Sensitized Increase of *Period* Gene Expression in the Mouse Caudate/Putamen Caused by Repeated Injection of Methamphetamine

TAKATO NIKAIDO, MASASHI AKIYAMA, TAKAHIRO MORIYA, and SHIGENOBU SHIBATA

Department of Pharmacology and Brain Science (T.N., M.A., S.S.) and Advanced Research Center for Human Sciences (T.M., S.S.), School of Human Sciences, Waseda University, Tokorozawa, Saitama, Japan

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ABSTRACT

Methamphetamine (MAP) causes the sensitization phenomena not only in MAP-induced locomotor activity, dopamine release, and Fos expression, but also in MAP-induced circadian rhythm. Cocaine-induced sensitization is reportedly impaired in *Drosophila melanogaster* mutant for the *Period* (*Per*) gene. Thus, sensitization may be related to induction of the *Per* gene. A rapid induction of *mPer1* and/or *mPer2* in the suprachiasmatic nucleus after light exposure is believed to be necessary for light-induced behavioral phase shifting. Although the caudate/putamen (CPu) expresses *mPer1* and/or *mPer2* mRNA, the function of these genes in this nucleus has not yet been elucidated. Therefore, we examined whether MAP affects the expression of *mPer1* and/or *mPer2* mRNA in the mouse CPu. Injection of MAP augmented the expression of *mPer1* but not *mPer2* or *mPer3* in the CPu, and this MAP-induced increase in *mPer1* expression lasted for 2 h. Also, the MAP-induced increase of *mPer1* mRNA was strongly antagonized by pretreatment with a dopamine D1 receptor and N-methyl-D-aspartate (NMDA) receptor antagonist, but not by a D2 receptor antagonist. Interestingly, application of either the D1 or the D2 agonist alone did not cause *mPer1* expression. The present results demonstrate that activation of both NMDA and D1 receptors is necessary to produce MAP-induced *mPer1* expression in the CPu. Repeated injection of MAP caused a sensitization in not only the locomotor activity but also *mPer1* expression in the CPu without affecting the level of *mPer2*, *mPer3*, or *mTim* mRNA. Thus, these results suggest that MAP-induced *mPer1* gene expression may be related to the mechanism for MAP-induced sensitization in the mouse.

A core clock mechanism in the mouse suprachiasmatic nucleus (SCN) seems to involve a transcriptional feedback loop in which CLOCK and BMAL1 function as positive regulators and the three *mPeriod (mPer)* genes, *Per1* (Sun et al., 1997; Tei et al., 1997), *Per2* (Albrecht et al., 1997; Shearman et al., 1997; Takumi et al., 1998), and *Per3* (Zylka et al., 1998) are involved in negative feedback (Dunlap, 1999). In addition, it was determined that two mouse cryptochrome genes, *mCry1* and *mCry2*, act in the negative limb of the clock feedback loop (Kume et al., 1999). It is already well known that the SCN contains a master pacemaker that regulates behavioral and physiological circadian rhythms such as locomotor activity, body temperature, and endocrine release (Inouye and Shibata, 1994). Interestingly, expression of the *Per* gene occurs not only in the SCN but also in other brain areas such as the cerebral cortex, caudate/putamen (CPu), and cerebellum (Albrecht et al., 1997; Shearman et al., 1997). However, the function of clock genes outside of the SCN has not been fully elucidated.

 Destruction of the SCN abolishes the circadian rhythms of many physiological functions (Inouye and Shibata, 1994). On the other hand, there are at least two oscillators outside the SCN: a food-associated oscillation entrained by daily restricted feeding (Mistlberger, 1994) and methamphetamine (MAP)-induced oscillation produced by its daily injection (Shibata et al., 1994, 1995). In addition, oral administration of MAP through drinking bottle initiates a circadian rhythm with a long free-running period even after SCN ablation (Honma et al., 1987). Based on these facts, it has been suggested that other circadian oscillators such as the MAP-induced rhythm exist in areas other than the SCN. Thus, it is possible that *mPer* mRNA outside of the SCN regulates the SCN-independent circadian rhythm, and rapid induction of *mPer* outside of the SCN by MAP may entrain the SCN-independent oscillation.

