Potent Inhibition of Platelet-Derived Growth Factor-Induced Responses in Vascular Smooth Muscle Cells by BMS-354825 (Dasatinib)

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ABSTRACT

Abnormal migration and proliferation of vascular smooth muscle cells (VSMCs) are key events in the pathogenesis of restenosis that undermine the long-term benefit of widely performed balloon angioplasty and stenting procedures. Platelet-derived growth factor (PDGF) is a potent mitogen and motogen for VSMCs and is known to play a prominent role in the intimal accumulation of smooth muscle cells. In this study, we analyzed the effects of a novel protein tyrosine kinase inhibitor, BMS-354825 (dasatinib), on PDGF-stimulated VSMCs. BMS-354825 is an orally bioavailable dual Src/Bcr-Abl tyrosine kinase inhibitor currently undergoing clinical trials in cancer patients. We found that BMS-354825 inhibited PDGF-stimulated activation of PDGF receptor (PDGFR), STAT3, Akt, and Erk2 in rat A10 VSMCs and in primary cultures of human aortic smooth muscle cells (AoSMCs) at low nanomolar concentrations. The 50% inhibition of the PDGFRβ tyrosine kinase activity in vitro by BMS-354825 was observed at 4 nM. Direct comparison of BMS-354825 and another PDGFR inhibitor, imatinib (Gleevec, STI571), in VSMCs indicated that BMS-354825 is 67-fold more potent than imatinib in inhibition of PDGFR activation. BMS-354825 also inhibited Src tyrosine kinase in A10 cells. At the cell level, PDGF stimulated migration and proliferation of A10 cells and human AoSMCs, both of which were inhibited by BMS-354825 in a concentration dependent manner in the low nanomolar range. These results suggest that BMS-354825 is a potent inhibitor of PDGF-stimulated VSMC activities and a potential agent for the development of a new therapy for vascular obstructive diseases such as restenosis.

Balloon angioplasty and stenting are widely used procedures for coronary artery disease. Unfortunately, renarrowing (restenosis) of the dilated artery occurs in 25 to 40% of patients within 6 months after these procedures, which requires repeat angioplasty or bypass surgery (Dangas and Kuepper, 2002; Michaels and Chatterjee, 2002). Therefore, restenosis represents a major problem limiting the long-term efficacy of these revascularization therapies. Moreover, after coronary artery bypass surgery, narrowing (stenosis) of the transplanted artery can also occur that necessitates further medical treatments.

Abnormal migration and proliferation of VSMCs are critical events in the pathogenesis of artery obstructive diseases such as restenosis (Bailey, 2002; Sanz-Gonzalez et al., 2004; Levitzki, 2005). Although several growth factors and cytokines are involved in the development of restenosis, many lines of evidence have indicated that PDGF plays a prominent role in the pathogenesis of restenosis. PDGF is the most potent mitogen and motogen for VSMCs (Heldin and Westermark, 1999). PDGF is present at sites of vascular injury from activated platelets, monocytes, and cells of the artery wall (Raines, 2004). Expression of exogenous PDGF in animal arteries can induce intimal thickening through stimulation of VSMC proliferation and migration and synthesis of ABBREVIATIONS: VSMC, vascular smooth muscle cell; PDGF, platelet-derived growth factor; CML, chronic myeloid leukemia; PDGFR, platelet-derived growth factor receptor; STAT, signal transducer and activator of transcription; Erk, extracellular signal-regulated kinase; IGF-1R, insulin-like growth factor-1 receptor; EGFR, epidermal growth factor receptor; BMS-354825, N-(2-chloro-6-methylphenyl)-2-[6-(4-(2-hydroxyethyl)-1-piperazinyl)-2-methyl-4-pyrimidinyl][amin]-5-thiazolecarboxamide monohydrate; DMEM, Dulbecco’s modified Eagle’s medium; FBS, fetal bovine serum; AoSMC, aortic smooth muscle cell; PBS, phosphate-buffered saline; BSA, bovine serum albumin; DMSO, dimethyl sulfoxide; SMC, smooth muscle cells; AG-1295, 6,7-dimethyl-2-phenylquinoxaline; PD180970, 6-(2,6-dichloro-phenyl)-2-(4-fluoro-3-methyl-phenylamino)-8-methyl-6H-furo[2,3-d]pyrimidin-7-one.
smooth muscle cells (AoSMCs) was obtained from Cambrex Bio Science Walkersville, Inc. (Walkersville, MD) and grown in SmgM-2 Bulletkit medium (Cambrex) plus 100 units/ml penicillin and 100 μg/ml streptomycin. Human AoSMCs were used before the 7th passage. Cells were maintained at 37°C in a humidified 95% air and 5% CO2 incubator.

