Resveratrol Sensitizes Acute Myelogenous Leukemia Cells to Histone Deacetylase Inhibitors through Reactive Oxygen Species-Mediated Activation of the Extrinsic Apoptotic Pathway

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ABSTRACT

Histone deacetylase inhibitors (HDACIs) activate the prosurvival nuclear factor-κB (NF-κB) pathway by hyperacetylating RelA/p65, whereas the chemopreventive agent resveratrol inhibits NF-κB by activating the class III histone deacetylase Sirt1. Interactions between resveratrol and pan-HDACIs (vorinostat and panobinostat) were examined in human acute myelogenous leukemia (AML) cells. Pharmacologically achievable resveratrol concentrations (25–50 μM) synergistically potentiated HDACI lethality in AML cell lines and primary AML blasts. Resveratrol antagonized RelA acetylation and NF-κB activation in HDACI-treated cells. However, short hairpin RNA Sirt1 knockdown failed to modify HDACI sensitivity, which suggests that factors other than or in addition to Sirt1 activation contribute to resveratrol/HDACI interactions. These interactions were associated with death receptor 5 (DR5) up-regulation and caspase-8 activation, whereas cells expressing dominant-negative caspase-8 were substantially protected from resveratrol/HDACI treatment, which suggests a significant functional role for the extrinsic apoptotic pathway in lethality. Exposure to resveratrol with HDACI induced sustained reactive oxygen species (ROS) generation, which was accompanied by increased levels of DNA double-strand breaks, as reflected inγH2AX and comet assays. The free radical scavenger Mn(III)tetrakis(4-benzoic acid)porphyrin chloride blocked ROS generation, DR5 up-regulation, caspase-8 activation, DNA damage, and apoptosis, which indicates a primary role for oxidative injury in lethality. Analyses of cell-cycle progression and 5-ethyl-2′-deoxyuridine incorporation through flow cytometry revealed that resveratrol induced S-phase accumulation; this effect was abrogated by HDACI coadministration, which suggests that cells undergoing DNA synthesis may be particularly vulnerable to HDACI lethality. Collectively, these findings indicate that resveratrol interacts synergistically with HDACIs in AML cells through multiple ROS-dependent actions, including death receptor up-regulation, extrinsic apoptotic pathway activation, and DNA damage induction. They also raise the possibility that S-phase cells may be particularly susceptible to these actions.

Introduction

Histone deacetylase inhibitors (HDACIs) represent a class of epigenetic agents that regulate gene expression by modifying chromatin structure. HDACIs promote histone acetylation, which leads to a more-relaxed configuration conducive to the transcription of genes implicated in differentiation and cell death (Bolden et al., 2006). However, HDACIs also kill transformed cells through alternative mechanisms, including induction of oxidative injury (Ruefli et al., 2001), interference with DNA repair machinery (Subramanian et al., 2005), and...
up-regulation of death receptors (Nebbioso et al., 2005), among others. The pan-HDACi vorinostat has been approved for the treatment of cutaneous T-cell lymphomas (Grant et al., 2007), and initial suggestions of HDACI activity in acute myelogenous leukemia (AML) were reported (Garcia-Manero et al., 2008).

HDACs are subdivided into four groups, as follows: class I, HDACs 1 to 3 and 8 (analogous to yeast Rpd); class II, HDACs 4 to 7, 9, and 10 (analogous to yeast Hda1); class III, NAD+－dependent sirtuins 1 to 7; class IV, HDAC11 (Glozak and Seto, 2007). Sirtuins have been implicated in the regulation of tumor initiation, progression, and chemoresistance; consequently, agents that modify sirtuin activity are currently a subject of interest for cancer therapy (Liu et al., 2009). Resveratrol is a naturally occurring polyphenolic compound extracted from grapes, and clinical trials are underway to explore its potential among patients with cardiovascular diseases or diabetes mellitus (Baur and Sinclair, 2006). Resveratrol has been associated with minimal toxicity, and plasma levels of >300 μM are achievable and well tolerated among humans (Howells et al., 2011). In preclinical studies, resveratrol exhibited activity against various malignant cell types, including AML (Tsan et al., 2002), through diverse mechanisms such as inhibition of IKK and NF-κB (Holmes-McNary and Baldwin, 2000), induction of oxidative injury (Low et al., 2010), and autophagy (Puissant et al., 2010). Resveratrol was shown to act as a Sirt1 agonist (Milne et al., 2010), although evidence indicating that this may involve indirect actions has emerged (Pacholec et al., 2010).