A Abbreviations: SCN, suprachiasmatic nucleus; MAP, methamphetamine; CPu, caudate/putamen; NMDA, N-methyl-D-aspartate; Per, Period; Tim, timeless; PB, phosphate buffer; PFA, paraformaldehyde.
Recently it was reported that sensitization to repeated cocaine exposure, a phenomenon also seen in humans and animal models and associated with enhanced drug craving, is eliminated in flies mutant for period, clock, cycle, and doubletime, but not in flies mutant for timeless (Andretic et al., 1999). We demonstrated that the MAP-induced free-running oscillation of rat locomotion with drinking application of MAP exhibits a sensitization phenomenon (Nikaido et al., 1999). Therefore, the next progressive step was to examine whether MAP induces Per expression in the CPu, and whether sensitization is involved in MAP-induced Per expression but not timeless expression.

Treatment with MAP is known to increase locomotor activity and Fos expression in the CPu (Graybiel et al., 1990). Pharmacological studies have further revealed that both MAP-induced hyperlocomotion and Fos expression in the CPu are attenuated by pretreatment with dopamine D1, D2, or NMDA receptor antagonists (Ujike et al., 1989; Kuribara and Uchihashi, 1993; Kuribara, 1994, 1995, 1996; Yoshida et al., 1995). Thus, it has been suggested that D1, D2, and NMDA receptors play an important role in the sensitization induced by repeated injection of MAP. This evidence suggests that MAP-induced Per expression may be involved in the activation of both dopamine and NMDA receptors. Therefore, in the first part of our present experiment, we examined the pharmacological characteristics of MAP-induced Per gene expression in the mouse CPu. Then, in the latter part, we examined the expression pattern of Per and timeless mRNA using animals sensitized to MAP.

![Fig. 1. Effect of 5 mg/kg of MAP on mPer1 expression in CPu, accumbens (NAcc), and piriform cortex (PC). Mice were decapitated 60 min after MAP injection. A, representative in situ hybridization autoradiograms of mPer1 on X-ray film. The brain areas surrounded by a broken line exhibit the NAcc, CPu, and PC areas. B, radioisotope in situ hybridization was performed for quantitative analysis purposes. Numbers in parentheses indicate the number of animals (*P < 0.05 in comparison with saline by Student’s t test). C, emulsion autoradiograms of mPer1 in the dorsal caudate. A 5-mg/kg injection of MAP increased mPer1 expression (black dots, middle panel), and there were many dots on cells showing the Fos immunoreactivity (right panel). Fos immunoreactive cells were stained by a brown color.](image)

![Fig. 2. Time course of MAP-induced locomotor activity (A) and mPer expression in the CPu or SCN (B). A, MAP increased mouse locomotion for 4 h. *: saline injection (n = 4); †, 5 mg/kg MAP injection (n = 4) (*P < 0.05 in comparison with saline by Student’s t test). B, relative value of mPer1 and mPer2 mRNA expression in the CPu. #: saline injection; †, 5 mg/kg MAP injection. Per expression observed in the saline group immediately after saline injection was set as 100%. Three to four animals made up each point (*P < 0.05 in comparison with saline by Student’s t test; *P < 0.05, **P < 0.01 in comparison with 0-min point by Dunnett’s test).](image)
Materials and Methods

Animals. In all experiments, we used 4- to 6-week-old male ddY mice (Takasugi, Saitama, Japan) maintained under a 12 h:12 h light/dark cycle. All animals were allowed free access to food and water and were treated in accordance with the Law no. 105 and Notification no. 6 of the Japanese Government.

Locomotor Activity Measurement. For assessment of the locomotor activity, mice were housed individually in transparent plastic cages (31 x 20 x 13 cm). Motor activity was measured using an infrared area sensor (F5E; Omron, Tokyo, Japan), and the activity count (number of movements) was recorded by computer and stored on disk at 5-min intervals.

