Effects of Valproic Acid Derivatives on Inositol Trisphosphate Depletion, Teratogenicity, Glycogen Synthase Kinase-3β Inhibition, and Viral Replication: A Screening Approach for New Bipolar Disorder Drugs Derived from the Valproic Acid Core Structure


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ABSTRACT
Inositol-1,4,5-trisphosphate (InsP3) depletion has been implicated in the therapeutic action of bipolar disorder drugs, including valproic acid (VPA). It is not currently known whether the effect of VPA on InsP3 depletion is related to the deleterious effects of teratogenicity or elevated viral replication, or if it occurs via putative inhibitory effects on glycogen synthase kinase-3β (GSK-3β). In addition, the structural requirements of VPA congeners to cause InsP3 depletion are unknown. In the current study, we selected a set of 10 VPA congeners to examine their effects on InsP3 depletion, in vivo teratogenic potency, HIV replication, and GSK-3β activity in vitro. We found four compounds that function to deplete InsP3 in the model eukaryote Dictyostelium discoideum, and these drugs all cause growth-cone enlargement in mammalian primary neurons, consistent with the effect of InsP3 depletion. No relationship was found between InsP3 depletion and teratogenic or elevated viral replication effects, and none of the VPA congeners were found to affect GSK-3β activity. Structural requirements of VPA congeners to maintain InsP3 depletion efficacy greater than that of lithium are a carboxylic-acid function without dependence on side-chain length, branching, or saturation. Noteworthy is the enantiomeric differentiation if a chiral center exists, suggesting that InsP3 depletion is mediated by a stereoselective mode of action. Thus, the effect of InsP3 depletion can be separated from that of teratogenic potency and elevated viral replication effect. We have used this to identify two VPA derivatives that share the common InsP3-depleting action of VPA, lithium and carbamazepine, but do not show the side effects of VPA, thus providing promising novel candidates for bipolar disorder treatment.

ABBREVIATIONS: VPA, valproic acid; GFP, green fluorescent protein; GSK-3β, glycogen synthase kinase 3β; HIV, human immunodeficiency virus; InsP3, inositol-1,4,5-trisphosphate; PO, prolyl oligopeptidase; VPD, valpromide; FACS, fluorescence-activated cell sorting; PBS, phosphate-buffered saline; DRG, dorsal root ganglia; NMRI, Naval Medical Research Institute (mouse model); GSM, GSK3 substrate modified.
ium works by “dampening down” an overactive inositol-1,4,5-trisphosphate (InsP$_3$) signaling cascade. The effect of bipolar disorder treatments on InsP$_3$ signaling is becoming increasingly apparent. Lithium acts as an uncompetitive inhibitor of a family of phosphatases that includes inositol monophosphate (Leech et al., 1993) and polyphosphatase (York et al., 1995), two enzymes involved in the breakdown and recycling of InsP$_3$. Both VPA and lithium decrease the amount of inositol (O'Donnell et al., 2003) and attenuate InsP$_3$ signaling (Li et al., 1993) in the rat brain. In addition, they change membrane lipid concentration, a process directly linked to InsP$_3$ signaling (Ding and Greenberg, 2003). Indeed, a recent study in patients with bipolar disorder suggested that altered InsP$_3$ signaling may be corrected by both VPA and lithium (Silverstone et al., 2002).

This inositol depletion theory of bipolar disorder drug action has been strengthened by two articles describing the effects of these treatments initially in Dictyostelium discoideum and later in primary rat neurons (Williams et al., 1999, 2002). Inhibition of the D. discoideum enzyme prolyl oligopeptidase (PO) gave rise to lithium resistance via the elevation of basal InsP$_3$ levels, thus overcoming the drug-induced InsP$_3$ depletion effect through an unknown mechanism. This work identified lithium, VPA, and carbamazepine as acting via InsP$_3$ depletion in primary mammalian neurons, a mechanism also controlled by PO activity in astroglialoma cell lines (Schulz et al., 2002). It is interesting that patients with bipolar disorder show altered activity of PO (Breen et al., 2004), suggesting altered InsP$_3$ signaling in this disorder. Finally, all three drugs alter inositol uptake in human astrocytoma cells (Wolfson et al., 2000). Despite these indications, both the primary target for VPA and carbamazepine in InsP$_3$ depletion and the structural requirements of VPA for this effect remain unknown, and no links exist between VPA side effects (such as teratogenicity) and inositol depletion.