**Immunoprecipitation and Immunoblotting.** Cells were lysed in ice-cold lysis buffer (25 mM Tris-HCl, pH 7.2, 150 mM NaCl, 25 mM NaF, 1 mM benzamidine, 1% Triton X-100, 1 mM Na3VO4, 20 mM p-nitrophenyl phosphate, 2 μg/ml leupeptin, 2 μg/ml aprotinin, and 100 μg/ml phenylmethylsulfonyl fluoride). Cell lysate supernatants were obtained by microcentrifugation at 4°C for 15 min and the pellet concentration was determined. For immunoblotting analysis of cell lysates, cell lysate supernatants were heat-denatured in SDS loading buffer. Antibodies to activated STAT3, activated Akt, activated Erk1/2, and phosphotyrosine were used to assess the activation of these molecules and receptor tyrosine kinases by immunoblotting. Immunoblotting was performed as described previously (Cunnick et al., 2001, 2002; Ren and Wu, 2003) using the SuperSignal West Pico chemiluminescent reagent (Pierce, Rockford, IL). Each immunoprecipitation was performed using 600 μg of cell lysate supernatant, 2 μg of specific antibody indicated in the figure legends, and 30 μl of Protein-A or Protein-G at 4°C for 2 h. Immune complex was collected by microcentrifugation and washed three times with the lysis buffer. Quantification of immunoreactive band intensity was achieved using the ImageQuant program (GE Healthcare, Little Chalfont, Buckinghamshire, UK).

**Protein Tyrosine Kinase Assays.** PDGFR was immunoprecipitated with an anti-PDGFR antibody (Calbiochem). Immunoprecipitates were washed twice with the lysis buffer and twice with kinase buffer (50 mM HEPES, pH 7.4, 10 mM MnCl2, and 0.1 mM Na3VO4). The immune complex kinase assay was performed in a 50-μl reaction mixture (kinase buffer plus 20 μl of [γ-32P]ATP, and 10 μM ATP) at 30°C for 15 min. Kinase reaction was terminated by addition of 17 μl of 4× SDS loading buffer and heat-denaturation at 95°C for 10 min. Samples were subjected to 8% SDS-polyacrylamide gel electrophoresis, transferred to nitrocellulose membrane. Phosphorylation was quantified by filmless autoradiography analysis or autoradiography. The immune complex kinase assay for c-Src was performed as described previously (Ren et al., 2004).

**Cell Migration Assay.** Cell migration was measured using the Transwell cell migration assay (Ren et al., 2004). Transwell cell culture insert polycarbonate membrane (6.5 mm, 8.0-μm pore size; Costar, Corning, NY) was coated with rat tail type I collagen (10 μg/ml in PBS) at 4°C for 18 h and air-dried. VSMCs (80% confluent) were serum-starved in DMEM/0.1% BSA for 18 h, detached from plates by digestion with 1:3 diluted trypsin-EDTA (Invitrogen), washed with DMEM, and resuspended in DMEM/0.1% BSA at 5 × 104 cells/ml for A10 cells or 2.5 × 104 cells/ml for human AoSMCs. Cell suspension (0.2 ml per well) was incubated with BMS-354825 or solvent (DMSO) in 1.5-ml microcentrifuge tube for 20 min before been placed in the upper chamber of Transwell. The lower chamber contained 0.6 ml of DMEM/0.1% BSA with or without 5 ng/ml PDGF-BB (PeproTech) and BMS-354825. After incubation at 37°C/5% CO2 for 4 h, cells remaining on the upper membrane surface were mechanically removed with a cotton swab. Migrated cells on the lower side of membrane were fixed and stained with the HEMA3 reagents (Fisher Scientific, Swaneey, GA) and enumerated under a microscope in eight randomly chosen fields with a 10 × 10 lens. Each field for quantification of the migrated cell number has an area of 0.8 × 0.6 mm.

