Xanthines Down-Regulate the Drug Transporter ABCG2 and Reverse Multidrug Resistance

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ABSTRACT
ABCG2 is an ATP-binding-cassette (ABC) transporter that confers multidrug resistance (MDR) to tumor cells by extruding a broad variety of chemotherapeutic agents, ultimately leading to failure of cancer therapy. Thus, the down-regulation of ABCG2 expression and/or function has been proposed as part of a regimen to improve cancer therapeutic efficacy. In this study, we found that a group of xanthines including caffeine, theophylline, and dyphylline can dramatically decrease ABCG2 protein in cells that have either moderate (BeWo, a placental choriocarcinoma cell line) or high (MCF-7/MX100, a breast cancer drug-resistant cell subline) levels of ABCG2 expression. This down-regulation is time-dependent, dose-dependent, and reversible. Using lysosomal inhibitors, we found that xanthines decreased ABCG2 by inducing its rapid internalization and lysosome-mediated degradation. As a consequence, caffeine treatment significantly increased the retention of an established ABCG2 substrate in MCF-7/MX100 cells but not in parental MCF-7 cells and sensitized the MDR cells to the chemotherapeutic agent mitoxantrone (MX); combination treatment with MX and caffeine decreased the IC50 of MX 10-fold and induced a greater degree of apoptotic cell death than MX treatment alone. Taken together, our results describe a novel function for this large class of therapeutically relevant compounds and suggest that a subset of xanthines could be developed as combination therapy to improve the efficacy of anticancer drugs that are ABCG2 substrates.

Introduction
The ABCG2 (breast cancer resistance protein) protein was first identified as a result of its overexpression in breast cancer cells exhibiting a MDR phenotype in the absence of other “classic” drug transporters (Doyle et al., 1998). Although a member of the same ABC family of membrane transporters as P-glycoprotein and multidrug resistance proteins, ABCG2 belongs to a unique subclass whose genes encode a half transporter with one classic nucleotide-binding domain and six transmembrane segments. Initially believed to exist as a homodimer, recent evidence suggests that ABCG2 may function as an oligomer consisting of 8 to 12 identical subunits (McDevitt et al., 2006) and transports a broad spectrum of natural and synthetic substrates including some cancer chemotherapeutic agents.

Given the role of ABCG2 in conferring MDR, there has been considerable focus on the expression of this transporter in human tumors and its correlation with clinical outcome. Elevated expression of ABCG2 was first reported in patients with acute myeloid leukemia (AML) (Ross et al., 2000) and subsequently confirmed in patients with AML and those with acute lymphocytic leukemia (van den Heuvel-Eibrink et al., 2002; Plasschaert et al., 2003; Wilson et al., 2006). ABCG2 conferred resistance to flavopiridol in patients with AML (Nakanishi et al., 2003), and its overexpression was higher after relapse of AML; hence, ABCG2 was suggested to act as a prognostic indicator for this disease (Benderra et al., 2004; Uggla et al., 2005). The role of ABCG2 in the resistant phenotype of solid tumors is less clear, although ABCG2 overexpression has been described in several tumor types (Diestra et al., 2002; Turner et al., 2006). ABCG2 conferred resistance to flavopiridol in patients with AML (Nakanishi et al., 2003), and its overexpression was higher after relapse of AML; hence, ABCG2 was suggested to act as a prognostic indicator for this disease (Benderra et al., 2004; Uggla et al., 2005). The role of ABCG2 in the resistant phenotype of solid tumors is less clear, although ABCG2 overexpression has been described in several tumor types (Diestra et al., 2002; Turner et al., 2006). Of interest, ABCG2 expression was rapidly increased in hepatoblastoma in response to certain chemotherapeutic drugs (Vander Borght et al., 2008), reminiscent of the induction of P-glycoprotein observed after treatment of metastatic sarcoma with doxorubicin (Abolhoda...
et al., 1999). ABCG2 is also expressed in adult normal tissues, including the intestine, colon, placenta, and blood-brain barrier, where it plays a role in restricting the oral bioavailability and pharmacokinetics of its substrates and probably plays a critical role in the homeostasis of endogenous compounds such as heme, porphyrins, riboflavin, and estrogens (Grube et al., 2007; Krishnamurthy et al., 2007; van Herwaarden et al., 2007; Ni et al., 2010).