In addition to histones, HDACIs promote the acetylation of diverse nonhistone proteins, including transcription factors such as NF-κB (Glozak et al., 2005). In previous studies, we reported that inhibitors of the NF-κB signaling pathway, including IKK and proteasome inhibitors, markedly increased the activity of HDACIs against myeloid leukemia cells (Dai et al., 2005, 2011b). Among other actions, these agents potently block RelA deacetylation which plays an important role in DNA binding and transactivation (Dai et al., 2005). It is known that, like class I HDACs (e.g., HDAC3), the class III HDAC Sirt1 deacetylates RelA and inactivates NF-κB (Chen et al., 2005). However, pan-HDACIs such as panobinostat (LBH-589) and vorinostat fail to target class III HDACs (Xu et al., 2007). Furthermore, sirtuin agonists were shown to inhibit NF-κB function by antagonizing RelA acetylation (Yeung et al., 2004; Dai et al., 2005). These observations raised the possibility that resveratrol might sensitize leukemia cells to HDACIs. To address this question, we examined interactions between resveratrol and two clinically relevant pan-HDACIs (vorinostat and panobinostat) in human myeloid leukemia cells. We report that resveratrol synergistically potentiated HDACI activity against myeloid leukemia cells in association with ROS-dependent activation of the extrinsic apoptotic pathway.

Materials and Methods

Cells and Cell Culture. U937 and MV-4-11 (bearing internal tandem duplications of FLT3) human leukemia cells were obtained from American Type Culture Collection (Manassas, VA) and were maintained in RPMI 1640 medium containing 10% fetal bovine serum, as described previously (Maggio et al., 2004). U937 cells were stably transfected with dominant-negative caspase-8 or empty vector as described previously (Rosato et al., 2007).

Bone marrow or peripheral blood samples (>65% blasts) were obtained, with informed consent, from patients with histologically documented AML who were undergoing routine diagnostic procedures, in accordance with the Declaration of Helsinki and with institutional review board approval (Virginia Commonwealth University Institutional Review Board no. HM 12517). Mononuclear cells were isolated through centrifugation at 400g for 30 min with Histopaque-1077 (Sigma-Aldrich, St. Louis, MO). Cell viability was regularly >95%, and all samples consisted of >70% blasts. All experiments were performed at a density of 1 × 10⁶ cells/ml, as described previously (Dai et al., 2011b).

Drugs and Chemicals. Resveratrol (3,4',5-trihydroxy-trans-stilbene) and sodium azide were purchased from Sigma-Aldrich. Vorinostat (suberoylanilide hydroxamic acid) and panobinostat were provided by Merck (Whitehouse Station, NJ) and Novartis (East Hanover, NJ), respectively. Mn(III)tetrakis(4-benzoic acid)porphyrin chloride (MnTBP), a cell-permeable superoxide dismutase mimetic and peroxynitrite scavenger, was obtained from CalvinBiochem (San Diego, CA). The selective NOX1 inhibitor 2-acetylsphingohexaenone (ML171) was purchased from Millipore Corporation (Billerica, MA). Reagents were formulated in dimethylsulfoxide and stored at −20°C. Sodium azide was dissolved in sterile phosphate-buffered saline before use. Stock solutions were diluted with serum-free RPMI 1640 medium before use, to ensure that the final concentration of dimethylsulfoxide did not exceed 0.02%.

Experimental Format. Logarithmically growing cells (2.5–4.0 × 10⁵ cells/ml) were exposed to various concentrations of HDACIs in the presence or absence of resveratrol for the indicated periods (generally 24–48 h), after which cells were processed and assayed.

RNA Interference. SureSilencing shRNA plasmids (nontoxic resistance) from Origene Technologies, Inc. (Rockville, MD) were transfected with these constructs by using an Amaxa Nucleofector device with cell line-specific Nucleofector kit C (Amaxa GmbH, Cologne, Germany), according to the manufacturer’s instructions, and clones with down-regulated expression of Sirt1 were selected with 400 μg/ml G418.

Assessments of Cell Death and Mitochondrial Membrane Potential. Cells were double-stained for 20 min at 37°C with a solution containing 25 μM 7-aminoactinomycin D and 40 nM 3,3'-dihexyloxacarbocyanine iodide and then were analyzed through flow cytometry with a FACSscan flow cytometer (BD Biosciences, San Jose, CA), as described previously (Maggio et al., 2004). In some cases, apoptosis was evaluated with staining for 30 min at room temperature with annexin V-propidium iodide (PI) (BD Biosciences Pharmingen, Franklin Lakes, NJ) and then flow cytometry, as described previously (Rosato et al., 2010).