Sample Preparation. Mice were deeply anesthetized with ether and intracardially perfused with 0.1 M phosphate buffer (PB), pH 7.4, containing 4% paraformaldehyde (PFA). Brains were removed, postfixed in 0.1 M PB containing 4% PFA for 24 h at 4°C, and transferred into 20% sucrose in PB for 72 h at 4°C. Brain slices (40 μm thick) including the CPu, accumbens, piriform cortex, and SCN were made using a cryostat (HM505E; Microm, Walldorf, Germany) and placed in 2× standard saline citrate until processing for hybridization.

In Situ Hybridization. The quantity of mPer1, mPer2, mPer3, or mTim mRNA expression in the various brain areas was studied by means of in situ hybridization. Slices were treated with 1 μg/ml proteinase K in 10 mM Tris-HCl buffer, pH 7.5, containing 10 mM EDTA for 10 min at 37°C followed by 0.25% acetic anhydride in 0.1 M triethanolamine and 0.9% NaCl for 10 min. The slides were then incubated in the hybridization buffer [60% formamide, 10% dextran sulfate, 10 mM triethanolamine and 0.9% NaCl for 10 min. Brain slices were fixed with 4% PFA and 10% dextran sulfate, 10 mM Tris-HCl, pH 7.4, 0.1 M NaCl, 1× Denhardt’s solution (0.02% Ficoll, 0.02% polyvinyl pyrrolidone, 0.02% bovine serum albumin), 0.2 M NaCl, and 0.25% sodium dodecyl sulfate] containing 33P-labeled cRNA probes for 16 h at 60°C. Radioisotope (1×33P]UTP-labeled antisense cRNA probes (PerkinElmer Life Sciences, Boston, MA) were made from restriction enzyme-linearized cDNA templates [nucleotide positions: mPer1 (538-1752), mPer2 (1-638), mPer3 (814-1955), mTim (236-909)] kindly provided by Dr. Okamura (Kobe University, Kobe, Japan). After a high-stringency posthybridization wash in 2× standard saline citrate/50% formamide, slices were treated with RNaseA (10 μg/ml) for 30 min at 37°C.

The radioactivity of each slice visualized on BioMax MR film (Eastman Kodak, Rochester, NY) was analyzed using a microcomputer interface to an image analysis system (MCID; Imaging Research Inc., ON, Canada) after conversion into optical density by 14C-autoradiographic microscales (Amersham Pharmacia Biotech, Buckinghamshire, UK). For data analysis, we subtracted the intensities of the optical density in sections from the corpus callosum from those in the SCN, CPu, piriform cortex, and accumbens and regarded this value as the net intensity for these areas. The intensity values of sections from the rostral to the caudal part of the SCN, CPu, piriform cortex, and accumbens (three to five sections per mouse brain) were then summed; the sum was considered to be a measure of the amount of mPer1, mPer2, mPer3, or mTim mRNA in this region. To express the relative mRNA abundance (Figs. 2–5 and 7), the intensity values of vehicle treatment were adjusted to 100.

Immunohistochemistry of Fos Protein and Emulsion Autoradiography of mPer1. The slices were fixed with 4% PFA and processed for immunohistochemistry according to the avidin-biotin-peroxidase complex method. Primary antibody (anti-Fos, 1:5000; Cambridge Research Biochemical, Northwich, UK) was diluted in 0.1 M phosphate buffer containing 1% normal goat serum in 0.3% Triton X-100.

For emulsion autoradiography, slices already processed for immunohistochemistry were dipped into emulsion (NTB2, Eastman Kodak; diluted 1:1 with distilled water) after hybridization with the mPer1 probe, air dried for 3 h, and stored in light-tight slide boxes at 4°C for 2 weeks. The slides were developed using a D19 developer (Eastman Kodak) and then fixed with Fujifilm (Fujifilm, Tokyo, Japan). Subnuclear slice distribution in the CPu was examined using an optical microscope. We did not adopt the quantitative analysis of emulsion autoradiogram because thickness of the coating could not be controlled using the present emulsion-dipping method.