Of interest is the observation that ABCG2 appears to play a critical role in both normal cells and cancer stem cells (CSCs); indeed, the observation that ABCG2 is often enriched in normal stem cells and CSCs relative to their more mature progeny has led to the proposed use of ABCG2 as a universal stem cell marker (Zhou et al., 2001; Bunting, 2002). The function of ABCG2 in CSCs is not well understood; however, some studies suggest that it is required for maintaining the “stemness” of the population, perhaps by enhancing proliferation and/or decreasing differentiation potential. Although this requires further investigation, there is ample evidence supporting a role for ABCG2 in protecting both normal and cancer stem cells from various stressors, including cancer chemotherapeutic drugs (Dean et al., 2005; Krishnamurthy and Schuetz, 2006).

Given the pleiotropic roles of ABCG2 in drug disposition, drug resistance, and CSC survival, inhibition or down-regulation of ABCG2 may be a valid approach to reverse ABCG2-mediated drug resistance and to increase oral absorption of certain chemotherapeutic drugs. Moreover, the putative role of ABCG2 in maintenance of CSCs suggests that ABCG2 inhibitors, unlike other drug transporter inhibitors, may function to reduce or eliminate the CSC population.

In the current study, we have investigated the use of xanthine derivatives as ABCG2 inhibitors. Xanthines are common in the human diet, where they are consumed in the form of nutrients, stimulants, and drugs. Caffeine (1,3,7-trimethylxanthine), the best known, best-studied, and most widely

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![Fig. 1: Caffeine down-regulated ABCG2 expression in cancer cell lines.](image-url)

**a** and **b**, ABCG2 protein levels were decreased by caffeine in a dose-dependent manner in ABCG2-overexpressing MCF-7/MX100 cells. **c** and **d**, caffeine (14 mM) reduced ABCG2 protein levels in a time-dependent manner, with maximum reduction achieved by 24 h in MCF-7/MX100 cells (c) and BeWo cells (d). **e** and **f**, caffeine-mediated down-regulation of ABCG2 protein was reversible. MCF-7/MX100 cells (e) and BeWo cells (f) were treated with caffeine for 24 h and then were washed with drug-free medium and incubated for the times indicated before lysate preparation and Western blot analysis. Graphs display quantifications of the Western blot using Quantity One software (Bio-Rad Laboratories, Hercules, CA). In all cases, levels of ABCG2 protein were normalized to a GAPDH loading control and to the untreated sample.
consumed xanthine, is a purine alkaloid, with a variety of pharmacological effects. It is used in the treatment of migraines and postprandial hypotension and obesity and as a respiratory stimulant in neonates (Sawynok, 1995). The effect of caffeine on cell cycle transition is sensitization of some human tumors to ionizing radiation and alkylating agents such as nitrogen mustard and cisplatin (Sarkaria et al., 1999). Analogous to caffeine, many other xanthine-related heterocyclins with enhanced potency and selectivity toward specific biological targets have provided powerful tools for research and potential therapeutic agents for intervention in a variety of diseases (Daly, 2007). For example, theophylline, a caffeine metabolite found in tea, is used for the treatment of asthma and has anti-inflammatory properties. Dyphylline [7-(2,3-dihydroxypropyl)-theophylline] is used as a bronchodilator in the treatment of asthma, chronic bronchitis, and emphysema. Additional xanthine derivatives are currently in preclinical and clinical studies (McCarty et al., 2002).

