Presynaptic Mechanism Underlying cAMP-Induced Synaptic Potentiation in Medial Prefrontal Cortex Pyramidal Neurons

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ABSTRACT

cAMP, a classic second messenger, has been proposed recently to participate in regulating prefrontal cortical cognitive functions, yet little is known about how it does so. In this study, we used forskolin, an adenylyl cyclase activator, to examine the effects of cAMP on excitatory synaptic transmission in the medial prefrontal cortex (mPFC) using whole-cell patch-clamp recordings from visually identified layer II–III or V pyramidal cells in vitro. We found that bath application of forskolin significantly increased the amplitude of excitatory postsynaptic currents (EPSCs) in a concentration- and age-dependent manner. This enhancement was completely abolished by coapplication of cAMP-dependent protein kinase (PKA) inhibitor and p42/p44 mitogen-activated protein kinase (MAPK) kinase inhibitor, but not application of either drug alone. The membrane-permeable cAMP analog adenosine 3',5'-cyclic monophosphorothioate, Sp-isomer, triethylammonium salt, or activation of β-adrenergic receptor by isoproterenol mimicked the effect of forskolin to potentiate EPSCs. However, neither exchange protein activated by cAMP (Epac) inhibitor breflidin A nor hyperpolarization and cyclic nucleotide-activated channel blocker 4-ethylphenylamino-1,2-dimethyl-6-methylaminopyrimidinium chloride (ZD7288) affected forskolin response. The augmentation of EPSCs by forskolin was accompanied by a reduction of the synaptic failure rate, coefficient of variation and paired-pulse ratio of EPSCs, and an increase in release probability and number of releasable synaptic vesicles. Forskolin also significantly increased the frequency of miniature EPSCs without altering their amplitude distribution. These results indicate that cAMP acts presynaptically to elicit a synaptic potentiation on the layer V pyramidal neurons of mPFC through converging activation of PKA and p42/p44 MAPK signaling pathways.

The prefrontal cortex (PFC) is believed to subserve a wide variety of cognitive processes, including working memory (Goldman-Rakic, 1995), selection and remembering of relevant stimuli (Rainer et al., 1998), and remembering of contextually relevant, cross-modal stimulus association (Fuster et al., 2000). In rodents, the medial aspect of PFC (mPFC), encompassing the infralimbic and prelimbic areas, is considered to be anatomically homologous to the human/primate dorsolateral PFC to execute these functions (Zahr et al., 1997; Birrell and Brown, 2000). Prefrontal dysfunction is commonly found in many psychiatric disorders, such as attention-deficient/hyperactivity disorder, post-traumatic stress disorder, the affective disorders, schizophrenia, and drug abuse (Heidbreder and Groenewegen, 2003).

cAMP is one of the best-studied second messengers. A substantial body of evidence implicates that cAMP activates cAMP-dependent protein kinase (PKA), thereby eliciting a long-lasting increase in transmitter release at many central nervous system synapses. The PFC is one of the brain areas with a high density of adenyl cyclase (EC 4.6.1.1) and cAMP is one of the best-studied second messengers. A classic example is cAMP-dependent modulation of transmitter release at the pupil-sphincter synapse (Averill et al., 1977). Nevertheless, the site of action of cAMP in the PFC has not been well characterized. Recent studies have shown that cAMP can modulate postsynaptic currents (EPSCs) in a variety of brain regions, including the hippocampus (Boulougouris et al., 1999), somatosensory cortex (Borland et al., 1997) and the locus coeruleus (Kimura et al., 1997). In contrast, effects of cAMP on presynaptic function have not been well characterized.

In this study, we used forskolin, an adenylyl cyclase activator, to examine the effects of cAMP on excitatory synaptic transmission in the mPFC using whole-cell patch-clamp recordings from visually identified layer II–III or V pyramidal cells in vitro. We found that bath application of forskolin significantly increased the amplitude of EPSCs in a concentration- and age-dependent manner. This enhancement was completely abolished by coapplication of the PKA inhibitor and the MAPK kinase inhibitor, but not application of either drug alone. The membrane-permeable cAMP analog adenosine 3',5'-cyclic monophosphorothioate, Sp-isomer, triethylammonium salt, or activation of β-adrenergic receptor by isoproterenol mimicked the effect of forskolin to potentiate EPSCs. However, neither exchange protein activated by cAMP (Epac) inhibitor breflidin A nor hyperpolarization and cyclic nucleotide-activated channel blocker 4-ethylphenylamino-1,2-dimethyl-6-methylaminopyrimidinium chloride (ZD7288) affected forskolin response. The augmentation of EPSCs by forskolin was accompanied by a reduction of the synaptic failure rate, coefficient of variation and paired-pulse ratio of EPSCs, and an increase in release probability and number of releasable synaptic vesicles. Forskolin also significantly increased the frequency of miniature EPSCs without altering their amplitude distribution. These results indicate that cAMP acts presynaptically to elicit a synaptic potentiation on the layer V pyramidal neurons of mPFC through converging activation of PKA and p42/p44 MAPK signaling pathways.

