Negative Gating Modulation by (R)-N-(Benzimidazol-2-yl)-1,2,3,4-tetrahydro-1-naphthylamine (NS8593) Depends on Residues in the Inner Pore Vestibule: Pharmacological Evidence of Deep-Pore Gating of KCa2 Channels

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ABSTRACT

Acting as a negative gating modulator, (R)-N-(benzimidazol-2-yl)-1,2,3,4-tetrahydro-1-naphthylamine (NS8593) shifts the apparent Ca2+-dependence of the small-conductance Ca2+-activated K+ channels KCa2.1–2.3 to higher Ca2+ concentrations. Similar to the positive KCa2 channel-gating modulators 1-ethyl-2-benzimidazolinone (1-EBIO) and cyclohexyl-[2-(3,5-dimethylpyrazol-1-yl)-6-methylpyrimidin-4-yl]-piperidine (CyP-TR), the gating site for NS8593 has been assumed to be located in the C-terminal region, in which these channels interact with their Ca2+ sensor calmodulin. However, by using a progressive chimeric approach, we were able to localize the site-of-action of NS8593 to the KCa2.2 pore. For example, when we transferred the C terminus from the NS8593-insensitive intermediate-conductance KCa3.1 channel to KCa2.3, the chimeric channel remained as sensitive to NS8593 as wild-type KCa2.3. In contrast, when we transferred the KCa2.3 pore to KCa3.1, the channel became sensitive to NS8593. Using site-directed mutagenesis, we subsequently identified two specific residues in the inner vestibule of KCa2.3 (Ser507 and Ala532) that determined the effect of NS8593. Mutation of these residues to the corresponding residues in KCa3.1 (Thr250 and Val275) made KCa2.3 insensitive to NS8593, whereas introduction of serine and alanine into KCa3.1 was sufficient to render this channel highly sensitive to NS8593. It is noteworthy that the same two residue positions have been found previously to mediate sensitivity of KCa3.1 to clotrimazole and 1-[(2-chlorophenyl)diphenylmethyl]-1H-pyrazole (TRAM-34). The location of Ser507 in the pore-loop near the selectivity filter and Ala532 in an adjacent position in S6 are within the region predicted to contain the KCa2 channel gate. Hence, we propose that NS8593-mediated gating modulation occurs via interaction with gating structures at a position deep within the inner pore vestibule.

Introduction

Within the superfamily of Ca2+-activated K+ channels (KCa), the group of small (KCa2.1–2.3) and intermediate-conductance (KCa3.1) channels are inward-rectifying, voltage-independent, and activated solely by intracellular Ca2+ (Stocker, 2004; Wulff et al., 2007). The opening of both KCa2 and KCa3.1 channels is initiated via Ca2+-binding to the N-lopes of calmodulin (CaM) constitutively attached to a calmodulin binding domain (CaMBD) located in the proximal intracellular C terminus (Xia et al., 1998; Fanger et al., 1999). The energy of the ensuing conformational change is transferred to the C terminus of calmodulin, which is then capable of interacting with intracellular targets.

ABBREVIATIONS: KCa, Ca2+-activated K+ channel; CaM, calmodulin; BMB, bicuculline methobromid; CaMBD, calmodulin binding domain; 1-EBIO, 1-ethyl-2-benzimidazolinone; GW542573X, 4-(2-methoxy-phenyl)carbamoyloxymethyl)piperidine-1-carboxylic acid tert-butyl ester; TM, transmembrane; HEK, human embryonic kidney; KCa2.1, big conductance Ca2+-activated K+ channel; KCa2.2, small conductance Ca2+-activated K+ channel; KCa3.1, intermediate conductance Ca2+-activated K+ channel; NS309, 6,7-dichloro-1H-indole-2,3-dione 3-oxide; NS8593, (R)-N-(benzimidazol-2-yl)-1,2,3,4-tetrahydro-1-naphthylamine; TRAM-34, 1-[(2-chlorophenyl)diphenylmethyl]-1H-pyrazole; WT, wild type; S6A, S6A-1, naphtho[1,2-d]thiazole-2-ylamine; UCL1684, 6,10-diaza-3(1,3)B[1,4]-dibenzena-1,5(1,4)-diquinolinacyclodecaphane; NS11757, (1H-benzimidazol-2-yl)-(6,7-dichloro-1,2,3,4-tetrahydronaphthalen-1-yl)amine; ICA-27243, N-(6-chloro-pyridin-3-yl)-3,4-difluoro-benzamide.
to the transmembrane (TM) regions to open the gate. Unlike KCa channels, which are gated by a rotational constriction of the intracellular aperture formed by the lower part of the four S6 TM helices, the physical gates of KCa,2 and KCa,3.1 channels seem to be deeply buried in the inner pore vestibule, close to or even overlapping with the K⁺ selectivity filter (Bruening-Wright et al., 2002, 2007; Klein et al., 2007; Garneau et al., 2009).

