by Bt2cAMP dose dependently (Fig. 4). These findings on the PCK gene expression are in agreement with those from a previous report in H4IIE rat hepatocyte cultures (O’Brien et al., 1994). Therefore, OA may affect the PKA signaling pathway in primary rat hepatocytes; however,

**TABLE 1** Comparative effects of treatment with OA, calyculin A, heme, CoCl2, and Bt2cAMP on HO-1 mRNA expression in primary cultures of rat hepatocytes

<table>
<thead>
<tr>
<th>Treatment</th>
<th>HO-1 mRNA induction (fold)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>1 ± 0.2</td>
</tr>
<tr>
<td>OA (10 nM)</td>
<td>13 ± 1.5*</td>
</tr>
<tr>
<td>Calyculin A (1 nM)</td>
<td>1 ± 0.2</td>
</tr>
<tr>
<td>Calyculin A (20 nM)</td>
<td>0.9 ± 0.1</td>
</tr>
<tr>
<td>Heme (10 μM)</td>
<td>40 ± 5*</td>
</tr>
<tr>
<td>CoCl2 (100 μM)</td>
<td>45 ± 6*</td>
</tr>
<tr>
<td>Bt2cAMP (250 μM)</td>
<td>19 ± 2*</td>
</tr>
</tbody>
</table>

**Fig. 3. Inhibition of the OA-dependent HO-1 mRNA induction by KT5720** and synergistic induction of HO-1 mRNA by OA and Bt2cAMP in rat hepatocyte cultures. Hepatocytes were cultured as described in Experimental Procedures. After 18 h in serum-free medium, cell culture was continued (A) in the presence of OA (10 nM), KT5720 (1 μM), and a combination of OA plus KT5720 for 6 h, or (B) in the presence of OA (5 nM), Bt2cAMP (100 μM), and a combination of OA plus Bt2cAMP for 6 h. Total RNA was assessed by Northern blot analysis and was sequentially probed with the 32P-labeled cDNAs of HO-1 and GAPDH. Autoradiograms were quantitated, and values represent the fold induction rate of HO-1 mRNA normalized to GAPDH from at least three independent experiments (mean ± S.E.).
strongly increased by OA (Fig. 5B). The turnover rate of HO-1 mRNA was determined in cell cultures after exposure to OA. As shown in Fig. 6, the half-life of HO-1 mRNA was slightly decreased during treatment with OA (4.7 h versus 4.2 h).

The data show that the induction of HO-1 gene expression by OA is primarily regulated on the transcriptional level and that de novo protein synthesis contributes partially to the HO-1 gene activation by OA in hepatocyte cultures.

OA-Dependent Induction of the Rat HO-1 Gene Promoter in Transiently Transfected Rat Hepatocyte Cultures. To investigate whether regulatory elements of the rat HO-1 5’-flanking promoter region are involved in the transcriptional regulation by OA, luciferase reporter constructs containing either the proximal 1338 or the 754 base pairs of the rat HO-1 promoter region were transiently transfected into primary rat hepatocyte cultures (Fig. 7, constructs 1 and 2; Table 2, pHO-1338 Luc and pHO-754 Luc). OA up-regulated the luciferase expression of these constructs 4- and 5.5-fold, respectively (Fig. 7), and a combination of submaximal doses of OA plus Bt2cAMP induced luciferase expression additively (Table 2). An HO-1 reporter construct with a deletion from −714 to −549 (Fig. 7, construct 3) and a construct lacking the CRE/AP-1 site (Fig. 7, construct 4) were not regulated by OA (Fig. 7). For a comparison, the regulation of a CAT reporter construct containing 2500 base pairs of the rat PCK promoter 5’-flanking region was examined in transfected rat hepatocytes (Table 2). Whereas treatment with OA alone had no effect, the Bt2cAMP-dependent induction of this reporter construct was inhibited by OA (Table 2, pPCK-2500 CAT).