We now show that caffeine and several related xanthines can dramatically decrease ABCG2 protein in multiple cell lines by targeting this cell surface transporter for internalization and lysosomal degradation. This down-regulation has functional consequences. Exposure to caffeine increased intracellular accumulation of the ABCG2 substrate mitoxantrone (MX), resulting in a 10-fold increased sensitivity of tumor cells to the cytotoxic effects of this agent. These findings suggest that xanthines can be developed as combination therapy to improve the efficacy of drugs that are ABCG2 substrates.

**Materials and Methods**

**Cell Culture and Chemicals.** BeWo cells were purchased from American Type Culture Collection (Manassas, VA) and maintained in F-12K medium (American Type Culture Collection) supplemented with 10% heat-inactivated fetal bovine serum (Atlanta Biologicals, Norcross, GA) at 37°C in 5% (v/v) CO₂. MCF-7 parental cells and the MX-resistant subline MCF-7/MX100 were kindly provided by Dr. Susan Bates (National Institutes of Health, Bethesda, MD). Both cell lines were grown in improved minimum essential medium (Invitrogen, Carlsbad, CA) containing 2 g/l glucose, 2 mM L-glutamine, 1 mM sodium pyruvate, and 10% fetal bovine serum at 37°C in 5% (v/v) CO₂. MCF-7/MX100 cells were maintained in the presence of 100 nM MX (Honjo et al., 2001). Caffeine and caffeine analogs [theophylline, pentoxifylline, dyphylline, paraxanthine, theobromine, 7-β-hydroxyethyltheophylline, and 7-methoxanthine], ammonium chloride, and leupeptin were purchased from Sigma-Aldrich (St. Louis, MO).

**Protein Analysis.** For Western blot analysis, cells were washed twice with ice-cold phosphate-buffered saline and lysed in radiomunoprecipitation assay buffer (50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 0.1% Triton X-100, 0.1% SDS, and 1% deoxycholate, sodium salt) plus protease inhibitor cocktail (Roche Diagnostics, Indianapolis, IN) and 100 mg/ml phenylmethylsulfonyl fluoride. Protein concentrations were determined using the BCA Protein Assay (Thermo Fisher Scientific, Waltham, MA). Equal amounts of total protein (5–15 g) were analyzed by 8% SDS-polyacrylamide gel electrophoresis followed by immunoblotting using mouse monoclonal antibody (clone BXP-21) against ABCG2 (1:1000; Kamiya Biomedical, Thousand Oaks, CA) or rabbit monoclonal antibody against GAPDH (1:1000; Cell Signaling). The secondary antibody was either horseradish peroxidase-conjugated goat anti-mouse IgG (1:2500; GE Healthcare, Chalfont St. Giles, Buckinghamshire, UK) or horseradish peroxidase-conjugated goat anti-rabbit IgG (Santa Cruz Biotechnology, Inc., Santa Cruz, CA). Immunoreactive bands were visualized using an enhanced chemiluminescent system (Thermo Fisher Scientific) according to the manufacturer’s recommendations.

For immunocytochemical analysis, cells grown on glass coverslips were washed three times with PBS, fixed in 4% paraformaldehyde solution, and permeabilized in 0.2% Triton X-100 for 10 min. Cells were washed with PBS three times at each interval. Coverslips were then treated with ice-cold phosphate-buffered saline, fixed in 4% paraformaldehyde solution, and permeabilized in 0.2% Triton X-100 for 10 min. Cells were washed with PBS three times at each interval. Coverslips were then treated with the following antibodies: (a) antisera against ABCG2 (1:50; Kamiya Biomedical, Thousand Oaks, CA); (b) antiserum against GAPDH (1:5000; Cell Signaling); (c) horseradish peroxidase-conjugated goat anti-mouse IgG (1:2500; GE Healthcare, Chalfont St. Giles, Buckinghamshire, UK). Immunoreactive bands were visualized using an enhanced chemiluminescent system (Thermo Fisher Scientific) according to the manufacturer’s recommendations.