**Cell Proliferation Analysis.** A10 cells and human AoSMC cells were seeded in triplicate at 2 × 104 per plate in 6-cm plates in DMEM/1% FBS with or without PDGF-BB (10 ng/ml). After 24 h (day 0), cells were treated with DMSO (solvent) or 10 to 50 μM BMS-354825. Medium was changed on day 3. Viable cell number was determined on day 6 as described previously (Dorsey et al., 2000).
Flow Cytometric Analysis. A10 cells (80% confluent) were serum starved in DMEM/0.1% BSA for 24 h. After which, PDGF-BB (10 ng/ml) and BMS-354825 or DMSO was added and the incubation was continued for another 16 h. Cells were collected by trypsinization and resuspended in PBS at 5 × 10⁶ cells/ml. Cells were fixed in cold 70% ethanol, washed with PBS, and incubated with propidium iodide (20 µg/ml) and RNase (200 µg/ml) for 1 h at room temperature at a cell concentration of 1 × 10⁶ cells/ml. Flow cytometric analysis of cell cycle phase distribution was performed using a Becton Dickinson FACSCalibur flow cytometer (Becton, Dickinson, San Jose, CA), and 1 × 10⁶ events were recorded for each sample.

**Results**

**BMS-354825 Inhibits PDGF-Activated Signaling Pathways in VSMCs.** To determine whether BMS-354825 could inhibit PDGFR activation in VSMCs, we treated serum-starved rat A10 cells with PDGF-BB (5 ng/ml) for 0 to 30 min in the presence or absence of 50 nM BMS-354825. Cell lysates were analyzed for activation of PDGFRβ, STAT3, Akt, and Erk2 by immunoblotting analyses (Fig. 1A). PDGFRβ is the predominant PDGF subunit in VSMCs (Raines, 2004) and the only functional PDGFR subunit in A10 cells (Rao et al., 1997). STAT3, Akt, and Erk2 signaling pathways have been reported to mediate PDGF-induced cell migration and proliferation in VSMCs (Graf et al., 1997; Shibata et al., 2003; Zhan et al., 2003; Neeli et al., 2004; Kim et al., 2005; Vantler et al., 2005). PDGF treatment markedly increased PDGFRβ tyrosine phosphorylation, which was detectable at the earliest time point examined (1 min). BMS-354825 (50 nM) effectively blocked PDGF-stimulated PDGFRβ tyrosine phosphorylation at all time points examined. Figure 1A also shows that STAT3, Akt, and Erk2 were activated by PDGF in A10 cells as measured by activation-specific phosphorylation and these were inhibited by BMS-354825.

We next immunoprecipitated PDGFRβ from A10 cells that had been treated with PDGF-BB (5 ng/ml, 5 min) in the presence of 0 to 25 nM BMS-354825 and then analyzed the PDGFR tyrosine phosphorylation by immunoblotting. As illustrated in Fig. 1B, tyrosine phosphorylation of PDGFR was induced by PDGF stimulation in A10 cells; the response was inhibited by BMS-354825 with an IC₅₀ of 3 nM in this assay. To confirm that BMS-354825 inhibits PDGFR tyrosine kinase activity, PDGFRβ was immunoprecipitated from A10 cells with or without prior PDGF stimulation (5 ng/ml, 5 min). The PDGFR tyrosine kinase activity was determined in vitro in the immune complexes by autophosphorylation in the presence of various concentrations of BMS-354825. As shown in Fig. 1C, BMS-354825 inhibited PDGFR tyrosine kinase activity in a concentration-dependent manner. The 50% inhibition of PDGFRβ phosphorylation was observed at 4 nM BMS-354825.