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cAMP-Dependent Synaptic Potentiation in the mPFC

Materials and Methods

Slice Preparation. All experiments were performed according to the guidelines laid down by the Institutional Animal Care and Use Committee of National Cheng Kung University (Tainan, Taiwan). The coronal brain slices containing the prelimbic area PFC (1.5–2.5 mm anterior to bregma) were prepared from 8- to 23-day-old male Sprague-Dawley rats for whole-cell, patch-clamp recordings using procedures described previously (Hirsch and Crepel, 1990). Most experiments (unless otherwise noted) were performed on 14- to 16-day-old rats. In brief, rats were killed by decapitation under halothane anesthesia, and coronal slices (200 μm) were prepared using a vibrating microtome (Leica VT1000S; Leica, Nussloch, Germany). The anterior cingulated cortex and the shoulder or frontal area 2 region of the frontal cortex (Paxinos and Watson, 1998) were used for recording. The slices were placed in a storage chamber of artificial CSF (ACSF) oxygenated with 95% O2/5% CO2 and kept at room temperature for at least 1 h before recording. The composition of the ACSF solution was 117 mM NaCl, 4.7 mM KCl, 2.5 mM CaCl2, 1.2 mM MgCl2, 25 mM NaHCO3, 1.2 mM Na2HPO4, and 11 mM glucose at pH 7.3 to 7.4 and equilibrated with 95% O2/5% CO2.

Electrophysiological Recordings. For whole-cell, patch-clamp recording, one slice was transferred to a recording chamber of standard design and fixed at the glass bottom of the chamber with a nylon grid on a platinum frame. The chamber consisted of a circular well of low volume (1–2 ml) and was perfused constantly at 32.0 ± 0.5°C with a speed of 2 to 3 ml/min. Visualized whole-cell patch-clamp recording of synaptically evoked EPSCs and miniature EPSCs (mEPSCs) was conducted using standard methods as described previously (Huang et al., 2002). The layer II–III or V pyramidal neurons were identified by their pyramidal shape, presence of a prominent apical dendrite, and distance from the pial surface with an upright microscope (Olympus BX50WI; Olympus, Tokyo, Japan) equipped with a water-immersion ×40 objective lens and a Nomarski condenser combined with infrared videomicroscopy. Patch pipettes were pulled from borosilicate capillary tubing and heat-polished. The electrode resistance was typically 3 to 6 MΩ. The composition of intracellular solution was 115 mM potassium glutamate, 20 mM KCl, 10 mM HEPES, 2 mM MgCl2, 0.5 mM EGTA, 3 mM Na2ATP, 0.3 mM Na3GTP, 5 mM QX-314, and sucrose to bring osmolarity to 290 to 295 mOsM and pH to 7.3.

After a high-resistance seal (>2 GΩ before breaking into whole-cell mode) was obtained, suction was applied lightly through the pipette to break through the membrane. The cell was then maintained at −70 mV for several minutes to allow diffusion of the internal solution into the cell body and dendrites. Recordings were made using an Axopatch 200B (Molecular Devices, Sunnyvale, CA) amplifier. Electrical signals were low-pass-filtered at 3 kHz, digitized at 10 kHz using a 12-bit analog-to-digital converter (Digidata 1200; Molecular Devices). An Intel Pentium-based computer with pCLAMP software (version 8.0; Molecular Devices) was used for online acquisition and offline analysis of the data. For measurement of synaptically evoked EPSCs, a bipolar stainless steel stimulating electrode was placed on layer II–III approximately 150 to 200 μm away from the apical dendrites of the recorded neurons (Fig. 1A) and the superfusate routinely contained bicuculline methiodide (10 μM) to block inhibitory synaptic responses. The strength of synaptic transmission was mostly quantified by measuring the initial rising slope of EPSC (2-ms period from its onset; picoampere per millisecond), which contains only a monosynaptic component. In some experiments, synaptic currents were recorded in the presence of NMDA receptor antagonist d-APV (50 μM) or AMPA/kainate receptor antagonist CNQX (20 μM), and the amplitude was measured over a 0.5- to 2-ms window concentrated around the peak. Series resistance (Rs) was calculated according to the equation Rs = 10 mV/I, where I was the peak of transient current (filtered with 10 kHz) evoked by the 10-nA testing pulse when the pipette capacitance was compensated fully. Only cells demonstrating <25 MΩ series resistance (usually 10–20 MΩ) were used in these experiments.

Miniature EPSCs were recorded from layer V pyramidal neurons held in voltage clamp at a potential of −70 mV in the presence of bicuculline methiodide (10 μM), tetrodotoxin (1 μM), and CdCl2 (100 μM) and analyzed offline using a commercially available software (Mini Analysis 4.3; Synaptosoft, Leonia, NJ). mEPSCs were abolished by CNQX (20 μM) plus d-APV (50 μM), indicating that these are glutamatergic events. The software detects events based on amplitudes exceeding a threshold set just above the baseline noise of the recording (3 pA). All detected events were re-examined and accepted or rejected based on subjective visual examination. The program then measured amplitudes and intervals between successive detected events. Background current noise was estimated from the baseline with no clear event and was subtracted from signals for analysis. The mEPSC frequencies were calculated by dividing the number of detected events by the total sampling periods of 5 to 10 min were analyzed for forskolin treatment. Events were ranked by amplitude and interevent interval for preparation of cu...
event intervals were made by performing a Kolmogorov-Smirnov t test. Amplitude histograms were binned in 1-pA intervals.