Discussion

In this study, it is shown in cultured rat hepatocytes that the serine threonine PP inhibitor OA up-regulates the gene expression of HO-1, which is the inducible enzyme of heme degradation. The OA-dependent HO-1 induction occurs on the transcriptional level and is mediated by a DNA sequence of the HO-1 gene promoter 5’-flanking region.

The OA-dependent increase of HO-1 gene expression is primarily regulated on the transcriptional level, as demon-
Procedures

Hepatocytes were cultured either in the absence (left) or presence of OA (right) (10 nM) for 6 h, after which cell culture was continued with ActD (1 μg/ml). Total RNA was isolated at the times indicated, and the levels of HO-1 mRNA were determined by Northern blot analysis. The primary plot is a semilog plot of individual points of two independent experiments (mean ± S.E.). The half-lives calculated from the graphs are indicated, respectively.

Stabilization of HO-1 message is not involved in the OA-dependent HO-1 gene regulation (Fig. 6), but protein synthesis de novo appears to contribute to this induction, as indicated by the inhibition of OA-dependent HO-1 mRNA expression by CHX (Fig. 5A) suggesting that protein(s) with a short half-life participate(s) in this regulatory pathway. Therefore, OA adds to the various stimuli that modulate the transcription rate of the HO-1 gene (Shibahara et al., 1987; Applegate et al., 1991; Durante et al., 1997; for review, see Choi and Alam, 1996). Several REs within the promoter 5′-flanking region that are involved in the activation of the human, mouse, and rat HO-1 genes have been characterized (for review, see Choi and Alam, 1996). As demonstrated by transient transfection of rat HO-1 gene reporter constructs into rat hepatocytes, the CRE/AP-1 element of the rat HO-1 gene (position –665 to –654) (Immenschuh et al., 1998a) is involved in the OA-dependent regulation of the HO-1 gene (Fig. 7). Although deletion of the HO-1 CRE/AP-1 element abolished the OA-dependent induction of luciferase reporter gene activity, it cannot be excluded that additional REs are involved in the OA-dependent gene regulation. A potential transcription factor (TF) that may mediate the OA-dependent transcriptional induction is the CRE-binding protein (CREB), which is activated on phosphorylation at Ser-133. Hagiwara et al. (1992) have demonstrated that OA inhibits the dephosphorylation of the Ser-133 of phospho-CREB, thereby augmenting cAMP-dependent gene expression. The hypothesis that CREB may mediate the OA-dependent HO-1 induction is supported by the observations that OA and Bt2cAMP elicit a synergistic effect on HO-1 mRNA up-regulation and that the specific PKA inhibitor KT5720 reduces the induction of HO-1 mRNA expression by OA (Fig. 3). Moreover, the pHO-1338 Luc and pHO-754 Luc HO-1 gene reporter constructs are up-regulated additively by submaximal doses of OA and Bt2cAMP (Table 2). Whether the Ser-133 of CREB is dephosphorylated by PP1 or PP2A appears to be cell type-dependent. Alberts et al. (1994) have shown that PP1 is the major regulator of dephosphorylation of CREB in fibroblasts. By contrast, others have demonstrated in rat liver and HepG2 hepatoma cells that PP2A dephosphorylates phospho-CREB 30-fold more efficiently than does PP1 (Wadzinski et al., 1993). The latter finding would correlate with our observation that OA, but not calyculin A, induced HO-1 gene expression in rat hepatocyte cultures at the applied concentrations (Table 1). OA has been reported to inhibit PP2A ~5- to 10-fold stronger than does calyculin A, whereas calyculin A is a significantly stronger inhibitor of PP1 than OA (Holmes and Boland, 1993). In contradiction to the idea that CREB may mediate the OA-dependent HO-1 induction on its own is the inhibitory effect of OA on the cAMP-dependent PCK gene expression. The cAMP-dependent induction of the PCK gene is known to be primarily mediated via a CRE (Hanson and Reshef, 1997); however, in our system of primary rat hepatocyte cultures, the cAMP-dependent induction of the PCK gene was inhibited by OA (Fig. 4, Table 2), as similarly reported in H4IIE hepatoma cells (O’Brien et al., 1994). It also has been shown that the liver-specific induction of the PCK gene promoter requires synergism of the TFs CREB and C/EBPα to mediate the full cAMP response in hepatic cells (Roesler et al., 1996). Other