Subsequent to the inositol depletion theory, a second target has been proposed for bipolar disorder drugs, the enzyme glycogen synthase kinase 3β (GSK-3β). This enzyme has been shown widely to be targeted by lithium (Klein and Melton, 1996; Stambolic et al., 1996), and it has also been reported to be directly inhibited by VPA (Chen et al., 1999) despite subsequent reports not finding this inhibition (Phiel et al., 2001; Hall et al., 2002). GSK-3β is still currently proposed to be involved in the therapeutic action of lithium in bipolar disorder treatment.

Although VPA is fast becoming the first-choice treatment for bipolar disorder worldwide—patients with newly diagnosed bipolar disorder are twice as likely to be prescribed VPA than lithium in the United States (Goodwin et al., 2003)—it also has some rare but severe side effects. These include teratogenicity (Nau et al., 1991), whereby mothers taking VPA during the first trimester of pregnancy have an increased chance of embryonic malformations (Loscher, 1999). Recent data suggest that this teratogenicity is caused by histone deacetylase inhibition (Phiel et al., 2001), giving rise to elevated histone acetylation and altered gene transcription. This effect has also been implicated in altering expression of the inositol biosynthetic enzyme ino1 (Kadow and Struhl, 1997), suggesting a possible link of teratogenic potency, histone deacetylase inhibition, and InsP$_3$ depletion. The mechanism by which VPA has been shown to increase viral load in HIV-positive patients (Jennings and Romanelli, 1999; Maggi and Halman, 2001) may also be mediated through its effects on promoter acetylation (Ylisastigui et al., 2004). Finally, an effect of VPA on GSK-3 activity, as seen for lithium (Stambolic et al., 1996), could also cause abnormal development. A correlation between the effect of both teratogenicity and viral amplification caused by VPA and its ability to cause InsP$_3$ depletion remains unknown. In this work, we used 10 compounds derived from the core structure of VPA to compare InsP$_3$, teratogenicity, viral amplification, and GSK-3β inhibitory effects, and we found that these adverse effects are discrete from that of InsP$_3$ depletion.

### Materials and Methods

#### Materials

All chemicals used were of analytical grade if not stated otherwise. Lithium chloride, VPA, myo-inositol, trichostatin A, dimethyl sulfoxide, and vigabatrin were supplied by Sigma Chemical (Poole, Dorset, UK). 2-Methyl-2-pentenoic acid (IX) was provided by Bachem UK Ltd. (St. Helens, Merseyside, UK). Valproic acid derivatives were synthesized according to methods described elsewhere (Hauck et al., 1991; Bojic et al., 1996, 1998; Levi et al., 1997; Gravemann, 2002). Standard gas chromatography-mass spectrometry purity analysis procedures demonstrate a chemical purity of the derivatives ≥95% and after suitable derivatization, an enantiomeric purity of ≥95% enantiomeric access of the chiral compounds. All VPA derivatives used in the in vitro experiments were dissolved in dimethyl sulfoxide to result in stock solutions of 1 M. PO activity in Dictyostelium discoideum was kindly supplied by Katwijk Chemie (Katwijk, The Netherlands). Prolyl oligopeptidase inhibitor Z-Pro-Pro-aldehyde-dimethyl acetal was provided by Bachem UK Ltd. (St. Helens, Merseyside, UK). Valproic acid derivatives were synthesized according to methods described elsewhere (Hauck et al., 1991; Bojic et al., 1996, 1998; Levi et al., 1997; Gravemann, 2002). Standard gas chromatography-mass spectrometry purity analysis procedures demonstrate a chemical purity of the derivatives ≥95% and after suitable derivatization, an enantiomeric purity of ≥95% enantiomeric access of the chiral compounds. All VPA derivatives used in the in vitro experiments were dissolved in dimethyl sulfoxide to result in stock solutions of 1 M. PO activity in Dictyostelium discoideum was kindly supplied by Katwijk Chemie (Katwijk, The Netherlands).