**Comparison of PDGFR Inhibition by BMS-354825 and Imatinib.** In addition to inhibiting Bcr-Abl and c-Kit, imatinib also inhibits PDGFR tyrosine kinase. To compare the inhibition of PDGFR in VSMCs by BMS-354825 and imatinib, A10 cells were preincubated with various concentrations of BMS-354825 or imatinib, stimulated with PDGF-BB, and activation of PDGFR, STAT3, Akt, and Erk2 were analyzed. Figure 2A shows that PDGF-stimulated PDGFRβ tyrosine phosphorylation was completely blocked by 50 nM BMS-354825. In contrast, 5 µM imatinib was required to achieve the same level of PDGFR inhibition in parallel experiments. Likewise, complete inhibition of PDGF-stimulated Akt and Erk2 activation were observed in cells treated with 50 nM BMS-354825 but only with 5 µM imatinib, whereas decrease in STAT3 tyrosine phosphorylation appeared at lower concentrations of imatinib (Fig. 2A). It is noteworthy that inhibition of Akt by both BMS-354825 and imatinib was observed only at the highest drug concentration tested; PDGFR is a strong activator of the phosphoinositide-3 kinase signaling pathway because it binds phosphoinositide-3 kinase directly. Comparison of 50% inhibition of
PDGFR tyrosine phosphorylation by BMS-354825 and imatinib in these experiments indicated that BMS-354825 was 67-fold more potent than imatinib in inhibiting PDGFR tyrosine phosphorylation (Fig. 2B).

**Inhibition of PDGFR in Human AoSMCs by BMS-354825.** To exclude the possibility that the potent inhibition of PDGFR by BMS-354825 is specific to rat VSMCs, we examined the effect of BMS-354825 in primary cultures of human AoSMCs. Human AoSMCs were preincubated with 0 to 50 nM BMS-354825 and then stimulated with PDGF-BB (5 ng/ml, 5 min). PDGFR tyrosine phosphorylation and activation of STAT3, Akt, and Erk2 were analyzed. PDGF markedly induced PDGFR tyrosine phosphorylation in human AoSMCs (Fig. 3). As observed in A10 cells, the PDGF-stimulated PDGFR tyrosine phosphorylation was completely blocked by 50 nM BMS-354825.

The primary culture of human AoSMCs seemed to have an elevated level of active STAT3 in the absence of PDGF stimulation (Fig. 3). Nevertheless, STAT3 was further activated by PDGF, which was blocked by BMS-354825. BMS-354825, however, was unable to reduce the active STAT3 to a level below the basal activation state in human AoSMCs. Complete inhibition of PDGF-stimulated Akt and Erk2 activation was achieved at 50 nM BMS-354825 in human AoSMCs (Fig. 3), which is similar to data obtained in A10 cells (Fig. 2).

**Inhibition of c-Src Tyrosine Kinase by BMS-354825 in VSMCs.** To determine whether BMS-354825 inhibits c-Src activity in VSMCs, c-Src was immunoprecipitated from serum-starved A10 cells treated with PDGF-BB (5 ng/ml) for 0 to 30 min in the presence or absence of 50 nM BMS-354825, and the Src tyrosine activity was determined by an immune complex kinase assay using a GST fusion protein of Gab1 fragment as an exogenous substrate (Ren et al., 2004). c-Src isolated from A10 cells without exposure to BMS-354825 and PDGF had detectable kinase activity (Fig. 4A). This kinase activity was reduced to a residual level in c-Src isolated from BMS-354825-treated cells (Fig. 4A). PDGF treatment did not lead to a detectable increase in c-Src kinase activity in A10 cells in repeated experiments. Nevertheless, as in unstimulated cells, the activity of c-Src tyrosine kinase in PDGF-stimulated A10 cells was suppressed by BMS-354825 to the residual level (Fig. 4A). This result indicated that c-Src was basally active in A10 cells and that BMS-354825 effectively inhibited the c-Src kinase activity in these cells.