Despite their structural and functional similarity, KCa,2 and KCa,3.1 channels have a very different pharmacology, the details of which are increasingly becoming better defined at the molecular level. Selective peptide inhibitors of KCa,2 channels, such as apamin or scyllatoxin, and KCa,3.1 channel-inhibiting peptides, such as charybdotoxin and maurotoxin, interact with extracellularly exposed amino acids in the outer pore vestibule (Ishii et al., 1997; Rauer et al., 2000; Castle et al., 2003) and are usually conceived to inhibit via a simple blocking mechanism (Lamy et al., 2010 describes an emergent different view on the apamin mode-of-action). The same applies to the positively charged small-molecule KCa,2-selective blockers such as 6,10-diaza-3(1,3)8(1,4)-dibenzena-1,5(1,4)-diquinolinacyclodecaphane (UC1684) (Campos Rosa et al., 2000), which were designed to mimic the charged part of the apamin molecule, as well as to bicusculine methiodide (Johnson and Seutin, 1997), a lower-affinity blocker.

In contrast, the inhibition by the established small-molecule blockers of KCa,3.1, the triarylmethanes, exemplified by clotrimazole and the more selective 1-[(2-chlorophenyl)diphenylmethyl]-1H-pyrazole (TRAM-34) (Wulf et al., 2001), is mediated via two amino acids located in the lower part of the pore loop (Thr250) and in the S6 segment (Val275), respectively (Wulf et al., 2001). These amino acids form part of a ring of hydrophobic residues, which line and thus isolate the upper part of the watery inner pore vestibule. Their position close to the selectivity filter suggested a model in which TRAM-34 coordinates via its aryl groups to Thr250 and Val275, in the S6 segment (Val275), respectively (Wulf et al., 2001). These amino acids conferring TRAM-34 sensitivity. We therefore conclude that NS8593 inhibition involves deep-pore amino acids and suggest that the compound’s negative gating modulation may result from a direct interaction with the physical gate of KCa,2 channels.

**Materials and Methods**

**Molecular Biology.** HEK293 cell lines stably expressing WT hKCa,2,3 and hKCa,3.1 channels were described previously (Hougaard et al., 2009). The N-terminal KCa,3.1-H11001 chimera was generated with overlapping polymerase chain reaction (Expand High Fidelity PCR System; Roche Diagnostics, Mannheim, Germany) using the oligonucleotides hK-hSiK3s GTCTCTGGCCGGCTGCCCAGAgatgtgtatgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtgtg
as templates in mutagenesis reactions. Four oligonucleotides (MWG-Biotech AG, Ebersberg, Germany) were used to introduce the mutations: hKCa₂.3 S507T, CTCATCCACATTCTTAsATTGTTATGGOGACA; hKCa₂.3 A532V, TCACTGGCATCATGCTGATAGGCTGACAGGCTT-TGGTGTGCCG; hKCa₃.1 T250S, GATCCCCATCATTCCTGCaATGC-GCTATGTTGACGTG; and hKCa₃.1 V275A, GCACTGGGAATCAGGGT-GcaTGTCGACAGGCTTGT. The mutagenesis reactions were performed using T7 DNA polymerase and T4 DNA ligase (New England Bioslabs, Ipswich, MA; E. coli XLI-Blue [Strategene] was transformed with an aliquot of the reaction, and the resulting plasmid DNA was purified using standard methods. All constructs were verified by sequencing.

Electrophysiology. All experiments were performed on transiently transfected HEK 293 cells in either the inside-out or the whole-cell configuration of the patch-clamp technique. Lipo-fectamine and standard transfection methods were used, and recordings were made 2 days after transfection. Cells for whole-cell experiments were detached by trypsinization and plated on coverslips (3.5 mm Ø) on the day of the experiments, whereas cells for inside-out recordings were plated 1 day before the experiments to attach them more firmly. For recordings, a coverslip was placed in a 15-μl recording chamber mounted on the cross-board of an inverted microscope (Olympus XI-70 equipped with fluorescence burner and filters; Olympus, Tokyo, Japan), and cell selection was guided by fluorescence from the cotransfected green fluorescent protein. The extracellular solutions contained 154 mM KCl, 10 mM HEPES, pH 7.4, 2 or 0.1 mM CaCl₂, and 1 or 3 mM MgCl₂ for inside-out/whole-cell experiments, respectively. Solutions on the intracellular side contained 154 mM KCl, 10 mM HEPES, pH 7.2, 10 mM EGTA or 1 mM EGTA plus 9 mM nitriloacetic acid, CaCl₂, and MgCl₂ to yield a calculated free Mg²⁺ concentration of 1 mM and calculated free Ca²⁺ concentrations of 0.01, 0.2, 0.3, 0.4, 0.5, 3, 10, and 30 μM. Solutions used for experiments with ATP were made with Na₂ATP (Sigma-Aldrich, St. Louis, MO) and adjusted to yield 30 μM free Ca²⁺ and 1.6 mM Mg²⁺-ATP. Cells or membrane patches were perfused at 1 ml/min by gravity from a 10-position solution exchanger. Patch pipettes were pulled from borosilicate (Vitrex Medical A/S, Herlev, Denmark) or soda lime glass (micro-hematocrit tubes; Kimble Chase, Rochester, NY) and had resistances of 2 to 3 MΩ when submerged in the bath solution. Positioning of the patch electrode was controlled with a Patchman micromanipulator (Eppendorf North America, New York, NY). Any initial voltage difference between the patch electrode and the integrated and grounded bath electrode was eliminated before the pipette was attached to the cell. Experiments were controlled with a HEKA EPC-9 or EPC-10 amplifier and Pulse software (HEKA, Lambrecht/Pfalz, Germany). Cells were clamped to a holding potential of at 0 mV, and KCa currents were elicited by 200-ms voltage ramps from −80 to +80 mV applied every 5 s with 400 nM free Ca²⁺ in the patch pipette. As published previously for recordings with symmetrical K⁺ concentrations in this expression system (Strøbaek et al., 2006; Hougaard et al., 2007), KCa₂.3 exhibited a much more pronounced inward rectification than KCa₃.1, which showed a nearly linear IV relationship. Both channels exhibited their characteristic pharmacology. KCa₂.3 was potently inhibited by NS8593 with an IC₅₀ of 104 nM (Tables 1 and 2) and almost fully blocked with 100 μM BMB (IC₅₀ 5 μM), which we routinely use to estimate leak when working with symmetrical K⁺ solutions. KCa₃.1, in contrast, was completely insensitive to 10 μM NS8593 and 100 μM BMB but could be blocked completely by TRAM-34 (Tables 1 and 2). NS309 increased both KCa₂.3 and KCa₃.1 currents but was distinctly more effective on KCa₃.1 in keeping with previous reports (Strøbaek et al., 2004).