#### D. discoideum Cell Culture and InsP$_3$ Analysis

Wild-type D. discoideum cells (Ax2g) were grown for 20 h in axenic media at 1 × 10$^6$ cells/ml in the presence of drugs at indicated concentrations or with vehicle-only control dimethyl sulfoxide. Cells were washed and resuspended in 1 ml of phosphate buffer and aerated for 10 min in the presence of the drug. Afterward, InsP$_3$ levels were measured by isotope dilution as reported previously (Williams et al., 1999). Protein was measured by Bradford assay (Bio-Rad Laboratories, Hemel Hempstead, UK).

#### Teratogenic Potency Assay

The exencephaly rate as a model for teratogenic effects was measured within the NRMI-exencephaly mouse model (Nau et al., 1981) at one or more concentrations of the substances (Hauck et al., 1991; Bojic et al., 1996, 1998; Volland, 2002) before being transformed to the arbitrary scale of teratogenic potency grading of 0 indicating no teratogenic potency to ++++ for compounds showing very high teratogenic potency (Table 1).

#### Dorsal Root Ganglion Explant Culture

Dorsal root ganglia (DRG) neuron explants from E18 rat embryos were plated onto poly-L-lysine (20 µg/ml)–and laminin (20 µg/ml)-coated glass cover slips, and cultures were incubated for 24 h in Dulbecco’s modified Eagle’s medium/10% fetal calf serum/1% penicillin/streptomycin.

### Table 1

<table>
<thead>
<tr>
<th>Teratogenic Potency Grading</th>
<th>Dose Range</th>
<th>Exencephaly Rate</th>
<th>Description</th>
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<tr>
<td>0</td>
<td>&gt;3</td>
<td>0</td>
<td>No teratogenic potency</td>
</tr>
<tr>
<td>+</td>
<td>2–3</td>
<td>1–5</td>
<td>Low teratogenic potency</td>
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<tr>
<td>++</td>
<td>2–2</td>
<td>5–25</td>
<td>Lower teratogenic potency than VPA</td>
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<tr>
<td>+++</td>
<td>2–3</td>
<td>25–60</td>
<td>Equal teratogenic potency to VPA</td>
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<tr>
<td>++++</td>
<td>1–2</td>
<td>40–60</td>
<td>Higher teratogenic potency than VPA</td>
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<td>+++++</td>
<td>0.25–1</td>
<td>40–60</td>
<td>Very high teratogenic potency</td>
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supplemented with 20 ng/ml nerve growth factor in the presence of specified drugs before fixation in 4% paraformaldehyde. DRG explants were washed twice with PBS, permeabilized in PBS/1% Triton X-100, and blocked in PBS/0.5% Triton2% bovine serum albumin. Cultures were stained with Alexa595-conjugated phalloidin (Molecular Probes, Eugene, OR) and anti-tubulin antibody (Sigma Chemical). Growth-cone sizes were determined in SimplePCI (Compix Inc., Imaging Systems, Cranberry Township, PA), and statistical analysis was carried out using the Student’s t test.

GSK-3β Kinase Assay: GSK-3β-specific activity was determined by measuring the transfer of [γ-32P]ATP from [γ-32P]ATP to the GSK-specific peptide substrate GSM, as described previously (Ryves et al., 1998). The final concentration of each assay component was as follows: 40 mM Tris, pH 7.5, 12.5 mM MgCl₂, 2 mM dithiothreitol, 400 μM GSM, 100 μM ATP, and 40,000 cpm/μl [γ-32P]ATP. Phosphate incorporation was linear with up to 200 units of kinase per assay for at least 10 min at room temperature (1 unit = 1 picomole of phosphate transferred to GSM peptide in 10 min). All experiments used 25 to 50 units of activity, which produced 12 to 15,000 cpm per assay under these conditions. Assays were conducted in triplicate, and activity was expressed as a percentage of no vehicle dimethyl sulfoxide control, with error bars showing standard deviation.