**Drug Application.** All drugs were applied by manually switching the superfusate. Drugs were diluted from stock solutions just before application. Forskolin, 1,9-dideoxy-forskolin (Dd-forskolin), KT5720, H-89, PD98059, U10126, SB203580, SP600125, and brefeldin A were dissolved in dimethyl sulfoxide stock solutions and stored at −20°C until the day of experiment. Other drugs used in this study were dissolved in distilled water. The concentration of dimethyl sulfoxide in the perfusion medium was 0.1%, which alone had no effect on basal synaptic transmission. Forskolin, isoprotanol, propranolol, 8-(4-chlorophenylthio)-2′-O-methyl-cAMP (8CPT-2Me-cAMP), adenosine 3′,5′-cyclic monophosphorothioate, S-isomer (Sp-cAMPS) triethylammonium salt, ZD7288, PD98059, U10126, SB203580, SP600125, CNQX, bicuculline methiodide, and D-APV were purchased from Sigma (St. Louis, MO); H-89 was purchased from Calbiochem (La Jolla, CA).

**Statistical Analysis.** The data for each experiment were normalized relative to baseline and are presented as means ± S.E.M. The numbers of experiments are indicated by N. The significance of the difference between the mean was calculated by paired or unpaired Student’s t test. Probability values (p) of less than 0.05 were considered to represent significant differences. Comparisons between control and experimental distributions of mEPSCs amplitude and inter-event intervals were made by performing a Kolmogorov-Smirnov test. Distributions were considered different using a conservative critical probability level of p < 0.01.

**Results**

A photograph of a slice used for recording is shown in Fig. 1A. The area used for recording is outlined by a circle. Figure 1B shows an infrared microscope image of a representative layer V pyramidal neuron. The results of the present study were obtained from 176 pyramidal neurons. These neurons had a mean resting membrane potential, spike height, and input resistance of −68.7 ± 0.8 mV, 93.7 ± 2.3 mV, and 286 ± 29 MΩ (n = 62), respectively, which are comparable with the values reported previously (Wang and O’Donnell, 2001). In all experiments, layer V pyramidal neurons were held at a holding potential of −70 mV, and EPSCs were evoked by the stimulation of layer II–III fibers with bipolar stimulating electrodes every 20 s in the presence of GABA<sub>A</sub> receptor antagonist bicuculline methiodide (10 μM). Because EPSCs were completely blocked by CNQX (20 μM) plus D-APV (50 μM) (Fig. 1C), they were predominantly mediated by ionotropic glutamate receptors.

**Potentiation of EPSCs by cAMP.** We initially examined the effect of the adenylyl cyclase activator, forskolin, on the evoked EPSCs. Typical responses are shown in Fig. 2A. Bath application of forskolin (25 μM) produced a rapid and sustained enhancement of evoked EPSCs on layer V pyramidal neurons. The mean EPSC slope measured 20 min after forskolin application was increased by 53.6 ± 5.8% of the control baseline (n = 8; p < 0.05, paired Student’s t test). No significant recovery was visible after drug washout of at least 20-min intervals. The magnitude of EPSC potentiation by forskolin was concentration-dependent (Fig. 2B), with an estimated EC<sub>50</sub> value of 21 μM. Because forskolin has been reported to possess many cAMP-independent actions, including the blockade of several types of K<sup>+</sup> conductance, it is possible that the effect of forskolin on EPSCs is caused by its nonspecificity (Laurenza et al., 1989). To exclude this possibility, an analog of forskolin, Dd-forskolin, which has no effect on adenylyl cyclase but does mimic many of the cAMP-independent actions of forskolin, was used. As shown in Fig. 2A, Dd-forskolin (25 μM) had no significant effect on EPSCs (4.2 ± 2.3%; n = 4; p > 0.05, paired Student’s t test). As on layer V pyramidal neurons, forskolin (25 μM) also increased the slope of EPSCs of layer II–III pyramidal neurons. The mean EPSC slope 20 min after forskolin application was increased by 48.7 ± 6.9% of the control baseline (n = 5; p < 0.05, paired Student’s t test) (Fig. 2C). Moreover, the facilitatory effect of forskolin on the layer V pyramidal neurons was robust at P8–P10 but became weaker as animal maturation (Fig. 2D), with the magnitude of potentiation by forskolin (25 μM) being 82.5 ± 7.8% at P8–P10 (n = 5), 53.6 ± 5.8% at P14–P16 (n = 10), and 32.8 ± 4.2% at P21–P23 (n = 5), indicating that the enhancement of synaptic transmission by forskolin is age-dependent. In the present study, forskolin was not observed to significantly change the holding current under voltage-clamp conditions (16.2 ± 2.2 pA for before and 19.3 ± 3.1 pA for 20 min after forskolin application; n = 55; p > 0.05, paired Student’s t test).

Additional evidence that elevation of cAMP levels can induce a synaptic potentiation of glutamatergic transmission on the layer V pyramidal neurons of mPFC came from exper-
iments using the nonhydrolyzable cAMP analog Sp-cAMPS. Similar to forskolin, bath application of Sp-cAMPS (100 μM) for 20 min produced a significant enhancement of EPSCs by 42.5 ± 5.7% of the control baseline (n = 5; p < 0.05, paired Student’s t test) (Fig. 3A).