We next tested the effect of NS8593 on KCa₂.3 chimeras that contained either the N- or the C-terminal regions of KCa₃.1. Because both chimeras contained the pore region of KCa₂.3, they exhibited the characteristic inward rectification of KCa₂.3. Surprisingly, NS8593 inhibited both chimeras as potently as the WT KCa₂.3 channel (Fig. 2, C and D), suggesting that the NS8593-induced negative gating modulation is not mediated via the N- or the C-terminal regions. Exchanging the C terminus between KCa₂.3 and KCa₃.1 also transferred the higher NS309 sensitivity of KCa₃.1 to the resulting chimeric channel (Fig. 2C, right). This observation is reminiscent of the increase in 1-EBIO sensitivity that was reported for a KCa₂.2 chimeric channel containing the C terminus of KCa₃.1 (Pedarzani et al., 2001) and suggests that the binding site of the positive gating modulator NS309 is also located in this region.

Amino Acids in the Pore Region Confer Sensitivity to NS8593. After having obtained the above results, we next focused our attention on the TM regions. Unfortunately, some KCa₂.3/KCa₃.1 chimeras in which various parts of TM5 and/or TM6 were swapped between the two channels did not seem to form functional channels or were expressed too poorly to allow for evaluation of modulator sensitivity (Supplemental Table 1). However, an important chimera that contained the transmembrane regions S1 to S4 from KCa₃.1 and the pore and C terminus of KCa₂.3 (Fig. 2E) expressed sufficiently well for pharmacological experiments. It is noteworthy that this chimera was as sensitive to NS8593 as the wild-type KCa₂.3 channel (Fig. 2E, left) and was further found to be insensitive to the KCa₂.3 pore blocker BMB. Both observations were surprising, the first suggesting that NS8593 might be exerting its effect by interacting with residues in the pore. As a consequence, we decided to follow-up this observation with point mutations in the inner pore re-
The latter observation could indicate that the presence of the S1 to S4 region from KCa3.1 in the chimera might have disturbed the architecture of the outer KCa2.3 vestibule, in which BMB, which exhibits voltage-dependent block (Grunnet et al., 2001) and displaces apamin (Finlayson et al., 2001), presumably binds. In this context, it should be mentioned that apamin was very recently reported to bind to the outer pore of KCa2 channels (Lamy et al., 2010),

Fig. 2. The inhibitory effect of NS8593 is not dependent on N- and C-terminal regions of KCa2.3 but on pore regions. Effect of NS8593, BMB, and NS309 on whole-cell currents from HEK293 cells expressing WT KCa2.3 (A), WT KCa3.1 (B), and the chimeric constructs KCa2.3-KCa3.1(292–427) (C), KCa3.1-KCa2.3(294–736) (D), and KCa3.1-KCa2.3(435–736) (E). Voltage ramps were applied every 5 s from a holding potential of 0 mV. Control traces (last sweep before compound application) are shown in black, and compound traces (last sweep before compound washout) are shown in red. Washout traces have been omitted for clarity. Symmetric K⁺ distributions with free [Ca²⁺] in the pipette solution buffered to 400 nM (see Materials and Methods for specifications).
where it apparently acts more as an allosteric modulator of the selectivity filter than as a direct blocker such as tetraethyl ammonium or the larger scorpion toxins tamapin and scyllatoxin, which are able to span the KCa2.3 channel pore (Weatherall et al., 2010).

Two Amino Acids in the Inner Pore Region Are Required and Sufficient for Inhibition by NS8593. Two amino acids in the inner pore vestibule were reported previously to confer TRAM-34 and clotrimazole sensitivity to KCa3.1 (Wulff et al., 2001). These amino acids, Thr250 and Val725 in the KCa3.1 numbering, are located on either side of the selectivity filter, and we therefore speculated whether the effect of NS8593 might also be dependent on these amino acids. As shown in Tables 1 and 2 and Fig. 3, mutations of each of the corresponding residues in KCa2.3 (Ser507 and Ala532) reduced the potency of NS8593 roughly 20-fold, whereas introduction of the double mutation produced a KCa2.3 channel that was completely insensitive to NS8593.