**Fig. 2.** Methylxanthines and caffeine analogs down-regulated ABCG2 expression. a. BeWo cells were treated with caffeine (14 mM), theophylline (14 mM), dyphylline (14 mM), theobromine (2.5 mM), paraxanthine (1.5 mM), 7-β-hydroxyethyltheophylline (14 mM), and 7-methoxanthine (1 mM) for 24 h, and ABCG2 protein levels were analyzed by Western blot. GDPH was used as a loading control. b. Theophylline decreased ABCG2 protein levels potently and dose dependently. BeWo cells were treated with theophylline at concentrations for 24 h. GDPH was used as a loading control. c. Quantification of the effect of theophylline on ABCG2. ABCG2 protein levels were normalized to the GDPH loading control, and the percentage of protein was determined relative to the levels in untreated cells.

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were incubated with 2% bovine serum albumin in 0.1% Triton X-100 PBS buffer at room temperature for 1 h, followed by incubation with monoclonal ABCG2 antibody BXP-21 (1:250) in 0.1% Triton X-100 in a humid chamber, washed three times with PBS, and then incubated with Alexa Fluor 488-conjugated goat anti-mouse IgG at 37°C for 1 h. Coverslips were mounted, sealed on glass slides with 4,6-diamidino-2-phenylindole mounting medium, and observed using an Eclipse TE2000-U confocal microscope (Nikon, Melville, NY).

Efflux Assays. Efflux assays were performed as described previously (Robey et al., 2001) with minor modifications. In brief, cells were collected and suspended in complete medium alone or complete medium containing 500 nM BODIPY-prazosin (an ABCG2 substrate; Invitrogen) with or without 10 μM fumitremorgin C (FTC), a known ABCG2 inhibitor (Rabindran et al., 2000), and incubated at 37°C in 5% CO₂ for 30 min. The incubations were stopped with the addition of 4 ml of ice-cold PBS, and cells were washed three times with ice-cold PBS and incubated for 1 h at 37°C in 5% CO₂ in complete medium with or without 10 μM FTC. Cells incubated in blank medium were used as the control for cell autofluorescence. Dead cells and debris were gated out for each treatment by propidium iodide staining. Fluorescence was visualized using a flow cytometer (FC500; Beckman Coulter, Fullerton, CA) with a 488-nm argon laser and 530-nm band-pass filter.

Cytotoxicity and Cell Death Assays. MX cytotoxicity was assayed as follows. BeWo or MCF-7/MX100 cells were plated in 96-well plates at a density of 5000 cells/well. Twenty-four hours after plating, cells were treated with or without caffeine for another 24 h. After caffeine pretreatment, the medium was replaced with medium containing increasing concentrations of MX, and cells were incubated for 72 h at 37°C. Cell death was assayed using CellTiter 96 AQ solution according to the manufacturer’s instructions (Promega, Madison, WI). Cytotoxicity was assessed by monitoring the absorbance at 490 nm using a Multiskan Spectrum microplate reader (Thermo Fisher Scientific). The IC₅₀ value was defined as the drug concentration resulting in 50% cell death. Both the fitted sigmoidal dose-response curve and IC₅₀ were calculated using Prism 4.

The Guava EasyCyte flow cytometry analysis system (Guava Technologies, Hayward, CA) was used to determine percentage of apoptotic cells. Assays were conducted according to the manufacturer’s instructions. In brief, total cells were collected and washed with ice-cold PBS; then 5 μl of annexin V-phycoerythrin, a marker for early apoptosis, and 5 μl of 7-amino-actinomycin, a cell-impermeant dye indicating late apoptosis or dead cells (PCA-96 Nexin Kit; Guava Technologies), were added to cell suspensions. After a 20-min incubation and thorough mixing, samples were analyzed on a Guava flow cytometer.