**Forskolin Enhances Synaptic Transmission through Converging Activation of PKA and p42/p44 MAPK Signaling Pathways.** An established signal transduction pathway for forskolin action is the activation of adenylyl cyclase, leading to an increase in intracellular cAMP levels and activating PKA, which in turn modulates the function of a series of cellular substrates by increasing their phosphorylation state. We first examined whether the cAMP-dependent synaptic potentiation is mediated by PKA activation. To test this possibility, two structurally unrelated PKA inhibitors, KT5720 and H-89, were used. In the presence of either of KT5720 (1 μM) or H-89 (1 μM), forskolin was still able to potentiate EPSCs but the magnitude of potentiation was significantly smaller than that induced in the interleaved control condition (p < 0.05; unpaired Student’s t test). On average, EPSC slope measured 20 min after forskolin application was increased by 38.4 ± 4.2% (n = 8) in the presence of KT5720 and 36.6 ± 4.1% (n = 5) in the presence of H-89 (Fig. 3B). Neither KT5720 nor H-89 alone had effects on EPSCs (Supplemental Fig. S1A). These results suggest that in addition to the PKA signaling cascade, other signal mechanisms are necessary for the induction of synaptic potentiation by forskolin.

At the calyx of Held, an increase in cAMP levels in the nerve terminal can facilitate transmitter release via the activation of Epac pathway (Kaneko and Takahashi, 2004). We next examined whether the cAMP-dependent synaptic potentiation seen in this study is mediated by Epac activation. To test this possibility, we used a selective Epac agonist, 8CPT-2Me-cAMP, which has been shown to have little effect on PKA (Enserink et al., 2002). To identify the saturating concentration of 8CPT-2Me-cAMP for activating Epac by external bath application, we treated slices with varying concentrations of 8CPT-2Me-cAMP. As shown in Fig. 3C, 8CPT-2Me-cAMP induced a concentration-dependent potentiation of EPSCs, and the magnitudes of potentiation caused by 50 and 100 μM 8CPT-2Me-cAMP were not significantly different (Fig. 3C). Thus, 50 μM 8CPT-2Me-cAMP was chosen to examine whether forskolin- and 8CPT-2Me-cAMP-induced synaptic potentiation use a similar induction mechanism.

The approach used to address this question is to determine whether the induction of one form of potentiation reduces or occludes the induction of the other form of potentiation. As shown in Fig. 3D, after 8CPT-2Me-cAMP-induced potentiation was fully established, application of forskolin still caused synaptic potentiation. On average, EPSC slope measured 20 min after forskolin application was increased by 51.3 ± 4.6% (n = 5), which was not significantly different from that found in slices without receiving 8CPT-2Me-cAMP. Because the small G-protein antagonist brefeldin A has recently been shown to effectively antagonize 8CPT-2Me-cAMP’s action on synaptic transmission at the crayfish neuromuscular junctions (Zhong and Zucker, 2005), we then used brefeldin A to characterize the role of Epac activation in forskolin-induced synaptic potentiation. Brefeldin A alone (100 μM) had no effect on basal synaptic transmission and did not affect forskolin-induced EPSC potentiation (Fig. 3E). On average, EPSC slope measured 20 min after forskolin application was increased by 49.7 ± 6.5% (n = 5), which was not significantly different from that of potentiation elicited under control condition. These results suggest that activation of Epac is not necessary for forskolin-induced synaptic potentiation in the layer V pyramidal neurons of mPFC.

To clarify the involvement of HCN channels in the induction of forskolin-induced synaptic potentiation, we compared the magnitude of synaptic potentiation in control slices with that obtained in slices preincubated in HCN channel blocker ZD7288. As shown in Fig. 3E, ZD7288 (30 μM) did not affect forskolin-induced EPSC potentiation (Fig. 3E). On average, EPSC slope measured 20 min after forskolin application was increased by 48.4 ± 5.7% (n = 5), which was not significantly different from that of potentiation elicited under control condition. Thus, these results rule out an involvement of HCN channels in the forskolin response.

In a number of systems, cAMP-induced MAPK activation
independently of PKA has been reported (Iacovelli et al., 2001; Troadec et al., 2002). To determine whether the cAMP-dependent synaptic potentiation is mediated by MAPK activation, forskolin-induced synaptic potentiation was attempted in the presence of multiple types of MAPK inhibitors. As shown in Fig. 3F, forskolin response was strongly reduced by p42/p44 MAPK signaling pathway inhibitors, PD98059 and U0126. On average, EPSC slope measured 20 min after forskolin application was increased by 18.6 ± 3.5% in the presence of PD98059 (50 μM) (n = 8; p < 0.05 compared with forskolin alone, unpaired Student’s t test) and 15.3 ± 3.4% in the presence of U0126 (10 μM) (n = 6; p < 0.05 compared with forskolin alone, unpaired Student’s t test). Neither PD98059 nor U0126 alone had effects on EPSCs (Supplemental Fig. S1B). In contrast, inhibition of p38 MAPK signaling pathway with SB203580 (1 μM) or c-Jun N-terminal kinase inhibitor SP600125 (20 μM) failed to affect the forskolin-induced synaptic potentiation (SB203580, 56.3 ± 4.6%, n = 4; SP600125, 51.6 ± 3.9%, n = 4) (Fig. 3H). In another experiment, we found that forskolin failed to potentiate EPSCs when H-89 and PD98059 or H-89 and U0126 were applied together (H-89 + PD98059, 2.8 ± 1.5%, n = 5; H-89 + U0126, 2.4 ± 1.3%, n = 4) (Fig. 3, G and H), suggesting that cAMP elevated by forskolin activates both PKA and p42/p44 MAPK signaling pathways to induced synaptic potentiation.