HIV-1 Infection Assay. HIV-1 vectors encoding green fluorescent protein (GFP) were pseudotyped with the vesicular stomatitis G-envelope protein as described previously (Besnier et al., 2002). In brief, 293T cells were transfected with three plasmids: p8.91, encoding HIV-1 gag-pol (HIV-1 structural and enzymatic proteins); pC-SGW, encoding an HIV-1 RNA, including the GFP gene; and pMDG, encoding the vesicular stomatitis G envelope protein. Forty-eight hours later, supernatant containing HIV-1 particles was collected and used to infect TE671 cells (American Type Culture Collection, Manassas, VA) plated at 10⁵ cells/well in six-well plates in the presence or absence of drug. Forty-eight hours later, infected cells were enumerated by fluorescence-activated cell sorting (FACS) (BD Biosciences, Cowley, Oxford, UK), and the percentage of infections was determined. Infections in the presence and absence of drug were compared. Viral doses were chosen to infect between 0.5 and 5% of the target cells to ensure linearity of the assay.

Results

Preliminary Screening for InsP₃-Depleting VPA Analogs. We have used D. discoideum to examine the effect of a set of VPA analogs (denoted I to IX) on InsP₃ levels (Figs. 1 and 2). Analogs were chosen by broad category, including branched and nonbranched side chain, saturated and unsaturated derivatives, R- and S-enantiomeric pairs, and analogs with a derivatized carboxylic acid function like VPD and hydroxamates. We included both lithium and VPA as reference substances. Cells were exposed for 20 h to drugs at a concentration of 0.5 mM, which is within the therapeutic range found in patient plasma undergoing VPA treatment. InsP₃ levels were measured using a direct InsP₃ binding protein assay. The results of this experiment clearly show that both VPA and lithium lower InsP₃ levels (Fig. 2, A and B), and some VPA analogs also exhibit this effect. To our knowledge, this represents the first direct assay for screening InsP₃-depletion efficacy of potentially new bipolar disorder drugs.

Structural comparison of analogs causing a strong InsP₃ reduction significantly below lithium treatment (Fig. 2B, compounds I, III, VIII, and IX) shows that these drugs have variable main- and side-chain length and contain both saturated and unsaturated bonds. All active compounds contain an acid group, whereas VPA derivatives containing modified acid groups like the amide or the hydroxamic acid function (Fig. 2, A and B; compounds V, VI, and VII, and VPD) showed less InsP₃ reduction. It is also noteworthy that two pairs of enantiomers with acid function showed opposite effects on InsP₃ reduction (compare compounds I and II, III and IV) with both of the S-enantiomers being the more potent derivative, although the enantiomers of the corresponding hydroxamic acid (compounds V and VI) do not show this effect. These results are in accordance with Pfieffer’s rule, which states that the greater the difference in the pharmacological effect of two enantiomers, the greater is the specificity of the active isomer for the response of the system under test; our results suggest a stereoselective mode of receptor interaction of valproic acid derivatives for InsP₃ depletion.

Teratogenic Potency of VPA Derivatives. We measured the teratogenic potency of the VPA analogs derived from the NMRI-exencephaly mouse model (Nau et al., 1981) and transformed the exencephaly rates measured previously into an arbitrary scale of teratogenic potency rating from nonteratogenic (0) to highly teratogenic (+++++, in which VPA is considered intermediate (++++) (Table 1 and Fig. 2C). Data are not available in this scale for compound IX, although it is known to be nonteratogenic (Phiel et al., 2001). VPD has been suggested to be converted to VPA in vivo and therefore shows teratogenicity (Radatz et al., 1998), but it is highly likely that it does not possess an intrinsic teratogenic potency. Although some of the teratogenic structural requirements like carboxylic acid function and the distinction of enantiomers are common to the observed requirements for InsP₃ depletion, there seems to be no direct correlation between InsP₃ depletion and teratogenic potency (Fig. 2, B and C). These data therefore provide the opportunity to select VPA derivatives that deplete InsP₃ without the potency for teratogenic side effects. Because the teratogenic effects of valproic acid may be caused by the inhibition of histone
deacetylases (Gurvich et al., 2004) and taking into account the relatively small number of VPA derivatives in this first set of analogs, these results also suggest no correlation between histone deacetylase inhibition and InsP₃ depletion.