Results

Xanthines Down-Regulate ABCG2 Expression in Cancer Cells. Xanthines, particularly methylxanthines, are found in more than 60 plant species and are among the most widely consumed substances in our diet. To evaluate the

![Fig. 3. Caffeine (Caff) decreased membrane-localized ABCG2 and altered its cellular localization.](https://example.com/f3.png)
effect of xanthines on the expression of the drug transporter ABCG2 in tumor cells, we used the MX-resistant human breast cancer cell line MCF-7/MX100, which is resistant to MX relative to its MCF-7 parental cells by virtue of its over-expression of ABCG2 (Fig. 1, a and c) and the human placental choriocarcinoma cell line BeWo (Fig. 1, b and d), which has intrinsic expression of functional ABCG2. Exposure to methylxanthine caffeine reduced ABCG2 protein levels in a dose-dependent (Fig. 1, a and b) and time-dependent (Fig. 1, c and d) manner. Effects could be seen as early as 6 h after treatment with the maximum decrease (~60–75%) achieved by 24 h (IC_{50} = 0.4 mM (BeWo) and 2.5 mM (MCF-7/ MX100)). This effect was reversible; when caffeine was removed and replaced with fresh medium 24 h after treatment, ABCG2 levels gradually recovered, reverting to normal levels within the next 24 h (Fig. 1, e and f). Of note, although some caffeine-induced cell cycle arrest was observed in MCF-7/ MX100 cells, this arrest was not observed in BeWo cells (data not shown), yet the degree of down-regulation of ABCG2 by caffeine in both cell lines was similar. Therefore, it does not appear that caffeine-induced cell cycle inhibition plays a role in the regulation of ABCG2 levels by xanthines. Moreover, caffeine did not have a general effect on membrane protein stability, because there was minimal impact on the expression of epidermal growth factor receptor and no apparent effect on Na^+/K^+-ATPase (data not shown).

A number of naturally occurring xanthines are structurally similar to caffeine and share some of its pharmacological effects. To determine the generality of the effect of caffeine on ABCG2 expression, we analyzed ABCG2 protein levels after exposure to several of these caffeine analogs. As shown in Fig. 2a, among the analogs we tested, theophylline, a direct metabolite of caffeine, was the most potent compound in terms of down-regulating ABCG2 protein levels (Fig. 2b); 0.2 mM of theophylline was sufficient to reduce ABCG2 protein levels by 50%, and the maximum decrease (90%) was achieved by 1.75 mM. Other compounds, such as paraxanthine and 7-β-hydroxyethyltheophylline, did not significantly affect ABCG2 expression levels. The structure-activity relationship of a large group of xanthines is currently under investigation.

Caffeine Induces Internalization and Lysosomal Degradation of ABCG2. To determine the molecular basis for the down-regulation of ABCG2, we first considered the possibility that caffeine affected the synthesis or processing of ABCG2 mRNA. Indeed, we have previously shown that caffeine can induce the alternative splicing of a subset of genes (Shi et al., 2008). However, quantitative polymerase chain reaction analysis of ABCG2 RNA from untreated and caffeine-treated cells revealed no impact on either ABCG2 RNA levels or splicing in either MCF-7/MX100 or BeWo cells (data not shown), suggesting that the effect of caffeine occurred at the level of protein synthesis or degradation.

To examine the fate of ABCG2 protein in the presence of caffeine, cells were subjected to immunofluorescence staining after caffeine treatment. As shown in Fig. 3a, both MCF-7/ MX100 and BeWo cells exhibited intense membrane staining and some cytoplasmic staining in the presence of an ABCG2-specific antibody. After treatment with high levels of caffeine, there was a significant decrease in the total fluorescence intensity in both cell lines, and membrane-localized ABCG2 was largely attenuated. At lower concentrations (0.8 mM), more intracellular staining was observed compared with that in the untreated cells, suggesting internalization of ABCG2 protein in the presence of caffeine. Again, this effect was time-dependent (Fig. 3b), with internalization apparent after 4 h of caffeine treatment and peaking between 10 and 24 h after caffeine exposure. By 24 h, little ABCG2-related staining was observed in the cell, suggesting that caffeine had induced the internalization and subsequent degradation of ABCG2 protein.