We next treated A10 cells with various concentrations of BMS-354825, immunoprecipitated c-Src and assayed its tyrosine kinase activity to determine the concentration-dependent effect of BMS-354825 on c-Src in A10 cells. Figure 2B shows that BMS-354825 potently inhibited c-Src tyrosine kinase in A10 cells. The IC50 for inhibition of c-Src autophosphorylation was 2.25 nM; the IC50 for inhibition of the exogenous substrate (GST-Gab1CT) was 2.0 nM.

**IGF-1R and EGFR in A10 Cells Are Not Sensitive to BMS-354825 Inhibition.** Besides PDGFR, A10 cells also express IGF-1R and EGFR. To assess whether BMS-354825 inhibits other receptor tyrosine kinases in VSMCs, we examined the effects of BMS-354825 on these two receptor tyrosine kinases in A10 cells. As shown in Fig. 5, IGF-1 induced IGF-1R tyrosine phosphorylation, whereas EGF induced EGFR tyrosine phosphorylation in A10 cells. No detectable inhibition of IGF-1R and EGFR tyrosine phosphorylation was observed when A10 cells were treated with up to 50 nM BMS-354825. These data are consistent with a previous report that the IC50s for inhibition of IGF-1R and EGFR by BMS-354825 in the in vitro kinase assays were >5 μM and 180 nM, respectively (Lombardo et al., 2004).

**BMS-354825 Inhibits PDGF-Stimulated VSMC Migration.** Migration of VSMCs plays a critical role in the development of restenosis. PDGF is a potent migratory stimulus for VSMCs. Transwell cell migration assay was performed to determine the effect of BMS-354825 on PDGF-induced VSMC migration. A10 cells had a low basal

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**Fig. 2.** Comparison of the inhibitory effects of BMS-354825 and imatinib in A10 cells. A, A10 cells were serum-starved, preincubated with indicated concentrations of BMS-354825 or imatinib for 20 min, and then stimulated with PDGF-BB (5 ng/ml, 5 min). Cell lysate supernatants were analyzed by immunoblotting similar to that described in Fig. 1A legend. B, quantification of PDGFR pTyr signal intensities. Data were from two experiments.

**Fig. 3.** Inhibitory effects of BMS-354825 in primary culture of human AoSMCs. Human AoSMCs were deprived from serum for 18 h, preincubated with indicated concentrations of BMS-354825 for 20 min, and then stimulated with PDGF-BB (5 ng/ml, 5 min). Cell lysate supernatants were analyzed for activation of PDGFR, STAT3, Akt, and Erk2 by immunoblotting.
migration activity. PDGF (5 ng/ml) stimulated A10 cell migration 7-fold in our assay. This response was inhibited by BMS-354825 in a concentration-dependent manner (Fig. 6, A and B). Consistent with the biochemical data, complete inhibition of PDGF-stimulated A10 cells migration was observed at 50 nM BMS-354825.

The primary human AoSMCs had a higher basal migration activity, which is 4 times that of A10 cells. In the presence of PDGF (5 ng/ml), migration of human AoSMCs was increased 3-fold. Again, PDGF-stimulated human AoSMC cell migration was inhibited by BMS-354825 (Fig. 6C).

**Inhibition of VSMC Proliferation by BMS-354825.** To determine the effect of BMS-354825 on PDGF-stimulated VSMC proliferation, rat A10 cells were incubated in medium containing low serum (1% FBS), PDGF (10 ng/ml), and/or 0 to 50 nM BMS-354825 for 6 days and viable cell numbers were determined. As shown in Fig. 7A, A10 cell proliferation was stimulated 5-fold by PDGF. BMS-354825 effectively inhibited PDGF-stimulated A10 cell proliferation. Complete inhibition was achieved at 50 nM BMS-354825. Similar results were obtained in human AoSMCs (Fig. 7B).