Defining VPA Analogs with a Common Mode of Action to Current Bipolar Disorder Treatments. The common increase in growth-cone size of primary rat DRG neurons has been suggested to be involved in the therapeutic effects of lithium, VPA, and carbamazepine (Williams et al., 2002). We therefore compared the effects of VPA, the four VPA analogs showing the most acute reduction in InsP₃ levels in *D. discoideum* (I, III, VIII, and IX), and one analog showing no InsP₃ depletion (VII) on the growth-cone size of rat DRG neurons. Cells were treated with these drugs for 1 day, fixed and stained with phalloidin, and growth-cone areas were measured. An indication of growth-cone morphology is shown (Fig. 3A) after staining with phalloidin and an anti-tubulin antibody. Both VPA and the four InsP₃-depleting drugs showed a significant 2-fold increase in growth-cone size (*p < 0.05*), consistent with earlier reports of an 81% increase caused by VPA (Williams et al., 2002) in these cells (Fig. 3B). No significant difference in growth-cone enlargement was seen between highly teratogenic (I and III) and non-teratogenic compounds (VIII and IX). Similar growth-cone enlargement effects were also seen using DRG neurons derived from mice or chick embryos (data not shown).

To confirm that these effects occurred through the modification of inositol-based signaling, cells were also treated in the presence of an inhibitor of the enzyme prolyl oligopeptidase (133 μM), whose inhibition leads to the elevation of intracellular InsP₃ levels and increased resistance to the effects of bipolar disorder treatments (Williams et al., 1999, 2002; Schulz et al., 2002) or myo-inositol (2 mM). The increase in growth-cone size caused by the VPA analogs was completely reversed by the addition of either PO inhibitor or myo-inositol (Fig. 3B), thus confirming that the effect of these drugs is through the modification of inositol-based signaling pathways within the mammalian growth cone. Therefore, these VPA analogs share the same mode of action seen with the three most commonly used bipolar disorder treatments: lithium, VPA, and carbamazepine.

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**Fig. 2.** Characterization of InsP₃-depletion efficacy and teratogenicity of VPA analogs and current bipolar disorder treatments. *D. discoideum* cells were treated with lithium, VPA, or compounds derived from the chemical structure of VPA (Fig. 1). A, cells were grown overnight in complete medium in the presence of VPA or one of its analogs at 0.5 mM, a concentration found in patient plasma undergoing VPA treatment, or with lithium at 10 mM. Changes in InsP₃ levels were subsequently measured by isotope dilution assay (Amersham Biosciences UK, Ltd., Little Chalfont, Buckinghamshire, UK; see Materials and Methods), and compared with vehicle-only (■) control or currently used bipolar disorder treatments (□). Results represent four experiments assayed in triplicate (± S.E.M.). B and C, comparison of relative efficacy of VPA congeners to cause InsP₃ depletion expressed as a percentage of that found for VPA or lithium (underline indicates increased InsP₃ levels). Compounds were tested for teratogenic rating in an in vivo model. Arbitrary scale of teratogenic ratings for these drugs, from not teratogenic (0) to highly teratogenic (+++ + + +), as described under Materials and Methods (*, inferred from Phiel et al., 2001; Gurvich et al., 2004). RM, racemic mixture; ND, not determined.