Measurement of ROS Production. Cells were treated with 20 μM 2,7'-dichlorodihydrofluorescein diacetate (Invitrogen, Carlsbad, CA) for 30 min at 37°C and then were analyzed through flow cytometry as described previously (Rosato et al., 2010).

Comet Assays. Single-cell gel electrophoresis assays were performed to assess both single- and double-strand DNA breaks in cells by using a comet assay kit (Trevigen, Gaithersburg, MD), according to the manufacturer’s instructions.

Western Blot Analyses. Whole-cell pellets were washed in phosphate-buffered saline and were lysed with loading buffer (Invitrogen) as described previously (Maggio et al., 2004). Alternatively, S-100 cytosolic fractions were prepared as described previously (Dai et al., 2005). Thirty micrograms of total protein for each condition were separated with a 4 to 12% Bis-Tris NuPage prefcast gel system (Invitrogen) and were electroblotted to nitrocellulose membranes. After incubation with the corresponding primary and secondary antibodies, blots were developed for enhanced chemiluminescence detection (PerkinElmer Life and Analytical Sciences, Waltham, MA). Primary antibodies were as follows: anti-caspase-8 and anti-PARP from Enzo Life Sciences (Plymouth Meeting, PA), anti-caspase-3.
from BD Biosciences Transduction Laboratories (Lexington, KY), anti-caspase-9, anti-DR5, anti-cytochrome c, and anti-p65 from BD Biosciences, anti-acetyl-NF-κB p65 (Lys310), anti-cleaved caspase-3, and anti-cleaved caspase-9 from Cell Signaling Technology (Danvers, MA), anti-Sirt1 from Santa Cruz Biotechnology (Santa Cruz, CA), anti-yH2AX (Ser139) from Millipore Corporation, anti-β-actin from Sigma-Aldrich, and anti-α-tubulin from Calbiochem. Secondary antibodies conjugated to horseradish peroxidase were obtained from KPL (Gaithersburg, MD).

**Immunoprecipitation Analyses.** RelA acetylation was evaluated through immunoprecipitation/Western blot analysis as described previously (Dai et al., 2005). Two hundred micrograms of nuclear extract for each condition were incubated overnight at 4°C with 1 μg of mouse monoclonal anti-NF-κB p65 antibody (Santa Cruz Biotechnology), with continuous shaking; 20 μl of Dynabeads (goat anti-mouse IgG; Invitrogen) for each condition were then added, and mixtures were incubated for an additional 4 h. After three washes with RIPA buffer, the bead-bound protein was eluted with vortex-mixing and boiling in 20 μl of 1× sample buffer. The samples were separated through SDS-polyacrylamide gel electrophoresis and were subjected to Western blot analysis as described above. Acetylated lysine-specific antibodies (Millipore Corporation) were used as primary antibodies.

**ELISA-Based NF-κB p65 Activity Analyses.** Nuclear protein for each condition were incubated overnight at 4°C with 1 μg of mouse monoclonal anti-NF-κB p65 antibody (Santa Cruz Biotechnology), with continuous shaking; 20 μl of Dynabeads (goat anti-mouse IgG; Invitrogen) for each condition were then added, and mixtures were incubated for an additional 4 h. After three washes with RIPA buffer, the bead-bound protein was eluted with vortex-mixing and boiling in 20 μl of 1× sample buffer. The samples were separated through SDS-polyacrylamide gel electrophoresis and were subjected to Western blot analysis as described above. Acetylated lysine-specific antibodies (Millipore Corporation) were used as primary antibodies.

**DNA Synthesis (S-Phase) Analyses.** Click-iT EdU CellCycle 488-red (7-aminoactinomycin D) assay kit (Invitrogen) was used, according to the manufacturer’s instructions, with flow cytometry to determine the S-phase population through incorporation of the thymidine analog EdU into genomic DNA during DNA synthesis. Alternatively, cells were double-stained with EdU-Alexa Fluor 488 and annexin V-allophycocyanin, after which flow cytometry was performed to determine the level of apoptosis (annexin V-positive) in the S-phase (EdU-positive) population.

**Statistical Analyses.** For analyses of cell death, ROS production, and NF-κB activity, values represent the mean ± S.D. of at least three separate experiments performed in triplicate. One-way analysis of variance of the Tukey-Kramer multiple-comparison test and Student’s two-tailed t test were performed. Analysis of synergism was performed through median dose effect analysis with Calcsyn ( Biosoft, Ferguson, MO).