Drugs and Application Schedule. The drugs used in this experiment consisted of methamphetamine HCl (Dainippon Co., Tokyo, Japan); SCH23390 (Funakoshi, Tokyo, Japan); (+)-sulpiride (Sigma, St. Louis, MO); and MK-801, SKF38393, and quinpirole (RBI/Sigma, Natick, MA). All drugs were dissolved into the physiological saline. Drugs were injected during the daytime because spontaneous locomotor activity and mPer1 expression in the CPu were low at this time (data not shown). To examine the blocking effect of receptor antagonists on MAP-induced mPer1 expression, these receptor antagonists were injected 15 min before MAP injection.

A single high-dose exposure to MAP or amphetamine sufficiently induces long-term behavioral and neurochemical sensitization (Ohno et al., 1994; Vanderschuren et al., 1999). Therefore, for sensitization experiments, MAP (5 mg/kg) was injected once, and then a small dose of MAP (0.5 mg/kg) was injected again after a 7-day interval.

Statistics. Results are expressed as the mean ± S.E.M. The significance of differences between groups was determined by two-way or one-way analysis of variance followed by Dunnett’s test or Student’s t test.

Fig. 3. Dose-response of MAP-induced locomotor activity (A) and mPer expression in the CPu (B). A, vertical values exhibit total locomotor count (number of movements) was recorded by computer and stored on disk at 5-min intervals. B, relative value of mPer1, mPer2, and mPer3 mRNA expression in the CPu. Per expression observed in the saline group was set as 100%. Three animals made up each point (P < 0.05 in comparison with saline by Dunnett’s test).
Results

Methamphetamine-Induced mPer1 Expression in the Various Brain Areas. It is well known that dopaminergic neurons innervate the CPu, accumbens, and piriform cortex. Therefore, we examined the amount of mPer1 expression in these brain areas. Figure 1 shows the representative brain areas responding to MAP and the sampling area for each brain slice. Basal level mPer1 expression was high in the piriform cortex but low in the CPu and accumbens (Fig. 1, A and B). Unrelated to basal expression, MAP significantly induced mPer1 expression in the CPu (P < 0.05, Student’s t test) and piriform cortex (P < 0.05, Student’s t test) but not in the accumbens (Fig. 1, A and B). Previous papers demonstrated a strong induction of Fos protein in the dorsal CPu (Yoshida et al., 1995); therefore, we examined the Fos expression and mPer1 induction using CPu slices. Interestingly, mPer1 mRNA and Fos immunoreactivity were coexpressed in the same striatal cells (Fig. 1C).

Time Course of MAP-Induced Locomotion and mPer1 Expression. A 5-mg/kg injection of MAP significantly increased the locomotion, which was maintained for 4 h (Fig. 2A). Two-way analysis of variance revealed an interaction between drug treatment and time course in CPu mPer1 (P < 0.01, Student’s test) and piriform cortex (P < 0.01, Student’s t test) but not in the accumbens (Fig. 1, A and B). The increased basal level of mPer1 mRNA expression in the CPu was set as 100%. Antagonists were injected 15 min before the 5-mg/kg MAP injection. Numbers in parentheses indicate the number of animals (**P < 0.01 in comparison with saline by Student’s t test; *P < 0.01 in comparison with MAP injection only by Dunnett’s test). B, representative in situ hybridization autoradiograms of mPer1 were developed on X-ray film. The D1 receptor antagonist (SCH) and NMDA receptor antagonist (MK-801), but not the D2 receptor antagonist (SUL), antagonized MAP-induced mPer1 expression. Calibration on the saline panel is 1 mm. C, relative value of mPer1 mRNA expression in the CPu. Per expression observed in the saline group was set as 100%. Antagonists were injected 15 min before the 5-mg/kg MAP injection, and then mice were decapitated 60 min after MAP injection. Numbers in parentheses indicate the number of animals (**P < 0.01 in comparison with saline by Student’s t test; *P < 0.01 in comparison with MAP injection only by Dunnett’s test). SCH, SCH23390; SUL, sulpiride.
sion in the CPu was attenuated by the D1 receptor antagonist and facilitated by the D2 receptor antagonist, we examined the effect of D1 and D2 receptor agonists on the basal level of mPer1 expression. Application of D1 receptor agonist SKF38393 did not affect mPer1 and mPer2 expression in the CPu (Fig. 5), whereas the D2 receptor agonist, quinpirole, slightly reduced mPer1 and mPer2 expression (Fig. 5).