**Fig. 3.** Enlargement of growth cones from DRG neurons after treatment with InsP₃-depleting drugs. Rat DRGs were cultured for 24 h in the presence of 0.5 mM VPA, analogs showing InsP₃ depletion in *D. discoideum*, or one analog showing no effect on InsP₃ levels in *D. discoideum* (Fig. 2A), and afterward, cells were fixed and stained before growth-cone size quantification. A, representative growth cones are shown for vehicle-only control (top left), 0.5 mM VPA (top right), or VPA with myo-inositol (2 mM) (bottom left) or prolyl oligopeptidase inhibitor (133 μM) (bottom right). B, quantification of growth-cone size for five VPA derivatives and VPA after treatment (■) with the indicated drug or with the drug plus insitol (□) or PO inhibitor (□). Data represent two to four independent experiments containing approximately 25 growth cones per experiment (± S.E.M.); *, *p < 0.05.
Effect of VPA Analogs on GSK-3β Activity. The first published report on the inhibitory effect of VPA on GSK-3β activity showed direct inhibition at therapeutically relevant concentrations (Chen et al., 1999). Although this result was not found in subsequent reports (Phiel et al., 2001; Hall et al., 2002; Williams et al., 2002), it still remains possible that modification of VPA in vivo may lead to a GSK-3β inhibitory compound. To look for effects of the VPA congeners on GSK-3β activity, we directly assayed purified mammalian GSK-3β activity in the presence of VPA and its congeners at 3 mM (Fig. 4). No changes were observed in GSK-3β activity with VPA or any congener tested. These assays were carried out at optimal magnesium concentrations, which were still found to lead to direct GSK-3β inhibition in earlier experiments (Chen et al., 1999).

Effect of VPA Analogs on Human Immunodeficiency Virus Infection. Recent data analyzing the treatment of HIV-positive patients with bipolar disorder have been conflicting, with some data suggesting that VPA treatment might increase viral loads by an undetermined mechanism, leading to worsening disease (Jennings and Romanelli, 1999; Maggi and Halman, 2001), whereas other data suggest that VPA might be protective against neuronal AIDS symptoms (Dou et al., 2003). We have therefore examined a series of VPA congeners for effects on in vitro HIV-1 infection (Fig. 5). In concordance with previous reports (Jennings and Romanelli, 1999; Maggi and Halman, 2001), we found that VPA increased HIV-1 vector infectivity at high (3 mM) concentrations (Fig. 5, A and B) but had little effect at low drug concentration (0.5 mM). Three compounds (I, V, and VI) increased HIV-1 vector infectivity 2-fold at low concentrations. All three of these compounds contained a seven-carbon backbone and a three-carbon side chain with a terminal triple bond and either carboxylic or hydroxamic acid groups. All three caused significant cell death at high (3 mM) concentrations, as indicated by a large reduction in fold HIV infection, indicating cytotoxicity. Five of these analogs had no significant effect on HIV-1 infectivity in this assay (II, IV, VIII, and IX and VPD). This effect was not common to InsP₃-depleting drugs including lithium (Fig. 2A), was not caused by an inhibitor of GABA transaminase (VGB), and was only partially caused by histone deacetylase inhibition. Furthermore, we could not reproduce the elevated infectivity shown by VPA and related compounds using a range of trichostatin A concentration (Fig. 5C). Similar results were also found using murine leukemia virus (data not shown).

Discussion

We have examined the effect of a set of 10 VPA congeners on InsP₃ depletion using the cellular slime mold D. discoideum (Figs. 1 and 2). This enabled the first partial characterization of the structural requirements of compounds, derived from the core structure of VPA, to deplete InsP₃. We report four valproic acid derivatives that deplete InsP₃ more strongly than lithium (Fig. 2). These four compounds contained a carboxylic acid group, whereas the analogs with amide or hydroxamic acid function were less potent, although a recent report by Shaltiel et al. (2004) shows high levels of inositol-depleting activity of a carboxamide VPA derivative. The active compounds varied in side-chain length, composition, and degree of saturation, but it is noteworthy that two pairs of enantiomers showed different potency in InsP₃ depletion (compounds I and III, II and IV) with the corresponding S-enantiomer being more potent. This observation suggests a stereoselective mode of receptor interaction. Unlike teratogenic rating, a hydrogen on the second carbon is not necessary for InsP₃ depletion (compound IX). Although these results provide the first indication of the structural requirements for InsP₃-depletion efficacy, a much larger cohort of VPA congeners must be analyzed for a complete structural definition of efficacy.