We next analyzed the effect of BMS-354825 on PDGF-induced change in cell cycle phase distribution. Figure 6C shows that PDGF decreased the percentage of cells in G1 phase from 88.65 to 65.44%, whereas it increased the percentages of cells in S and G2 phases from 10.4 and 0.99% to 23.83 and 10.74%, respectively. The effect of PDGF was suppressed by BMS-354825, such that in the percentages of cells in G1, S, and G2 phases were 88.29, 7.99, and 3.75%, respectively, in cells treated with PDGF in the presence of 50 nM BMS-354825 (Fig. 7C).

A previous study showed that the PDGFR specific tyrosine kinase inhibitor AG-1295 could inhibit porcine SMC proliferation, and the effect was reversible and not toxic (Banai et al., 1998). To evaluate whether the inhibitory effect of BMS-354825 is also reversible and not toxic, we performed experiments to determine the reversibility of the inhibitory effect of BMS-354825 on PDGF-stimulated cell proliferation.

**Fig. 4.** Inhibition of Src tyrosine kinase by BMS-354825. A, c-Src was immunoprecipitated from serum-starved A10 cells (0) or A10 cells stimulated with PDGF (5 ng/ml) for the indicated time in the presence of absence of 50 nM BMS-354825. Src kinase activity was determined by autophosphorylation and phosphorylation of GST-Gab1CT in the immune complex kinase assay (Ren et al., 2004). After autoradiography, the filter was used for immunoblotting analysis with an anti-Src antibody (bottom). B, c-Src was immunoprecipitated from serum-starved A10 cells treated with the indicated concentrations of BMS-354825 for 20 min and Src kinase activity was determined. C, quantification of radioactive signal from c-Src autophosphorylation and phosphorylation of GST-Gab1CT. The data were from two experiments performed in duplicate (n = 4).

**Fig. 5.** IGF-1R and EGFR are insensitive to BMS-354825. A10 cells were serum-starved, pretreated with indicated concentrations of BMS-354825 (20 min), and then stimulated with IGF-1 (10 ng/ml, 10 min) or EGF (10 ng/ml, 10 min). IGF-1R or EGFR was immunoprecipitated from cell lysate supernatants and analyzed by antibodies to pTyr and IGF-1R (A), or pTyr and EGFR (B).

**Fig. 6.** Effect of BMS-354825 on PDGF-stimulated cell migration. Transwell cell migration assay was performed using PDGF-BB (5 ng/ml) as the chemoattractant in the presence or absence of BMS-354825 as indicated. A, for A10 cells, 1 × 10⁴ cells were loaded into the upper chamber of each well. Each field was 0.8 × 0.6 mm. The data were from two (lanes 2–4) to four experiments performed in duplicate. B, representative views of migrated cells from each sample. Different microscopes were used to acquire data for A and B. Each area in these photos is 1.0 × 0.8 mm. C, for human AoSMCs, 0.5 × 10⁴ cells per well were used. The results were from two experiments performed in duplicate.
BMS-354825 is an orally bioavailable protein tyrosine kinase inhibitor currently in human clinical trials as a dual Src/Bcr-Abl inhibitor. We illustrate here that BMS-354825 is a potent inhibitor of PDGF-stimulated cellular responses in VSMCs. It is possible that inhibition of Src tyrosine kinase could contribute to the inhibitory effects of BMS-354825 in VSMCs, such as the STAT3 pathway (Wang et al., 2000; Bowman et al., 2001). However, biochemical evidence suggests that inhibition of PDGFR plays a major role in suppression of PDGF-stimulated responses in VSMCs by BMS-354825.