Sensitized Expression of Locomotion and mPer mRNA in the CPu with Repeated Injection of MAP. In preparing the four experimental groups, MAP or saline was injected initially into all mice, then half of the MAP- (5 mg/kg) or saline-injected groups received another injection of MAP (0.5 mg/kg) or saline. Figure 6, A and B demonstrates the time course of locomotion (Fig. 6A) after injection of saline or MAP (0.5 mg/kg) and total locomotor counts (Fig. 6B) 60 min after injection, respectively. Small doses of MAP at 0.5 mg/kg did not increase locomotion in the saline-pretreated group but significantly increased locomotion in MAP (5 mg/kg)-pretreated mice \(F(3,12) = 6.9, P < 0.01\) (Fig. 6B).

A second injection of MAP (0.5 mg/kg) strongly increased mPer1 mRNA in the CPu of mice pretreated with MAP at 5 mg/kg \(F(3,12) = 20.1, P < 0.01\) (Fig. 7, A and B), but 0.5 mg/kg of MAP did not affect the expression of mPer2 \(F(3,12) = 1.8, P > 0.05\) or mTim mRNA \(F(3,12) = 1.1, P > 0.05\) (Fig. 7B).

Discussion

In the present experiment, we demonstrated that MAP dose dependently induces the expression of mPer1 in the CPu and piriform cortex but not in the accumbens or SCN, whereas MAP did not affect the levels of mPer2 and mPer3 in these brain areas. Coadministration of D1 receptor antagonist SCH23390 or NMDA receptor antagonist MK-801 significantly attenuated MAP-induced expression of mPer1, but D2 receptor antagonist sulpiride did not block this expression. Interestingly, Fos induction in the CPu by MAP injection was reportedly attenuated by either SCH23390 or MK-801 (Konradi et al., 1994, 1996; Ohno et al., 1994; Yoshida et al., 1995). Treatment with MK-801 attenuated MAP-induced mPer1 expression in the CPu. Previous reports demonstrated that MK-801 inhibited amphetamine-induced glutamate release in the ventral tegmental area (Wolf and Xue, 1999), MAP-induced striatal dopamine release (Finnegan and Taraska, 1996), and striatal Fos expression (Ohno et al., 1994). Thus, NMDA receptor mechanisms are also involved in MAP-induced biochemical responses such as mPer1 and Fos expression. From a pharmacological point of view, above-mentioned articles have suggested that there may be common neural mechanisms between the expression of Fos and of mPer1 induced by MAP. The present double-staining experiment demonstrated a dense expression of both Fos and mPer1 mRNA in the dorsomedial regions of the CPu, which also support the above possibility.

In the present experiment, MAP increased the level of mPer1 but did not affect the levels of mPer2 and mPer3 in the CPu. Application of forskolin induced Per1 but not Per2 expression in Rat-1 cells with the induction of cyclic AMP-responsive element binding protein phosphorylation; then it initiated the oscillation of Per2 expression (Yagita and Oka-
mura, 2000). Interestingly, we found that the promoter region of mPer1 contains a total of four cyclic AMP-responsive elements (Yamaguchi et al., 2000). This cyclic AMP-responsive element site may be responsible for the mPer1 induction that occurs with MAP application. In fact, not only D1 and NMDA receptor activation (Das et al., 1997) but also MAP application (Muratake et al., 1998) reportedly cause cyclic AMP-responsive element binding protein phosphorylation.