Previous investigation of a variety of valproic acid derivatives in an exencephaly model of an NMRI mice strain (Nau et al., 1981; Spiegelstein et al., 2003) revealed that the intrinsic structural requirements for the teratogenic potency are the following: 1) a carboxylic acid group; 2) a hydrogen atom at the second carbon atom; and 3) a branching at the second carbon atom with two side chains containing at least three carbon atoms at each side chain. Furthermore, unsaturated derivatives with one double or triple bond are found to exhibit a higher teratogenic potency, as are R-enantiomers of an enantiomeric pair at the second carbon atom, suggesting a stereogenic mode of action for the teratogenic effects of valproic acid derivatives (Hauck et al., 1991; Bojic et al., 1996, 1998). Teratogenic potential of the currently analyzed drugs is consistent with these findings (Table 1).

Comparison of InsP₃ depletion efficacy with the teratogenic potency of the VPA congeners showed no relationship between these two effects, because S-2-pentyl-4-pentynoic acid (I) depleted InsP₃ (Figs. 2 and 3) and was highly teratogenic (Fig. 2C), whereas 2-ethyl-4-methyl-pentanoic acid (VIII) and 2-methyl-2-pentoic acid (IX) showed similar InsP₃ depletion effects but were not teratogenic (Fig. 2). These results suggest that it is possible to isolate VPA derivatives with putative bipolar disorder efficacy without teratogenic side effects, and because a correlation between histone deacetylase inhibition and the teratogenic potency of VPA derivatives has been suggested, these results infer that there is no correlation between histone deacetylase inhibition and InsP₃ depletion. It is interesting to note that O’Loinsigh et al. (2004) recently studied the enantiomeric forms of 2-pentyloxy-4-pentynoic acid (compounds I and II) and defined the R-enantiomer to show cognition enhancement in water maze tests, whereas the S-enantiomer showed antiproliferative and prodifferentiative effects. This suggests that these latter actions are involved in either teratogenic, viral replication, or InsP₃ depletion effects, whereas the R-enantiomer may function through other means.
It is possible that analysis of InsP$_3$-depleting drugs in a single model system, such as D. discoideum, will not yield results similar to those found in primary mammalian neurons. To examine this, we tested a non-InsP$_3$-depleting compound (VII) and all four compounds that strongly deplete InsP$_3$ (I, III, VIII, and IX) on rat DRG neurons. We found that all drugs which depleted InsP$_3$ in D. discoideum also caused an effect consistent with InsP$_3$ depletion in mammalian neurons (Williams et al., 2002). This effect, seen as the doubling of the growth-cone size, was not seen for a non-InsP$_3$ depletion compound (VII). These results suggest that this growth-cone enlargement effect, shared by the commonly used bipolar disorder treatments (Williams et al., 2002), can now be extended to defined VPA analogs. These results also confirm D. discoideum as a good model system for testing bipolar disorder drugs. The reversal of these effects using either myo-inositol or inhibitors of prolyl oligopeptidase is consistent with the drugs working through InsP$_3$ depletion, as shown for lithium, VPA, and carbamazepine, and that the teratogenic effect of VPA is independent of its InsP$_3$ depletion action.