### Table 7

<table>
<thead>
<tr>
<th>Construct</th>
<th>Fold Stimulation</th>
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<tbody>
<tr>
<td>1</td>
<td>1 4.0 ± 1.1*</td>
</tr>
<tr>
<td>2</td>
<td>1 5.5 ± 1.0*</td>
</tr>
<tr>
<td>3</td>
<td>1 1.2 ± 0.2</td>
</tr>
<tr>
<td>4</td>
<td>1 1.3 ± 0.2</td>
</tr>
<tr>
<td>5</td>
<td>0.1 0.1 ± 0.1</td>
</tr>
</tbody>
</table>

Fig. 6. Effect of OA on the half-life of HO-1 mRNA in rat hepatocyte cultures. Rat hepatocytes were cultured as described in Experimental Procedures. Hepatocytes were cultured either in the absence (left) or presence of OA (right) (10 nM) for 6 h, after which cell culture was continued with ActD (1 μg/ml). Total RNA was isolated at the times indicated, and the levels of HO-1 mRNA were determined by Northern blot analysis. The primary plot is a semilog plot of individual points of two independent experiments (mean ± S.E.). The half-lives calculated from the graphs are indicated, respectively.

Fig. 7. OA-dependent regulation of DNA sequences of the rat HO-1 gene promoter 5′-flanking region in transiently transfected rat hepatocyte cultures. The indicated rat HO-1 gene sequences were cloned into pGL3Luc (constructs 1–4) as described in Experimental Procedures. The reporter constructs were transiently transfected into primary rat hepatocyte cultures and, after 24 h, the transfected cells were treated for 12 h with OA (10 nM). The rate of induction in each experiment relative to the control was determined. Regulation of luciferase activity of pGL3prom is shown as a control. The values are from at least three independent experiments (mean ± S.E.). Student’s t test for paired values: * indicates significant differences control versus OA, P < .05. In construct 3, H indicates HindIII restriction site.
TABLE 2
Regulation of DNA sequences of the rat HO-1 and PKC gene promoter

<table>
<thead>
<tr>
<th>Construct</th>
<th>Fold induction of reporter gene activity</th>
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<tr>
<td></td>
<td>OA</td>
</tr>
<tr>
<td>PHO-1338 Luc</td>
<td>2.5 ± 0.2</td>
</tr>
<tr>
<td>PHO-754 Luc</td>
<td>3 ± 0.5</td>
</tr>
<tr>
<td>pCCK-2500 CAT</td>
<td>1 ± 0.2</td>
</tr>
<tr>
<td>pGL3prom Luc</td>
<td>0.1 ± 0.2</td>
</tr>
</tbody>
</table>

The indicated rat HO-1 or PKC gene sequences were cloned into pGL3Luc or pcAT, as indicated, and the reporter constructs were transiently transfected into primary rat hepatocyte cultures. After 24 h, the transfected cells were treated for 12 h with OA (5 mM), Bt2cAMP (100 μM) or a combination of OA plus Bt2cAMP. The rate of induction in each experiment relative to the control was determined. The values are from three independent experiments (mean ± S.E.). Student’s t test for paired values: *indicates significant difference OA versus OA + Bt2cAMP; P ≤ 0.05; **significant difference OA versus Bt2cAMP; P ≤ 0.65.


Rosenberger SF and Bowden GT (1996) Okadaic acid stimulated TRE binding activity in a papilloma producing mouse keratinocyte cell line involves increased AP-1 expression. Oncogene 15:2301–2308


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