**Results**

**Resveratrol Synergistically Interacts with HDACIs in Human Myeloid Leukemia Cells.** Coadministration of HDACIs with resveratrol leads to enhanced DNA damage, mitochondrial injury, and caspase-3, caspase-9, and caspase-8 activation. Whereas resveratrol or HDACIs (LBH-589 or vorinostat) administered individually for ≥16 h minimally induced γH2AX, an indicator of DNA double-strand breaks (Rosato et al., 2010), combined treatments sharply increased the levels of γH2AX expression in U937 and MV-4-11 cells (Fig. 2A). After drug treatment for 24 h, single- and double-strand DNA breaks in U937 cells were analyzed with comet assays. In this assay, denatured broken DNA fragments migrate out of the cell under the influence of an electric field, producing a comet tail, whereas undamaged DNA migrates more slowly and remains within the confines of the nucleus (Dai et al., 2008). As shown in Fig. 2C, coadministration of resveratrol with either LBH-589 or vorinostat resulted in clear increases in the numbers of comet-positive cells, compared with treatment with the agents individually. The appearance of DNA comet tails in cells exposed to resveratrol with LBH-589 or vorinostat occurred substantially before the induction of massive apoptosis, i.e., 8 h (Fig. 2C) versus 24 h (Fig. 1C). Furthermore, coadministration of resveratrol with LBH-589 or vorinostat resulted in early (4–8 h) and marked increases in the release of mitochondrial cytochrome c into the cytosol, compared with individual treatment (Fig. 2B). Exposure to resveratrol with either LBH-589 or vorinostat led to clearly increased cleavage/activation of caspase-3, caspase-9, and particularly caspase-8, which was accompanied by marked PARP degradation, in U937 (Fig. 2D) and MV-4-11 cells (Fig. 2E).

**Resveratrol Blocks HDACI-Mediated RelA Acetylation and NF-κB Activation.** Previous studies showed that interference with RelA/p65 acetylation and NF-κB activation, e.g., by IKK inhibitors, dramatically increased HDACI lethality in AML cells (Dai et al., 2005), which raises the possibility that a Sirt1 agonist such as resveratrol might act similarly by promoting Sirt1-mediated RelA deacetylation. To address this, the effects of resveratrol on RelA/p65 acetylation and NF-κB activation were examined in U937 cells exposed to HDACIs. As shown in Fig. 3A, ELISA-based NF-κB activity analysis of nuclear extracts showed that exposure to both LBH-589 and vorinostat induced p65-specific NF-κB activation, as reported previously (Dai et al., 2005), whereas this event was significantly blocked by resveratrol (P < 0.001 in each case). Western blot analyses of whole-cell lysates demonstrated that coadministration of resveratrol with either LBH-589 or vorinostat clearly decreased K310
acetylation of p65 (Fig. 3B). Immunoprecipitation analyses also revealed that resveratrol coadministration substantially decreased HDACI-induced p65 acetylation (Fig. 3C). Together, these findings indicate that resveratrol attenuates RelA/p65 acetylation and NF-κB activation in HDACI-treated AML cells, similar to effects observed with agents that directly target the NF-κB signaling pathway, such as IKK and proteasome inhibitors (Dai et al., 2005, 2011b).

Knockdown of Sirt1 Fails to Attenuate HDACI Lethality. To investigate the functional significance of Sirt1 perturbations in the regulation of HDACI lethality, U937 cells were stably transfected with constructs encoding shRNA specifically targeting human Sirt1 (shSirt1) or a scrambled sequence as a negative control (shNC). Two shSirt1 clones (designated 36 and 45) that displayed sharp reductions in Sirt1 expression, compared with control cells, were isolated (Fig. 3D, left). Contrary to expectations that Sirt1 knockdown would exert effects opposite those of the Sirt1 agonist resveratrol, both shSirt1 clones appeared slightly more sensitive (rather than resistant) to LBH-589 or vorinostat, compared with control cells, although differences did not achieve statistical significance (P > 0.05) (Fig. 3D, right). This finding suggests that mechanisms other than or in addition to Sirt1 activation by resveratrol contribute to the potentiation of HDACI antileukemic activity by this agent.