Presynaptic Expression of Forskolin-Induced Synaptic Potentiation. To dissect the synaptic site of action for forskolin, the effects of forskolin on the AMPA and NMDA receptor-mediated component of synaptic currents were examined. If forskolin-induced synaptic potentiation were expressed presynaptically, changes in both of the magnitude of AMPA receptor-mediated EPSC (EPSCAMPA) and NMDA receptor-mediated EPSC (EPSC_NMDA) by forskolin would be expected. The EPSCAMPA was recorded from the layer V pyramidal neurons in the presence of NMDA receptor antagonist d-APV (50 μM) at a holding potential of −70 mV, and the EPSC_NMDA was recorded in the presence of AMPA/kainate receptor antagonist CNQX (20 μM) at a holding potential of +40 mV to remove the voltage-dependent block of Mg2+. As illustrated in Fig. 4, A and B, forskolin (25 μM) increased the amplitude of EPSCAMPA by 51.4 ± 5.6% (n = 5) of control baseline. Comparable results were obtained with EPSC_NMDA (48.9 ± 5.2% of control baseline, n = 5).

Fig. 3. Coapplication of PKA and p42/p44 MAPK inhibitors blocks the forskolin-induced EPSC potentiation. A, summary of experiments (n = 5) showing that bath application of nonhydrolyzable cAMP analog Sp-cAMPS increased EPSCs. B, summary of experiments showing that KT5720 (1 μM, n = 8; ○ or H-89 (1 μM, n = 5; ●) treatment partially blocked the forskolin-induced synaptic potentiation. C, summary of experiments showing that bath application of Epac agonist 8CPT-2Me-cAMP (10–100 μM) enhanced EPSCs in a concentration-dependent manner. Data are derived from four to six cells. D, summary of experiments (n = 5) showing that treatment of 8CPT-2Me-cAMP did not affect the forskolin-induced synaptic potentiation. E, summary of experiments showing that forskolin-induced synaptic potentiation was not significantly affected by prior treatment with either brefeldin A (BFA, 100 μM; n = 5; ○) or ZD7288 (30 μM, n = 5; ●). F, summary of experiments showing that PD98059 (50 μM, n = 8; ○) or U0126 (10 μM, n = 6; ●) treatment significant reduced the forskolin-induced synaptic potentiation. G, summary of experiments showing that forskolin-induced EPSC potentiation was completely abolished by prior coapplication of H-89 (1 μM) and PD98059 (50 μM) (n = 5, ○) or H-89 (1 μM) and U0126 (10 μM) (n = 4, ●). Sample traces are averages of five consecutive EPSCs recorded at time indicated by the numbers on the graph. H, histogram comparing the effects of different antagonists on the forskolin-induced synaptic potentiation. The magnitude of potentiation was measured 20 min after forskolin application. Data are taken from B, E, F, and G, *, significant difference from control group at p < 0.05.
To further test the possibility that forskolin induces synaptic potentiation through a presynaptic mechanism, we examined the effect of forskolin on the failure rate of single-fiber EPSCs evoked by minimal stimulation, which reflects changes in the presynaptic transmitter release (Stevens and Wang, 1994). As a typical example shown in Fig. 4C, the expression of forskolin-induced synaptic potentiation was accompanied by a decrease in synaptic failure rate. On average, the failure rate was decreased from 0.51 ± 0.03 to 0.18 ± 0.03 after forskolin application (n = 6; p < 0.05, paired Student’s t test) (Fig. 4D). We also addressed the synaptic locus of forskolin-induced synaptic potentiation by examining the trial-to-trial amplitude fluctuation in EPSCs with the variance analysis. The CV value varies with quantal content but is independent of changes in the postsynaptic response to a fixed amount of transmitter and is a useful measure of changes in presynaptic function (Bekkers and Stevens, 1990). Because variance analysis is best done on unitary synaptic responses, our strategy was to carry out a variance analysis of unitary single-fiber EPSCs evoked by minimal stimulation before and after forskolin application. We found that after the induction of synaptic potentiation by forskolin, the value of CV for unitary EPSCs was decreased from 0.85 ± 0.04 to 0.49 ± 0.03 (n = 6; p < 0.05, paired Student’s t test) (Fig. 4D).

When the excitatory afferents to the central neurons are activated twice with a short interval between each stimulus, the response to the second stimulus is generally facilitated in relation to the initial stimulus. This phenomenon is called paired-pulse facilitation and is attributed to an increase in the amount of transmitter release to the second stimulus (Zucker, 1989). On the other hand, the manipulation of presynaptic transmitter release may result in the change in the magnitude of paired-pulse facilitation. If the forskolin-induced synaptic potentiation involves a presynaptic mechanism of action, it will be associated with a decrease in the ratio of paired-pulse (PPR). To test this hypothesis, PPR (using an interpulse interval of 50 ms) was determined before and during application of 25 μM forskolin for 20 min. Under control conditions, the ratio of the slope of the second EPSC divided by the first one was 1.48 ± 0.04 (n = 6). We found that forskolin significantly decreased the PPR to 1.15 ± 0.05 (n = 6; p < 0.05, paired Student’s t test) (Fig. 4D), suggesting an increase in glutamate release probability after forskolin application.