Light exposure strongly increases Per1 and Per2 but not Per3 expression in the SCN of mice, rats, and hamsters (Shigeyoshi et al., 1997; Yan et al., 1999; Horikawa et al., 2000); however, in our study, MAP failed to change mPer gene expression in the SCN. We found a significant circadian oscillation of Per1 and Per2 in the hamster SCN with a peak at subjective day (Horikawa et al., 2000). Local injection of NMDA into the SCN at subjective night causes the induction of Per1 in the hamster (Moriya et al., 2000). On the other hand, stimulation of the D1 receptor in the SCN produces Fos induction in early developmental rodents but not in adults (Viswanathan et al., 1994; Weaver and Reppert, 1995; Grose and Davis, 1999). Thus, the reason why MAP failed to produce mPer1 expression in the SCN may be related to weak contribution of D1 receptors in the adult SCN. Changes in mPer1 mRNA levels of CPu detected by in situ hybridization exhibited a circadian rhythm with a peak at early subjective night (data not shown). Therefore, it is interesting to determine whether transiently induced mPer1 in the CPu at subjective day may cause a phase shift of circadian rhythm of mPer1 expression in the CPu. The answer requires an examination of circadian time course of mPer1 expression in the CPu subsequent to MAP treatment, and this important question should be the follow-up to this article.

In this experiment, SCH23390 and MK-801 administration alone lowered the expression of mPer1 in the CPu, suggesting a tonic activation of the mPer1 gene through D1 and/or NMDA receptors in the CPu. Actually, MK-801 decreased the MAP-induced mPer1 expression but slightly augmented the MAP-induced locomotion. On the contrary, sulpiride strongly attenuated the MAP-induced locomotion without affecting mPer1 expression. These results seemingly indicate that MAP-induced locomotor stimulation is not sufficient for induction of mPer1 by MAP. One of our previous articles supports this idea by showing that the D1 and NMDA receptor blockade abolished MAP-induced anticipatory behavior without attenuating its induction of hyperlocomotion (Shibata et al., 1995).

In the present experiments, repeated administration of MAP caused sensitization in mPer1 expression but not in mPer2, mPer3, or mTim expression in the CPu. Therefore, behavioral sensitization is associated with mPer1 expression in the CPu but not that in the SCN, suggesting the important role of mPer1 gene expression in MAP-induced behavioral sensitization. Furthermore, we demonstrated that the MAP-induced free-running oscillation of rat locomotion with drinking application of MAP exhibited a sensitization phenomenon (Nikaido et al., 1999). Interestingly, Andretic et al. (1999) reported that flies mutant for period, clock, cycle, and double-time lack sensitization to repeated cocaine administration, but flies mutant for timeless do not. On the other hand, in contrast to Drosophila melanogaster, mutation of the Clock gene that regulates circadian rhythm in mice does not affect acute or sensitized responses to cocaine (Sidiropourou et al., 2000). Thus, in mammals, the Clock gene is not required for the induction of behavioral sensitization to cocaine. Therefore, we should investigate whether the sensitized increase in mPer1 expression reflects the result or cause using mPer1 gene mutant mice. Taken together, the results seem to indicate that MAP-induced sensitized expression of mPer1 may be related, at least, to the sensitized phenomenon.

In conclusion, the present results demonstrate that the activation of both D1 and NMDA receptors plays an important role in causing MAP-induced expression of mPer1 in the CPu. Furthermore, behavioral sensitization induced by repeated MAP injection is associated with sensitization of MAP-induced mPer1 expression.

References


Send reprint requests to: Shigenobu Shibata, Department of Pharmacology and Brain Science, School of Human Sciences, Waseda University, Tokyo, 152-8558, Japan. E-mail: shibata@human.waseda.ac.jp