It has been reported previously that wild-type ABCG2 is primarily degraded through the lysosome-mediated pathway, although misfolded ABCG2 is degraded by ubiquitin-mediated proteasomal machinery (Nakagawa et al., 2009). To determine whether caffeine or its analog, theophylline, enhanced ABCG2 lysosomal degradation, we examined the effect of lysosome inhibitors on caffeine-mediated ABCG2 down-regulation. Figure 4 shows a Western blot analysis of ABCG2 expression after methylxanthine exposure in the presence or absence of the lysosomal inhibitors ammonium chloride (NH_4Cl) (Fig. 4a) or leupeptin (Fig. 4b). Ammonium chloride, which inactivates lysosomal enzymes by neutralizing the luminal pH of the lysosome, prevented caffeine and theophylline from decreasing ABCG2 protein levels (Fig. 4a). This effect could be seen as early as 4 h after treatment (data not shown). A similar result was observed when the lysosomal inhibitor leupeptin was used (Fig. 4b).

To confirm this result, immunofluorescence was determined in cells treated with caffeine in the absence or presence of either NH_4Cl or leupeptin (Fig. 4, c and d). As shown previously, caffeine caused an overall decrease in the ABCG2 fluorescence signal, and the ratio between the membrane and cytosol signals remained similar to that in the untreated samples. In the presence of NH_4Cl, a decrease in ABCG2 signal intensity was still observed after caffeine treatment, but there was a marked increase in cytosol staining, particularly around the perinuclear region. This intracellular accumulation of ABCG2 was even more prominent after treatment with the specific lysosomal protease inhibitor leupeptin. These results suggest that caffeine induced the internalization of ABCG2 protein, probably through endocytosis, and directed it to the lysosomal degradation pathway, which was blocked by NH_4Cl or leupeptin treatment.

Caffeine Potentiates Cytotoxicity of the ABCG2 Substrate Mitoxantrone. To determine the functional impact of the down-regulation of ABCG2 by methylxanthines, we determined whether caffeine could affect the accumulation of an ABCG2 substrate, using a flow cytometry assay (Robey et al., 2001). MCF-7/MX100 (a high ABCG2 expresser) and its parental cell line MCF-7 (a low ABCG2 expresser) were treated with caffeine for 24 h, and then cells were collected...
Fig. 5. Caffeine inhibited efflux of an ABCG2 substrate and sensitized cells to the chemotherapeutic agent mitoxantrone. a, MCF-7/MX100 cells were treated with either 14 mM caffeine or 10 μM FTC 24 h before assay of cells for their ability to efflux the ABCG2-specific substrate BP. Both caffeine (red dashed line) and FTC treatments (black dotted line) increased the mean fluorescence intensity compared with that for untreated cells (black solid line).
and incubated with the ABCG2-specific fluorescence substrate Bodipy-prazosin (BP). After incubation, cells were grown in fresh media without substrate, and the intracellular concentration of BP was determined as a measure of ABCG2 expression/function. The highly potent but toxic ABCG2 inhibitor, FTC, was used as a positive control. As shown in Fig. 5a, accumulation of BP in the highly expressing MCF-7/MX100 cells was markedly increased in the presence of caffeine, suggesting that caffeine inhibited transport of this substrate by ABCG2. In contrast, there was little effect of caffeine on BP accumulation in the low-expressing MCF-7 parental cell line (Fig. 5b), confirming that the effect of caffeine on BP uptake was specific for ABCG2.