The importance of Ser507 and Ala532 was verified by showing that the reverse KCa3.1 mutants (T250S and V275A) became sensitive to NS8593 and that the KCa3.1 double mutant was at least as sensitive to NS8593 as the wild-type KCa2.3 channel (Fig. 3 and Tables 1 and 2). As a control of the effect of NS8593 might also be dependent on these amino acids as functionally important pore residues.

**TABLE 1**

<table>
<thead>
<tr>
<th>Chimeras</th>
<th>NS8593 IC50</th>
<th>Current Inhibition by 100 μM BMB</th>
<th>Current Increase by 30 nM NS309</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>μM</td>
<td>%</td>
<td>fold</td>
</tr>
<tr>
<td>KCa2.3 WT</td>
<td>0.104 ± 0.034 (13)</td>
<td>83 ± 13 (10)</td>
<td>2.1 ± 0.5 (6)</td>
</tr>
<tr>
<td>KCa3.1 WT</td>
<td>&gt;10 (11)</td>
<td>0 ± 0 (5)</td>
<td>5.1 ± 2.1 (6)</td>
</tr>
<tr>
<td>KCa2.3-KCa3.1S507T</td>
<td>0.117 ± 0.013 (3)</td>
<td>57 ± 24 (3)</td>
<td>2.9 ± 0.6 (3)</td>
</tr>
<tr>
<td>KCa3.1-KCa2.3A532V</td>
<td>0.115 ± 0.040 (6)</td>
<td>91 ± 3 (6)</td>
<td>1.1 ± 0.5 (6)</td>
</tr>
<tr>
<td>KCa3.1-KCa2.3S507T</td>
<td>0.947 ± 0.031 (4)</td>
<td>0 ± 0 (3)</td>
<td>2.3 ± 0.4 (3)</td>
</tr>
</tbody>
</table>

**TABLE 2**

<table>
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<tr>
<th>Mutants</th>
<th>NS8593 IC50</th>
<th>Current Inhibition by 100 μM BMB</th>
<th>TRAM-34 IC50</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>μM</td>
<td>%</td>
<td>μM</td>
</tr>
<tr>
<td>KCa2.3 WT</td>
<td>0.104 ± 0.034 (13)</td>
<td>83 ± 13 (10)</td>
<td>&gt;20a</td>
</tr>
<tr>
<td>KCa2.3S507T</td>
<td>2.5 ± 0.78 (9)</td>
<td>42 ± 18 (5)</td>
<td>&gt;1 (3)</td>
</tr>
<tr>
<td>KCa2.3A532V</td>
<td>3.1 ± 1.3 (4)</td>
<td>67 ± 12 (2)</td>
<td>2.7 (1)</td>
</tr>
<tr>
<td>KCa2.3S507T + A532V</td>
<td>&gt;10 (3)</td>
<td>65 ± 15 (3)</td>
<td>0.064 ± 0.068 (2)</td>
</tr>
<tr>
<td>KCa3.1 WT</td>
<td>&gt;10 (11)</td>
<td>0 ± 0 (5)</td>
<td>22 ± 14 (2)</td>
</tr>
<tr>
<td>KCa3.1T250S</td>
<td>0.503 ± 0.209 (11)</td>
<td>0 ± 0 (5)</td>
<td>21 ± 4.3 (3)</td>
</tr>
<tr>
<td>KCa3.1V275A</td>
<td>3.5 ± 1.6 (14)</td>
<td>0 ± 0 (7)</td>
<td>&gt;20a</td>
</tr>
<tr>
<td>KCa3.1S507T + V275A</td>
<td>0.056 ± 0.024 (3)</td>
<td>1 ± 1 (2)</td>
<td>&gt;20a</td>
</tr>
</tbody>
</table>

* Wulff et al. (2001).
**K_{Ca2.3}**

**K_{Ca3.1}**

\[ VTNFLGAMWLISITFLS\text{G}GYDMVPHTYC\text{K}GVCLLTGIMGAGCTAL \]
\[ ATGLHD\text{T}LWLIPITFTLT\text{G}GDVVP\text{G}TWGKIVCLCTGV\text{G}CCTAL \]
modulation: 1) dependence on [Ca\textsuperscript{2+}]; and 2) reversibility in presence of the positive modulator NS309.

As described previously (Strøbaek et al., 2006), current reduction by the negative gating modulator NS8593 is strongly dependent on the [Ca\textsuperscript{2+}], with the compound’s potency decreasing with increasing [Ca\textsuperscript{2+}]. As shown in Fig. 5A, perfusion of 1 μM NS8593 onto KCa\textsubscript{2.3} currents activated with 300 nM [Ca\textsuperscript{2+}], in inside-out patches blocked 74 ± 6% (n = 6) of the calcium-dependent inward current at –80 mV. However, when the same patch was subsequently exposed to 30 μM [Ca\textsuperscript{2+}] to maximally activate KCa\textsubscript{2.3}, 1 μM NS8593 only blocked 21 ± 5% (n = 6) of the current. After washout and another control for absence of contaminating leak current by a switch to 10 nM [Ca\textsuperscript{2+}], KCa\textsubscript{2.3} was then exposed to 0.5 μM [Ca\textsuperscript{2+}], and 1 μM NS8593 was observed to exhibit an intermediate potency (58 ± 14% blockade, n = 5). In contrast, inhibition of the KCa\textsubscript{3.1} double mutant (KCa\textsubscript{3.1, T250S-V275A}) by NS8593 was much less calcium-dependent (Figs 5B and 6). In a similar inside-out experiment, in which [Ca\textsuperscript{2+}] varied from 0.3 to 30 μM, NS8593 at 1 μM inhibited 87 ± 3% (n = 3) at 0.3 μM, 80 ± 4% (n = 3) at 0.5 μM, and 70 ± 6% (n = 5) of the current at 30 μM [Ca\textsuperscript{2+}]. However, the reduced calcium-dependence of the inhibitory effect of NS8593 on the KCa\textsubscript{3.1} double mutant was not simply the result of the slightly higher affinity of NS8593 to the mutant channel because inhibition remained less calcium-sensitive even when the NS8593 concentration was reduced 10-fold from 1 μM to 100 nM (Fig. 6).