First, our data show that BMS-354825 is a potent inhibitor of PDGFR. The IC_{50} for inhibition of PDGFR\(\gamma\) tyrosine kinase in PDGF-stimulated A10 cells is 3 nM. A similar IC_{50} (4 nM) was obtained in the in vitro kinase assay. While this work was in progress, Lombardo et al. (2004) reported the kinase selectivity profile of BMS-354825, which showed that BMS-354825 inhibited PDGFR in vitro with an IC_{50} of 28 nM. Although Lombardo et al. (2004) performed no cell-based assay in their study, our data and theirs are consistent in demonstrating that BMS-354825 is a potent PDGFR inhibitor in vitro. The slight difference in IC_{50} for inhibition of PDGFR may reflect different assay conditions used to determine the PDGFR kinase activity between these two studies.

We found that Src is basally active in A10 cells and that BMS-354825 inhibits the basal Src activity (IC_{50} = 2 nM). Src kinase activity may be essential for cellular activities such as migration (Heldin and Westermark, 1999). Thus, blocking Src activity could contribute to the overall effect of BMS-354825 in inhibition of PDGF-stimulated VSMC migration. However, we did not detect measurable PDGF-induced Src activation in A10 cells, whereas marked activation of PDGFR was readily detectable. Thus, Src kinase activity alone is insufficient to account for the higher migratory activity of VSMCs observed in the presence of PDGFR. Furthermore, although imatinib has no Src tyrosine kinase inhibitor activity (Druker et al., 1996), it suppressed the PDGFR-stimulated activation of PDGFR, STAT3, Akt, and Erk2 in A10 cells. Therefore, inhibition of the PDGFR tyrosine kinase activity alone could have major effects on blocking PDGF-stimulated responses in VSMCs.

Several studies have shown that PDGFR inhibitors such as imatinib and AG-1295 are effective in suppressing restenosis and stenosis in experimental animals (Myllarniemi et al., 1999; Fishein et al., 2000; Sihvola et al., 2003; Levitzki, 2005). Inhibition of PDGFR in VSMCs by imatinib and AG-1295 requires micromolar concentrations (Fishein et al., 2000; Sanz-Gonzalez et al., 2004). In contrast, near-complete inhibition of PDGFR activation and PDGFR-stimulated activities in VSMCs was observed with 50 nM BMS-354825 in this study. Direct comparison of BMS-354825 and imatinib in A10 cells indicates that BMS-354825 is 67-fold more potent that imatinib (Fig. 2). Thus, BMS-354825 is a much more potent PDGFR inhibitor than imatinib.

Inhibition of PDGF-stimulated VSMC proliferation by BMS-354825 is reversible. It has been reported that the vascular endothelial growth factor receptor is not sensitive to BMS-354825 inhibition (Lombardo et al., 2004). These observations suggest that BMS-354825 is unlikely to cause unmanageable damage to the vascular system. In support of this notion, imatinib, which also inhibits PDGFR, does not cause an adverse effect in the cardiovascular system in CML patients who have been treated with the drug for more than 4 years (Lahaye et al., 2005). The reversibility of the inhibitory effect of PDGF-stimulated VSMC proliferation by BMS-
354825 also raises the question of whether BMS-354825 will be effective in suppressing restenosis. It is possible that restenosis-promoting activities peak shortly after vascular injury and lessen afterward. Therefore, one would predict that VSMC proliferation is not likely to resume when BMS-354825 is terminated after sufficient period of treatment unless there is new vascular injury.

BMS-354825 is currently being developed as an anticancer drug. We demonstrated here that it is very effective in inhibiting PDGF-stimulated VSMC migration and proliferation. VSMC migration and proliferation are critical events in the development of restenosis, which is a major problem for the long-term efficacy of the widely performed angioplasty and stenting procedures for coronary artery disease. Thus, in addition to being a promising anticancer drug, BMS-354825 is a potential novel therapeutic agent for cardiovascular diseases involving abnormal VSMC activities such as restenosis and stenosis.

References
Deininger M, Buchdunger E, and Druker BJ (2005) The development of imatinib as a potential novel therapeutic agent for cardiovascular diseases involving abnormal VSMC activities such as restenosis and stenosis.

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