MX is used in the treatment of certain cancers, including metastatic breast cancer and acute myeloid leukemia, for which it has been shown to induce programmed cell death or apoptosis (Kluza et al., 2004). Therefore, to further confirm the functional effect of caffeine on ABCG2 and to investigate the potential of using methylxanthines as chemosensitizing agents to treat ABCG2-mediated multidrug resistance, we analyzed the effect of caffeine on MX-induced apoptosis in MCF7/MX100 cells. Cells were pretreated with caffeine for 24 h to achieve maximum down-regulation of ABCG2 protein and then were treated with 10 or 100 μM MX for an additional 24 h. The apoptotic status of the cells was determined using annexin staining and flow cytometry. As shown in Fig. 5c, compared with untreated cells, treatment with caffeine alone (7 mM) induced relatively little apoptosis (1.6%), whereas MX alone caused a dose-dependent apoptotic effect. However, pretreatment of cells with caffeine before MX exposure (10 μM) increased MX-induced apoptosis ~3-fold and shifted the IC50 by ~10-fold (IC50 = 946.5 ± 1.71 μM for MX alone and 81.4 ± 1.31 μM for caffeine and MX) (Fig. 5d). Parental MCF-7 cells were intrinsically more sensitive to MX (IC50 = 3.233 ± 1.84 μM) because of their relatively low levels of ABCG2; as expected, caffeine had little effect on MX cytotoxicity in MCF-7 cells (IC50 = 2.174 ± 1.44 μM for caffeine-treated cells), again supporting the hypothesis that caffeine is sensitizing cells to MX by down-regulating ABCG2 expression.

**Discussion**

In the present study, we show that xanthines, including the widely consumed methylxanthine caffeine and the respiratory therapeutic drug theophylline, can down-regulate the MDR membrane protein ABCG2 by inducing its translocation and subsequent lysosomal degradation. As a consequence of this down-regulation, xanthines inhibited efflux of an ABCG2 substrate and sensitized drug resistance breast cancer cells to mitoxantrone, an antineoplastic agent commonly used to treat acute myeloid leukemia, metastatic breast cancer, and non-Hodgkin’s lymphoma.

Previous studies have suggested that caffeine can modulate cancer drug sensitivity. The many cellular effects attributed to caffeine have implicated a number of pathways/targets.

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**Table 1**

<table>
<thead>
<tr>
<th>Compound</th>
<th>IC50 (μM)</th>
<th>IC50 (μM)</th>
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<tbody>
<tr>
<td>FTC</td>
<td>19.6 ± 0.4</td>
<td>19.6 ± 0.4</td>
</tr>
<tr>
<td>Caffeine</td>
<td>946.5 ± 1.71</td>
<td>81.4 ± 1.31</td>
</tr>
<tr>
<td>MX</td>
<td>104.3 ± 2.17</td>
<td>104.3 ± 2.17</td>
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Results are expressed as means ± S.D.; n = 3. Cells were treated with 10 or 100 μM MX for 24 h, and the percentage of apoptotic cells was determined. Results are expressed as means ± S.D.; n = 8.
are currently using caffeine and caffeine analogs as 1) tools to further dissect the mechanism underlying regulation of degradation of ABCG2 protein and 2) as lead compounds for the identification/development of more potent/less toxic agents. Regarding this latter point, we have recently investigated the effect of a synthetic xanthine, 1,3-dipropyl-8-cyclopentylxanthine (DPCPX), on ABCG2 levels. DPCPX is in clinical trials for the treatment of cystic fibrosis; single doses up to 1000 mg were tested with no apparent toxicity (McCarty et al., 2002).

Of importance, DPCPX was able to down-regulate ABCG2 levels to the same degree as caffeine, but at concentrations (5–50 μM, data not shown) that are more likely to be achievable in vivo. Animal studies are planned to evaluate the antitumor effect of this agent in combination with chemotherapeutic drugs that are ABCG2 substrates.