The NS8593 inhibition of KCa channels at low [Ca\textsuperscript{2+}] is essentially abolished upon the addition of the positive modulator NS309 (Strøbaek et al., 2006; Ji et al., 2009). Figure 7 compares the ability of NS309 to reverse the inhibitory effect of NS8593 on KCa\textsubscript{2.3} (Fig. 7A) and the KCa\textsubscript{3.1, T250S-V275A} mutant (Fig. 7B). In both cases, upon stabilization of the whole-cell current (black traces, left), 1 μM NS8593 was superfused causing nearly 100% inhibition (red traces, middle, after 150 s). Increasing concentrations of NS309 were then cosuperfused with NS8593 leading to a concentration-dependent reversal and “overshoot” of the KCa\textsubscript{2.3} current, whereas the KCa\textsubscript{3.1, T250S-V275A} current remained largely inhibited even at an NS8593 concentration of 1 μM (green traces, middle, after 250 s). This series of experiments has been quantified (Fig. 7, right), in which the fold decrease/increase relative to the control current is plotted for the NS8593 (1 μM) equilibrium inhibition and for the combined NS8593 + NS309 (1 μM) effect. The overall conclusion from these experiments is that NS8593-mediated inhibition of the KCa\textsubscript{3.1, T250S-V275A} construct is not reversible with a positive modulator and in this respect has lost its negative modulation character and rather resembles a blocker.

Finally, in a last attempt to demonstrate negative gating modulation characteristics on the KCa\textsubscript{3.1, T250S-V275A} construct, we performed a series of inside-out experiments with combined high [Ca\textsuperscript{2+}] (30 μM) and Mg\textsuperscript{2+}-ATP (1.6 mM) or NS309 (1 μM). The rationale for using Mg\textsuperscript{2+}-ATP quote back to the observation that the Ca\textsuperscript{2+}-dependent open channel probability \( P_\text{d}(\text{max}) \) value of KCa\textsubscript{3.1} is significantly lower than the corresponding value for KCa\textsubscript{2.2} in some studies (Gerr-lach et al., 2001; Jones et al., 2007), and that this value can be increased significantly in the presence of Mg\textsuperscript{2+}-ATP. In our hands, Mg\textsuperscript{2+}-ATP and NS309 both induced slight increases (28%, n = 2, and 23%, n = 3, respectively), in the maximal current level of KCa\textsubscript{3.1, T250S-V275A} (results not shown), and the ensuing addition of NS8593 (0.3 μM) caused current reductions that were not significantly different from the effects without Mg\textsuperscript{2+}-ATP or NS309 (55 ± 8%, n = 5 versus 52 ± 7%, n = 3, and 51 ± 7%, n = 4, respectively). Likewise, the higher NS8593 concentration of 1 μM blocked the KCa\textsubscript{3.1, T250S-V275A} mutant channel by 70% in both the absence (n = 5) and the presence of 1.6 mM ATP (n = 2).

Taken together, these data suggest that although the KCa\textsubscript{3.1, T250S-V275A} mutation perfectly transfers the interaction site for NS8593 in terms of potency, the characteristic coupling to the gate is less pronounced in this construct, as shown by diminished Ca\textsuperscript{2+} - and NS309-dependence of inhibition and by the preservation of its inhibitory potency in the presence of Mg\textsuperscript{2+}-ATP.

**Discussion**

In comparison with the classic KCa\textsubscript{2.2} channel blocker apamin, NS8593 acts by a different mechanism and via a different site because KCa\textsubscript{2.3} channels mutated in the apamin binding site remain sensitive to its actions (Sørensen et al., 2008). NS8593 produces a rightward-shift in the KCa\textsubscript{2.2} channel Ca\textsuperscript{2+} activation curve and has therefore been termed a negative gating modulator rather than a pore blocker (Strøbaek et al., 2006; Sørensen et al., 2008). This article attempts to define the site-of-action for NS8593.