Forskolin Enhances Frequency of mEPSCs. To further confirm the possibility that forskolin potentiates synaptic transmission through a presynaptic mechanism, we examined the effects of forskolin on mEPSCs in the presence of tetrodotoxin (1 μM) and CdCl2 (100 μM). mEPSCs in the layer V pyramidal neurons were measured under voltage clamp at −70 mV and were pharmacologically isolated from spontaneous inhibitory currents by the inclusion of 10 μM bicuculline methiodide in the ACSF perfusing the slices. The mEPSCs were totally blocked by bath coapplication of CNQX (20 μM) and d-APV (50 μM), confirming them to be true glutamate receptor-mediated events (data not shown). Under control conditions, mEPSCs had a mean amplitude of 5.98 ± 0.23 pA and a variable frequency ranging from 1.9 to 2.7 Hz (mean, 2.13 ± 0.19 Hz; n = 5). In five pyramidal neurons tested, forskolin (25 μM) markedly increased the mean frequency of the mEPSCs from 2.13 ± 0.19 to 5.23 ± 0.21 Hz (p < 0.05, paired Student’s t test) (Fig. 5, A and F). Significant differences in cumulative interevent interval distributions were observed in all five cells tested during forskolin application (i.e., forskolin shifted the interevent interval distribution of mEPSCs to shorter intervals; p < 0.01, Kolmogorov-Smirnov test). A typical example of recorded cell is shown in Fig. 5D. However, there was no significant effect of forskolin (25 μM) on the mEPSC amplitude. This can be observed by a lack of effect of forskolin on either the amplitude histogram (Fig. 5B) or the cumulative probability plots (Fig. 5C, p = 0.94; Kolmogorov-Smirnov test). The mean amplitude of mEPSCs recorded in the presence of forskolin (25 μM) was 6.21 ± 0.19 pA, which was of comparable amplitude with that of mEPSCs recorded under control conditions (5.98 ± 0.23 pA; p = 0.78, paired Student’s t test). Therefore, these data further suggest that forskolin may act presynaptically to enhance the amount of glutamate release without changing the postsynaptic sensitivity to glutamate.

Forskolin Increases the Number of Releasable Vesicles and Release Probability. Although the above results
are consistent with a presynaptic site for the expression of forskolin-induced synaptic potentiation, they may be attributed to a number of presynaptic mechanisms, including an increase in the number of readily releasable quanta (synaptic vesicles) \((N)\) or an increase in release probability \((P)\) (Trudeau et al., 1996; Kaneko and Takahashi, 2004). To determine which mechanism underlies the forskolin-induced synaptic potentiation, we first used the approach of high-frequency repetitive stimulation (20 stimuli at 100 Hz) (Schneggenburger et al., 1999), which induced a strong depression of EPSCs (Fig. 6A). In this approach, assuming that depression is largely caused by depletion of readily releasable quanta, \(N\) multiplied by mean quantal size \((q)\) can be estimated from zero time intercept of a line fitted to a cumulative amplitude plot of EPSCs (Fig. 6B), and \(P\) can be estimated from the first EPSC amplitude divided by \(Nq\). During stimulation at 100 Hz, EPSCs underwent a marked depression and reached a steady low level. Forskolin (25 \(\mu\)M) potentiated the first few EPSCs during a train of tetanic stimulation (Fig. 6A). Cumulative amplitude plot of EPSCs before and after forskolin application indicated that forskolin increased \(Nq\) by 23.2 ± 4.1\% \((n = 6; p < 0.05,\) paired Student’s \(t\) test) (Fig. 6C) and \(P\) by 55 ± 11\% \((n = 6; p < 0.05,\) paired Student’s \(t\) test) (Fig. 6D). Given that forskolin had no significant effect on \(q\) (Fig. 5E), these results suggest that forskolin increases both the number of readily releasable quanta and release probability.

Another way of assessing the release probability is to evaluate the speed of block of NMDA receptors by irreversible open channel blocker MK-801 (Rosenmund et al., 1993). Repeated activation of synapses in the presence of MK-801 results in a progressive decline in the amplitude of the NMDA receptor-mediated synaptic current, and the rate of decay depends on the probability of transmitter release at synapses. If the probability of transmitter release is higher, a larger proportion of postsynaptic NMDA receptors is blocked at any one time, resulting in a faster decline of EPSC\(_{\text{NMDA}}\) (Rosenmund et al., 1993). The EPSC\(_{\text{NMDA}}\) was recorded in the presence of CNQX (20 \(\mu\)M) and at a holding potential of +40 mV. After confirming a stable baseline at a basal stimulus frequency of 0.1 Hz, the stimulation was stopped, and MK-801 (40 \(\mu\)M) was applied. Stimulation was restarted 10 min later, and the EPSC\(_{\text{NMDA}}\) was recorded in the continuous presence of MK-801. From five such experiments, the time course of decline could be fitted by a double exponential function, with a fast time constant of 82 s and a slow time constant of 186 s. In the presence of forskolin (25 \(\mu\)M), the fast and slow time constants were 31 and 285 s, respectively (Fig. 7). Forskolin also apparently increased the proportion of the fast component. Because \(P\) is inversely proportional to the decay time constant in the MK-801 experiments (Rosenmund et al., 1993), these results further support the proposal that forskolin increases \(P\).