One complication of most of the chemotherapeutic regimens currently in use is that their targets are also expressed in normal cells, which is the case with ABCG2, which is found in normal stem cells, endothelial barriers, and excretory cells of the liver, kidney, and intestines (Robey et al., 2009). This situation presents both advantages and disadvantages. On the one hand, down-regulation of ABCG2 in secretory tissues may enhance drug bioavailability; in contrast, decreased ABCG2 levels may also lead to greater toxicity of chemotherapeutic agents in some cell types. Thus, as is the case with most cancer-directed agents, unwanted effects of candidate xanthines on normal cells/tissues will need to be considered as in vivo studies progress. It is also important to note that ABCG2 is expressed in cancer stem cells, in which it may play a role in stemness maintenance and proliferation. Thus, down-regulation of ABCG2 may prove useful for the elimination of this oftentimes drug-resistant subpopulation.

Studies addressing these questions are underway.

It has been reported previously that ABCG2 can be degraded through both proteasome- and lysosome-mediated pathways. Mutated or misfolded newly synthesized ABCG2 protein is rapidly eliminated via ubiquitin-mediated proteasome degradation before reaching the membrane (Nakagawa et al., 2009). In contrast, membrane-associated wild-type ABCG2 is normally degraded through the lysosome (Wakabayashi et al., 2007). How xanthines induce the translocation and lysosomal degradation of ABCG2 is not yet clear. One possibility is that xanthines directly interact with ABCG2 and negatively regulate its stability; indeed, it has been reported that some nucleosides and nucleoside analogs can bind to the drug pocket of ABCG2 (de Wolf et al., 2008), although there is no evidence that nucleobases can do the same. Of note, we have observed that some of the xanthines tested did not mimic the effect of caffeine on ABCG2. For example, theophylline, dyphylline, and theobromine actively reduced the protein level of ABCG2, whereas 7-(β-hydroxyethyl)theophylline, paraxanthine, and 7-methylxanthine did not have a significant impact on its expression. The facts that the size of the side chain modification on the xanthine ring did not correlate with activity and that caffeine is closely related to a nucleobase rather than a nucleoside reduce the likelihood that xanthines act by direct binding to ABCG2. Although we have not yet ruled out this possibility, our current hypothesis favors a model whereby active xanthines affect/induce a signaling event that triggers the translocation and degradation of ABCG2, perhaps via a regulated process such as ubiquitin-mediated endocytosis. Recent studies have demonstrated the importance of monoubiquitination as a critical signal for selective degradation (d’Azzo et al., 2005). For example, the ion transporter epithelial Na+ channel can be recognized by the E3 ligase Nedd4-2 for ubiquitination in response to metabolic stress, triggering its subsequent endocytosis and lysosomal degradation (Bhalla et al., 2006). Although we have yet to determine whether a caffeine-induced signaling pathway regulates ABCG2 degradation or which pathway is involved, a previous study demonstrating that inhibition of phosphatidylinositol 3-kinase activity led to the intracellular translocation of ABCG2 and subsequent decrease in the efflux of ABCG2 substrates (Bleau et al., 2009) supports this hypothesis and suggests an avenue for further investigation. Clarifying the mechanisms by which xanthines induce lysosomal degradation of ABCG2 will inform the development of more specific ABCG2 modulators.

Taken together, our data demonstrate that a class of xanthine derivatives, including many already in clinical use, can modulate the expression of ABCG2 and sensitize cells to chemotherapeutic agents that are substrates for this transporter. This result defines a new function for this large family of compounds and identifies a new class of agents that may be useful in sensitizing tumors to the cytotoxic effects of chemotherapeutic drugs. As we further dissect the mechanism/pathway through which xanthines induce the degradation of ABCG2, it is likely that more specific and efficacious inhibitors will be identified.

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Authorship Contributions

Participated in research design: Ding, Shi, and Scotto.
Performed experiments: Ding and Pabon.
Wrote or contributed to the writing of the manuscript: Ding and Scotto.

References

tance-associated protein BCRP/PMXR/ABCP/ABCG2 in human tumours detected...


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