The selectivity of NS8593 for KCa\textsubscript{2.2} channels over the structurally related KCa\textsubscript{3.1} channel allowed us to use a chimera/mutagenesis approach to identify regions and specific amino acids important for the NS8593 effect. In short, we have shown that KCa\textsubscript{2.3} loses sensitivity toward NS8593 inhibition by the point mutations S507T and A532V and, on the other hand, that KCa\textsubscript{3.1} gains sensitivity to NS8593 by the equivalently positioned mutations T250S and V275A. These results have at least two noteworthy implications. First, according to the generally accepted gross architecture of 6-TM K’-channels, both of these amino acids are located in the inner pore vestibule (inner pore helix of S5 and S6, respectively) close to the inside of the selectivity filter, in which they define open channel properties such as inward rectification/divalent cation block. Hence, because of the coupling between NS8593 inhibition and Ca\textsuperscript{2+}-dependent gating (Strøbaek et al., 2006), we assume that Ser507 and Ala532 are positioned close to the physical gate of the channel. Second, these positions are the same two positions that define KCa\textsubscript{3.1} sensitivity (and KCa\textsubscript{2.2} insensitivity) toward triaryl methane such as TRAM-34 and clotrimazole (Wulff et al., 2001) and to arachidonic acid (Hamilton et al., 2003). This coincidence of equivalent sites for selective inhibitors of the two channels underscores the close structural resemblance

![Fig. 3. Ser507 and Ala532 confer NS8593 sensitivity to KCa\textsubscript{2.3}, and their transfer into KCa\textsubscript{3.1} renders KCa\textsubscript{3.1} sensitive to NS8593. Effects of 1 and 10 μM NS8593 on KCa\textsubscript{2.3} (left) and KCa\textsubscript{3.1} (right) single and double mutants, in which inner pore residues were mutated to the equivalent amino acids of the other channel. A partial sequence alignment of hKCa\textsubscript{3.1} and hKCa\textsubscript{2.3} in the P-loop and the S6 region with the position of the residues highlighted is shown on top. Experimental details and color-coding are as stated in Fig. 1.](molpharm.aspetjournals.org)
between the KCa2 and KCa3 families. However, our data also point toward significant functional differences between the two channel families. Although NS8593 inhibition via this region in KCa2.3 exhibits strong negative gating modulation (abolishable by Ca2\(^{++}\) and coapplication of positive modulators), Ca2\(^{++}\)-dependence and NS309-mediated reversion are barely detectable for its inhibition of the KCa3.1T250S/V275A construct. The present results seem not to support the interpretation that the increased potency and abolished gating dependence is due solely to a low \(P_{o}(\text{max})\) value (favoring a possible closed state binding of NS8593), because combined experimental conditions tending toward a high \(P_{o}\) (high \([\text{Ca}^{2+}]_i/\text{NS309 or Mg}^{2+}\text{-ATP}\)), do not shift the potency significantly. It is noteworthy that the same is the case for the TRAM-34/clotrimazole inhibition of KCa3.1, in which we have previously found that clotrimazole and TRAM-34 inhibited NS309 or naphtho[1,2-d]thiazol-2-ylamine (SKA-31) activated KCa3.1 channels with essentially the same potency as nonactivated channels (Strobaek et al., 2004; Sankaranarayanan et al., 2009). Together, this series of results may warrant the consideration of whether these residue positions are simply less intimately coupled to the gating process in KCa3.1 than the equivalent positions in KCa2.3.

Equivalent deep-pore amino acid positions allowing 3 or- ders of magnitude of inhibitory selectivity between KCa2 and KCa3 families provoke the question of whether interaction with these positions might potentially also cause selectivity between the KCa2.1, KCa2.2, and KCa2.3 subtypes, a matter of considerable pharmacological and potential clinical relevance: KCa2.1 and KCa2.2 are mostly expressed in the cortical/limbic structures of the brain, whereas KCa2.3 is prefer- entially expressed in the basal ganglia and in other subcortical regions (Sailer et al., 2004). KCa inhibitors have

Fig. 4. The KCa2.3S507T/A532V and the KCa3.1T250S/V275A double mutants exhibit similar biophysical and pharmacological properties as the WT channels. A, overlay of the calcium-response curves of KCa2.3 and the two double mutants measured from inside-out patches exposed to increasing Ca2\(^{++}\) concentrations. Data points are mean ± S.D. from seven or eight experiments per construct. The fit of the data to the Hill equation yielded the following results: KCa2.3 (EC50 480 ± 50 nM, \(n_H = 4.8 ± 1.0\)); KCa2.3S507T/A532V (EC50 590 ± 40 nM, \(n_H = 4.4 ± 0.5\)), and KCa3.1T250S/V275A (EC50 520 ± 70 nM, \(n_H = 3.8 ± 0.9\)) B, whole-cell recording showing effects of the pore-blocking toxins apamin (left) and charybdotoxin (right) on the KCa2.3S507T/A532V and the KCa3.1T250S/V275A double mutants. Experimental details as stated in text to Fig. 1. (C) Rectification of the different KCa2.3 and KCa3.1 constructs determined by the ratio of the current amplitude at \(-80\) and \(+80\) mV. Values are mean ± S.D. (n = 2–15 per data point).