\section*{\(\beta\)-Adrenergic Receptor Agonist Isoproterenol Potentiates EPSCs.} The final test was to determine whether activation of receptors that are positively coupled to elevate cAMP can mimic forskolin to potentiate synaptic transmission on the layer V pyramidal neurons of mPFC. The \(\beta\)-ad-
benevolent receptor agonist isoproterenol has been shown to mimic the enhancement effect of cAMP elevation on synaptic transmission of many brain regions (Herrero and Sánchez-Prieto, 1996; Huang et al., 1996). Although the action of β-adrenergic receptors on the glutamatergic transmission of the mPFC region has not been established, β-adrenergic receptors have been shown to mediate many noradrenaline functions in the mPFC (Bing et al., 1992), and noradrenaline has been reported to facilitate the release of glutamate from presynaptic terminals that synapse onto layer V pyramidal neurons of mPFC (Marek and Aghajanian, 1999). Thus, we conducted a series of experiments to test the hypothesis that activation of β-adrenergic receptors would induce a cAMP-mediated synaptic potentiation. As shown in Fig. 8A, bath application of isoproterenol (15 μM) for 20 min induced EPSC potentiation by 43.5 ± 4.3% of the control baseline (n = 5; p < 0.05, paired Student’s t-test). The response to isoproterenol was completely blocked by propranolol (20 μM), a selective β-adrenergic receptor antagonist, suggesting that this effect is indeed mediated by the activation of β-adrenergic receptors (Fig. 8B). In addition, isoproterenol was still able to potentiate EPSCs in the presence of H-89 (1 μM), but the magnitude of potentiation was significantly smaller than that induced in the interleaved control condition (p < 0.05, unpaired Student’s t-test) (Fig. 8C). On average, EPSC slope measured 20 min after isoproterenol application was increased by 28.5 ± 3.5% (n = 5) in the presence of H-89. Coapplication of H-89 (1 μM) and PD98059 (50 μM) completely blocked isoproterenol-induced EPSC potentiation (n = 5; 2.5 ± 1.2% of preisoproterenol baseline) (Fig. 8D). These data are consistent with the hypothesis that the enhancement action of β-adrenergic receptor activation on glutamatergic transmission in the mPFC is mediated by both PKA and p42/p44 MAPK signaling cascades.

**Discussion**

We have, for the first time, systematically examined the role of cAMP elevation in the regulation of synaptic transmission on layer V pyramidal neurons of mPFC. Our results indicate that forskolin and membrane-permeable cAMP analogs induce synaptic potentiation through presynaptic mechanisms. Most importantly, we provide pharmacological evidence that cAMP acts on both PKA and p42/p44 MAPK signaling pathways to enhance transmitter release from presynaptic nerve terminal. Furthermore, activation of β-adrenergic receptor with isoproterenol mimics forskolin and elicits a cAMP-dependent synaptic potentiation.

**Presynaptic Locus of Expression of cAMP-Dependent Synaptic Potentiation.** Various approaches were taken to determine the site of action of forskolin in enhancing transmission on layer V pyramidal neurons of mPFC. Based on these experiments, it is likely that forskolin-induced synaptic potentiation is primarily of presynaptic origin. Three lines of evidence support this conclusion. First, forskolin increases equally the AMPA receptor- and NMDA receptor-mediated component of EPSCs (Fig. 4, A and B). Second, the increase in synaptic transmission by forskolin was accompanied by a decrease in the synaptic failure rate, magnitude of CV, and PPR (Fig. 4D), which are generally considered to indicate a presynaptic mode of drug action (Zucker, 1989; Bekkers and Stevens, 1990; Stevens and Wang, 1994). Third, forskolin significantly increased the frequency of mEPSCs but did not affect the amplitude of mEPSCs (Fig. 5). A change in the amplitude of mEPSCs has traditionally been interpreted as a postsynaptic modification, whereas a change in their frequency is typically associated with mechanisms that increase the probability of transmitter release. Thus, the lack of effect of forskolin on the amplitude of mEPSCs also implies that forskolin-induced synaptic potentiation is not mediated by a change in postsynaptic sensitivity to glutamate.

**Forskolin Increases the Number of Releasable Vesicles and Release Probability.** Using three independent approaches, the paired-pulse stimulation, high-frequency stimulation, and MK-801 protocols, we have shown that forskolin increases the release probability, P. The high-frequency stimulation protocol also indicates that forskolin increases the number of releasable vesicles, N. In cerebellar parallel-Purkinje cell synapses, the effect of forskolin has been attributed primarily to an increase in P (Chen and Regehr, 1997). At the calyx of Held, forskolin has also been...
proposed to facilitate transmitter release by increasing both \( P \) and \( N \) (Kaneko and Takahashi, 2004). cAMP also regulates release from dentate granule cells in the hippocampus by changing both \( P \) and \( N \) (Weisskopf et al., 1994). The fit of a double exponential function to the data in MK-801 experiments predicts vesicle populations having different release probability (Rosenmund et al., 1993). The finding that forskolin selectively accelerated the fast decay time constant and increased the relative proportion of the fast-decay component suggests that forskolin increases \( P \) and the proportion of vesicles with high \( P \) as reported previously (Kaneko and Takahashi, 2004).

**Molecular Mechanism of cAMP-Mediated Synaptic Potentiation.** Forskolin has been reported to possess many cAMP-independent actions, including the blockade of several types of potassium currents, which could result in prolongation of presynaptic action potentials and consequent increase in transmitter release (Hoshi et al., 1988). However, the cAMP-independent action of forskolin could be mimicked by its analog Dd-forskolin, which is unable to activate adenyl cyclase. In our experiments, Dd-forskolin had no significant effect on EPSCs (Fig. 2, A and C). Thus, the effect of forskolin is not caused by its nonspecificity. This idea was also supported by the finding that nonhydrolyzable cAMP analog Sp-cAMPS mimicked forskolin to potentiate EPSCs (Fig. 3A). Consistent with this idea, we have found that the activation of \( \beta \)-adrenergic receptors that are coupled to \( G_x \) proteins and activation of cAMP-dependent signaling pathways also elicit a cAMP-mediated synaptic potentiation (Fig. 8).