The inhibition of GSK-3β by VPA remains a contentious issue, because the first report concerning this issue showed the direct inhibition of GSK-3β in vitro at physiological levels of VPA (Chen et al., 1999), but this result has yet to be confirmed (Phiel et al., 2001; Hall et al., 2002; Williams et al., 2002) and it is still currently considered to be a direct inhibitor of GSK-3β. Here we show that VPA does not cause a direct inhibition of GSK-3β. These results, however, do not exclude the possibility of modified VPA structures, produced through in vivo metabolic processes, playing a role in its action. Indeed, VPA has also been shown to be metabolized (Granneman et al., 1984) with structurally related products causing altered in vivo effects. Although we have not eliminated all structural changes possible by biotransformation, we have shown no direct inhibition of GSK-3β by any VPA-related compound tested. Subsequent to the first reported inhibition of GSK-3β by VPA (Chen et al., 1999), in vivo studies have suggested that VPA may function to mimic the inhibition of GSK-3β by elevating the expression of β-catenin, a GSK-3β target, which is degraded upon phosphorylation (Phiel et al., 2001). However, this effect was shown to correspond to VPA’s teratogenic action because of its histone deacetylase inhibitory effect. Because we have determined which of these congeners are teratogenic, the potential effects on GSK-3β can be eliminated by the choice of nonteratogenic VPA derivatives.

In addition to teratogenicity, we examined the possibility that the InsP$_3$ depletion effect may be related to an increase HIV-1 infectivity. HIV is widespread, with up to 46 million people infected, many of whom have developed AIDS. Recent data analyzing the treatment of HIV-positive patients with bipolar disorder have been conflicting. Some data suggest that VPA treatment might increase viral loads by an unidentified mechanism, leading to worsening disease (Jennings and Romanelli, 1999; Maggi and Halman, 2001), whereas other data suggest that VPA might be protective against neuronal AIDS symptoms (Dou et al., 2003). To examine this, we exposed cells to combined drug and HIV-1 vector for 48 h, after which GFP-expressing infected cells were enumerated by FACS, thus measuring the effect of a drug on the ability of HIV-1 to infect human cells in culture. The HIV-1 vectors are nonreplicative, and therefore this assay measures the effect of the drugs on the ability of HIV-1 to infect the target cells, reverse-transcribe its RNA to DNA, deliver its genome to the nucleus, and integrate it into the genome.
host chromosome. We therefore used this assay to measure the effects of bipolar disorder treatments and VPA analogs on HIV infectivity. In addition, we used an inhibitor of GABA transaminase (vigabatrin) to mimic the proposed antiepileptic mechanism of drug action (Loscher, 1999) and trichostatin A, which mimics the histone deacetylase inhibition caused by VPA (Phiel et al., 2001), to examine the effects of VPA in this assay.

In concordance with previous HIV data (Jennings and Romaneli, 1999; Maggi and Halman, 2001), we implicate VPA as being able to increase HIV-1 infectivity in vitro. No correlation was found between InsP3-depleting efficacy and HIV-1 infectivity, suggesting an unrelated mechanism. In support of this, no increase in viral load has been found in patients using other InsP3-depleting bipolar disorder treatments. Comparison of the teratogenic ratings of compounds with effects on HIV-1 infection shows that broadly, the more teratogenic compounds show the highest increase in HIV infectivity (compounds I, III, V, and VI and VPA) (Figs. 2C and 5B), as reported for other viral activities (Michaelis et al., 2004). This suggests that teratogenicity and viral infectivity may be linked, although only a small increase in HIV infectivity was found using the teratogenic inhibitor to histone deacetylase, trichostatin A. No significant increase in viral infectivity is produced by compounds VIII and IX or by an inhibitor to GABA transaminase (vigabatrin), suggesting that this effect is not mediated through altered GABA signaling.

We screened 10 VPA congeners for their ability to affect InsP3 depletion, to cause in vivo teratogenicity and in vitro effects on viral replication. We found no relationship between these VPA side effects and the InsP3-depletion efficacy. We also found no indication of direct GSK-3β inhibition by VPA or any of the tested congeners. Instead, we found some correlation between teratogenicity and effects on HIV replication. This approach has allowed the first preliminary definition of changes in VPA structure that do not cause a teratogenic mechanism of drug action (Loscher, 1999) and trichostatin A, which mimics the histone deacetylase inhibition caused by VPA (Phiel et al., 2001), to examine the effects of VPA in this assay.

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