Fig. 5. Inhibition of WT KCa2.3 (A) by NS8593 is more Ca2\(^{++}\)-dependent than inhibition of the KCa3.1T250S/V275A mutant (B). Inside-out patches were exposed to a \([\text{Ca}^{2+}]_i\) of 0.01, 0.3, 0.5, or 30 \(\mu\text{M}\) as indicated in the presence or absence of NS8593. The main figures show a continuous plot of the currents recorded at \(-75\) mV. The inserts show control and NS8593 current traces. Experimental details and color-coding are as described in Fig. 1.
been considered for improvement of cognitive performance (primarily KCa2.2) (Hammond et al., 2006), whereas there is evidence for the use of KCa2.3 inhibitors for mood disorders such as depression (Jacobsen et al., 2008). Unfortunately, both Ser507 and Ala532 are conserved among the KCa2 subtypes (Ser330 and Ala355 in KCa2.1; Ser359 and Ala384 in KCa2.2), which implies that selective inhibition of the KCa2 channel members may be difficult to achieve. In support of this interpretation, a detailed structure-activity analysis on NS8593 analogs revealed no subtype-selectivity between KCa2.2 and KCa2.3, despite the achievement of considerably increased potency for negative gating modulation (Sørensen et al., 2008). In a study of non–apamin-displacing KCa2 inhibitors of a different chemotype [4-(aminomethylaryl)pyrazolopyrimidine], subtype selectivity was also not observed (Gentles et al., 2008), but whether these compounds are interacting with the same site as NS8593 is currently not known. However, it might of course be possible to achieve some functional selectivity among the KCa2 subtypes based on the different Ca²⁺ concentrations in the microscopic environment surrounding the channel’s calmodulin and its phosphorylation state (Bildl et al., 2004; Allen et al., 2007) in different types of neurons and brain regions.

An Emerging Picture of KCa2 Channel-Gating Pharmacology. Because the predominant effects of the prototypical positive and negative gating modulators are left-shifting/right-shifting of the [Ca²⁺]ᵢ-response curve, respectively, their phenomenological mode-of-actions were originally attributed to selective increases/decreases in the “apparent Ca²⁺ affinity” of KCa2 channels, not excluding a priori an interference with the genuine CaM binding affinity for Ca²⁺.

The demonstration that the C terminus is the site of 1-EBIO-mediated positive modulation of KCa2.1 (Pedarzani et al., 2001), a finding that has been confirmed by the Kₐ₃.₂/₃-KCa2.2-selective cyclohexyl-[2-(3,5-dimethyl-pyrazol-1-yl)-6-methylpyrimidin-4-yl]-amine (Kₐ₃.₂/₃>Kₐ₃.₁ = Kₐ₃.₂/₃) (Hougaard et al., 2008), further strengthened the view of positive modulation occurring by a comparatively simple “local” C-terminal mechanism. We initially imagined a similar C-terminally “delimited” action for negative gating modulation, in particular with reference to the quite similar negative gating effect of casein kinase 2-mediated phosphorylation of Thr80 in the attached CaM (Bildl et al., 2004; Allen et al., 2007).

However, a number of findings significantly complicate this simple, unifying picture. First, localization of the site-of-action for NS8593 to deep-pore amino acids immediately excludes the possibility that positive and negative modulators simply share the same binding site and just differ in their (positive or negative) coupling to the gating process in analogy to the action of benzodiazepines on the GABAA receptor (Sieghart, 1994). In addition, recent findings show...
that NS309, which is a classic positive modulator like 1-EBIO, strengthens the link between CaM-Ca$^{2+}$ and channel opening rather than increasing Ca$^{2+}$ binding to CaM per se (Li et al., 2009). Furthermore, we have described recently a K$_{Ca}$2.1-selective activator (K$_{Ca}$2.1$\pm$/K$_{Ca}$2.2 = K$_{Ca}$2.3$\pm$/K$_{Ca}$3.1) 4-(2-methoxy-phenylcarbamoyloxy)methyl-piperidine-1-carboxylic acid tert-butyl ester (GW542573X), which exhibits a complex, partial agonist-like mode of action (Hougaard et al., 2009) that is also independent of the CaMBD/C-terminal region but is dependent on selective interaction with Ser293 (Leu476 in K$_{Ca}$2.3) in the S3 segment. In line with the present analysis of the interaction site for negative gating modulation by NS8593, the GW542573X results were interpreted as evidence for K$_{Ca}$2.1 activation via “deep-pore” gating structures. At the present time we therefore favor the hypothesis, that the diverse positive/negative modulator pharmacology directly reflects the complexity and extended participation of even remotely positioned parts of K$_{Ca}$2 (and accessory proteins) in the gating process. Cysteine scanning experiments have clearly shown that the gate of both K$_{Ca}$2.2 and K$_{Ca}$3 channels is positioned very close to or even encompasses the quite outwardly displaced K$^+$ selectivity filter (Bruening-Wright et al., 2002, 2007; Klein et al., 2007; Garneau et al., 2009), that finding has to be reconciled with the primary Ca$^{2+}$-binding event occurring on CaM at the cytoplasmic C terminus. We think of K$_{Ca}$2.2 (and K$_{Ca}$3) channel gating as a series of events comprising Ca$^{2+}$-binding, CaM/CaMBD/C-terminal conformational change, leading to a transduction via S6 (possibly involving S5 stabilisation) to deep-pore gating structures and eventual opening of the channel. The emerging complexity of the gating modulators, in terms of site-of-actions, selectivity, and mode-of-actions, most likely reflects the existence of several points for pharmacological intervention along the chain of molecular events leading from Ca$^{2+}$ binding to the eventual channel opening. Both negative and positive allosteric-like gating modulation can be achieved, as well as partial agonism-like activation. However, no simple unifying relation seems to exist between the position of interaction sites and mode-of-actions. Despite the present results obtained with NS8593, we do therefore not exclude that future negative gating modulators of K$_{Ca}$2 channels might act on different sites than Ser507 and Ala532. Indeed, different sites of modulation coupled to different selectivities are not unprecedented for K$^+$ channel gating modulators. Although the “classic” K$_{Ca}$7.2-K$_{Ca}$7.5 channel activator retigabine interacts with a hydrophobic pocket formed upon channel opening between the cytoplasmic parts of S5 and S6 (Wuttke et al., 2005), the more K$_{Ca}$7.2/7.3-selective activator N-(6-chloro-pyridin-3-yl)-3,4-difluoro-benzamide (ICA-27243) was found recently to act through a voltage-sensor domain site located in transmembrane segments S1–S3 (Padilla et al., 2009). In analogy, alternative gating inhibitor sites on K$_{Ca}$2 channels may exist, possibly providing a better opportunity for the achievement of subtype-selectivity.