What is the molecular target of cAMP? Previous studies have shown that cAMP-dependent synaptic potentiation is mediated mainly by activating PKA in a variety of brain regions, including the hippocampus, amygdala, cerebellum, and striatum (Chavez-Noriega and Stevens, 1994; Huang et al., 1996, 2002; Salin et al., 1996; Chen and Regehr, 1997). In contrast, at the calyx of Held, presynaptic cAMP is proposed to facilitate synaptic transmission via activating Epac pathway (Kaneko and Takahashi, 2004). The cAMP-dependent potentiation of synaptic transmission at crayfish glutamatergic neuromuscular junctions is mediated by acting on Epac and HCN (Zhong and Zucker, 2005). However, we found that forskolin still caused synaptic potentiation when forskolin was applied in the presence of PKA, Epac, or HCN inhibitors. Furthermore, although the application of a selective Epac agonist 8CPT-2Me-cAMP potentiated EPSCs, it did not occlude the subsequent forskolin-induced synaptic potentiation. However, antagonists of PKA and p42/p44 MAPK each reduced forskolin-induced synaptic potentiation, and together they almost fully abolished the potentiation. Our findings that the effects of PKA and p42/p44 MAPK activation seem to be additive, suggesting the possibility that coincident activation of these two signaling pathway is required for the induction of cAMP-dependent synaptic potentiation on layer V pyramidal neurons of mPFC. The biological step downstream of PKA and p42/p44 MAPK responsible for the cAMP-induced synaptic potentiation remains to be determined.

**Fig. 7.** Forskolin accelerates the time course of blocking NMDA receptor-mediated EPSC (EPSC\(_{\text{NMDA}}\)) by MK-801. EPSC\(_{\text{NMDA}}\) was evoked at 0.1 Hz at a holding potential of +40 mV in the presence of MK-801 (40 μM). Data are derived from a different group of cells, one group in the presence of forskolin (25 μM; \( n = 5 \)) and the other in its absence (\( C, n = 5 \)). The numbers of sample traces (superimposed) indicate the sequence of stimulation. Ordinate indicates the amplitude of EPSC\(_{\text{NMDA}}\) normalized to the initial amplitude. Abscissa indicates the time after starting stimulation in the presence of MK-801. Mean relative amplitudes derived each from five cells were fitted with double exponential functions.

**Fig. 8.** \( \beta \)-Adrenergic receptor agonist isoproterenol enhances synaptic transmission. A, summary of experiments (\( n = 5 \)) showing that bath application isoproterenol (15 μM) for 20 min increased EPSCs. B, summary of experiments (\( n = 4 \)) showing that isoproterenol-induced EPSC potentiation was completely abolished by prior treatment with \( \beta \)-adrenergic receptor antagonist propanolol (20 μM). C, summary of experiments showing that H-89 (1 μM, \( n = 5 \)) treatment partially blocked the isoproterenol-induced synaptic potentiation. D, summary of experiments showing that isoproterenol-induced synaptic potentiation was completely abolished by prior coapplication of H-89 (1 μM) and PD98059 (50 μM, \( n = 5 \)). Sample traces are averages of five consecutive EPSCs recorded at the times indicated by the numbers on the graph.
Given that forskolin increases both N and P, the target of these kinases seems to be in both the vesicular trafficking mechanism and the exocytotic mechanism. Indeed, at many synapses, activation of PKA has been shown to phosphorylate one or more proteins, either associated with or part of the protein complex that is necessary for the exocytosis of synaptic vesicles and underlies synaptic facilitation (Nagy et al., 2004). Activation of p42/p44 MAPK was also reported to facilitate glutamate release from rat brain synaptosomes by phosphorylating the synaptic vesicle membrane protein synapsin I, thereby regulating its interaction with the actin cytoskeleton, leading to the recruitment of releasable synaptic vesicles from a distal pool (Jovanovic et al., 2000).

We were surprised to find that the facilitatory effect of forskolin decreased with postnatal development (Fig. 2D). This developmental decline of forskolin-induced synaptic potentiation is not unique to excitatory afferents to layer V pyramidal neurons of mPFC; it was also reported at the calyx of Held (Kaneko and Takahashi, 2004). Although the molecular mechanism underlying this phenomenon remains unclear, a developmental decrease in the molecular target downstream of PKA and/or p42/p44 MAPK seems to contribute to this phenomenon. Further work, involving the use of functional knockout of candidate proteins, is needed to assess this hypothesis.

In summary, cAMP acts presynaptically, via activating the PKA and p42/p44 MAPK signaling pathways, to induce synaptic potentiation on layer V pyramidal neurons of mPFC. This enhancement is a result of increase in both release probability and number of releasable vesicles. The finding that β-adrenergic receptor activation mimics the forskolin action provides a major advance in establishing a role for more physiologically relevant stimuli in eliciting such synaptic modification. Given the importance of mPFC for cognitive functions, our findings may provide novel pharmacological strategies to treat human cognitive deficits in the future.

References


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