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Authorship Contributions

Participated in research design: Jenkins, Strøbæk, Christoffersen, and Wulff.

Conducted experiments: Jenkins, Strøbæk, and Hougaard.

Contributed new reagents or analytic tools: Jensen, Hummel (molecular biology), and Sørensen (chemistry).

Performed data analysis: Jenkins, Strøbæk, and Wulff.

Wrote or contributed to the writing of the manuscript: Jenkins, Strøbæk, Christoffersen, and Wulff.

References


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Supplemental Data for Molecular Pharmacology article:

Negative Gating Modulation by (R)-N-(Benzimidazol-2-yl)-tetrahydro-1-naphtylamine (NS8593) Depends on Residues in the Inner Pore Vestibule: Pharmacological Evidence of Deep-Pore Gating of KCa2 Channels

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This file contains:

1 Supplemental Table with information on additional chimeras.
**Supplemental Table 1: Additional Chimeras**

### Non-functional or pharmacologically "oddly"-behaving chimeras

<table>
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<tr>
<th>Chimeras</th>
<th>IC&lt;sub&gt;50&lt;/sub&gt; [nM]</th>
<th>Fold current increase by NS309</th>
<th>% current inhibition by BMB</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>K&lt;sub&gt;Ca.2.3-KCa.3.1&lt;/sub&gt;(233-281)</td>
<td>Not tested</td>
<td>No current increase (n = 5 cells)</td>
<td>Not tested</td>
<td>&lt; 200 pA current (n = 5 cells)</td>
</tr>
<tr>
<td>K&lt;sub&gt;Ca.2.3-KCa.3.1&lt;/sub&gt;(233-427)</td>
<td>No inhibition at 0.3 µM (n=1) or 10 µM (n=2).</td>
<td>No current increase at 0.03 µM (n=2) and only small increase at 10 µM (n=2)</td>
<td>Not tested</td>
<td>Inhibition by ChTX (n = 1) and TRAM-34 (n=3)</td>
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<tr>
<td>K&lt;sub&gt;Ca.3.1-KCa.2.3&lt;/sub&gt;(639-731)</td>
<td>Not tested</td>
<td>Not modulated at 0.3 µM (n = 2)</td>
<td>Not tested</td>
<td>No current (n = 10 cells)</td>
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### Functionally "redundant" chimeras

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<th>Chimeras</th>
<th>IC&lt;sub&gt;50&lt;/sub&gt; [nM]</th>
<th>Fold current increase by NS309</th>
<th>% current inhibition by BMB</th>
<th>Comments</th>
</tr>
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<tbody>
<tr>
<td>K&lt;sub&gt;Ca.2.3-KCa.3.1&lt;/sub&gt;(292-427)</td>
<td>250 ± 60 (4)</td>
<td>126 ± 2 (2)</td>
<td>89 ± 5 (4)</td>
<td>Increased Ca&lt;sup&gt;2+&lt;/sup&gt;-sensitivity in inside-out experiments</td>
</tr>
<tr>
<td>K&lt;sub&gt;Ca.2.3-KCa.3.1&lt;/sub&gt;(378-427)</td>
<td>107 ± 11 (2)</td>
<td>199 ± 15 (3)</td>
<td>90 ± 6 (3)</td>
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<tr>
<td>K&lt;sub&gt;Ca.3.1-KCa.2.3&lt;/sub&gt;(294-642)-K&lt;sub&gt;Ca.3.1&lt;/sub&gt;</td>
<td>457 (1)</td>
<td>130 ± 11 (2)</td>
<td>91 ± 13 (2)</td>
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<tr>
<td>K&lt;sub&gt;Ca.2.3-KCa.3.1&lt;/sub&gt;(404-427)</td>
<td>52 (1)</td>
<td>209 (1)</td>
<td>100 (1)</td>
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<tr>
<td>K&lt;sub&gt;Ca.3.1-KCa.2.3&lt;/sub&gt;(644-731)</td>
<td>&gt;10 µM (2)</td>
<td>400 ± 0 (2)</td>
<td>0 (2)</td>
<td></td>
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</table>
Table 1: List of chimeras not included in the main article because they were either found to be non-functional/low expressing (1-3) or because they were considered to yield only supportive but not additional information with respect to defining the site- or mode-of-action of NS8593 (4-8). The cartoons are color coded with the K_{Ca}2.3 derived sequence in green and the K_{Ca}3.1 sequence in blue. Data are given as n (the number of independent experiments) ± SEM (standard error of the mean). All chimaeras were constructed, expressed and evaluated as detailed in the Materials and Methods section of the article. The concentrations used of NS309 (6,7-dichloro-1H-indole-2,3-dione 3-oxime) and BMB (bicuculline methobromid) were 30 nM and 100 µM, respectively.