Resveratrol/HDACI Activity Proceeds through a ROS-Dependent Process. Earlier studies showed that both resveratrol (Schilder et al., 2009; Low et al., 2010) and HDACIs (Ruefi et al., 2001; Rosato et al., 2010) could trigger cell death through an oxidative injury-mediated process. Therefore, the effects of resveratrol with or without HDACIs on the generation of reactive oxygen species (ROS) were examined. Exposure of U937 cells to LBH-589 or vorinostat alone triggered modest increases in ROS levels, which decreased slightly after 24 h of exposure (Fig. 4A). In contrast, resvera-
trol alone induced sharp increases in ROS levels, which persisted over the 24-h exposure period, and coadministration of HDACIs did not increase ROS accumulation further (Fig. 4A). ROS generation in cells exposed to resveratrol with or without HDACIs was largely abrogated by the ROS scavenger MnTBAP (Fig. 4B), which led to substantial protection from resveratrol/HDACI-induced cell death (Fig. 4C) as well as loss of mitochondrial membrane potential (Supplemental Fig. 2A) and cleavage of caspase-3 and PARP (Supplemental Fig. 2B). It is known that NOXs share the capacity to trans- 
port electrons across the plasma membrane and to reduce oxygen to superoxide, thereby generating downstream ROS (Gianni et al., 2010). In this context, the selective NOX1 inhibitor ML171 (Gianni et al., 2010) was used to assess the functional role of NOXs in the lethality of the resveratrol/HDACI regimen. As shown in Fig. 4D, 1-h pretreatment with ML171 significantly prevented apoptosis induced by coadministra-
tion of resveratrol and LBH-589 (P = 0.0079, with versus without ML171) or vorinostat (P = 0.0419) in U937 cells. Catalases efficiently decompose H$_2$O$_2$ derived from superoxide (O$_2^-$), a reaction that is catalyzed by superoxide dismutase in cells (Mesquita et al., 2010). Therefore, two catalase inhibitors were used to analyze the functional role of catalase in cell death induced by resveratrol with or without HDACIs. It was found that the specific catalase inhibitor 3-amino-1,2,4-triazole (Mesquita et al., 2010) failed to attenuate the lethality of the resveratrol regimen (data not shown). One-hour pretreatment with sodium azide, a known catalase inhibitor and a donor of nitric oxide in the presence of catalase and H$_2$O$_2$ (Ogino et al., 2001), was reported
previously to prevent apoptosis through the extrinsic pathway (Kim et al., 1997) substantially prevented apoptosis induced by exposure to resveratrol and LBH-589 ($P = 0.0110$, with versus without azide) or vorinostat ($P = 0.0151$) in U937 cells (Fig. 4E).

The ROS Scavenger MnTBAP Blocks DR5 Up-regulation, Caspase-8 Cleavage, and DNA Damage in Cells Exposed to Resveratrol and HDACI. As shown in Fig. 2D, caspase-8 exhibited pronounced cleavage in U937 cells after exposure to resveratrol and HDACI, which indicated activation of the extrinsic apoptotic cascade. In this context, resveratrol by itself induced expression of DR5 (Fig. 5A), as reported for diffuse large B-cell lymphoma cells (Hussain et al., 2011). Coadministration of LBH-589 clearly enhanced resveratrol-mediated DR5 up-regulation (Fig. 5A), which was accompanied by increased caspase-8 cleavage (Fig. 5B) and γH2A.X expression (Fig. 5A). In the presence of MnTBAP, however, DR5 induction and caspase-8 cleavage/activation were entirely abrogated and γH2A.X expression (DNA damage) was largely prevented (Fig. 5, A and B). Together, these findings indicate that induction of ROS acts upstream of other lethal events (e.g., activation of the extrinsic death pathway and DNA damage) and thus plays a primary functional role in the antileukemic activity of the resveratrol/HDACI regimen.

There Is Evidence of a Functional Role for the Extrinsic Pathway in Resveratrol/HDACI Lethality. To assess more definitively the functional significance of activation of the extrinsic apoptotic pathway in this setting, U937 cells expressing dominant-negative caspase-8 were used (Rosato et al., 2007). Cleavage/activation of both caspase-8 and caspase-3 induced by resveratrol with LBH-589 or vorinostat was dramatically decreased in dominant-negative caspase-8–transfected cells, compared with their empty-vector–transfected counterparts (Fig. 5C). Consistent with these results, resveratrol/HDACI lethality was significantly attenuated in dominant-negative caspase-8–transfected cells ($P < 0.02$ or $P < 0.05$, compared with empty-vector–transfected control
Collectively, these findings indicate that ROS-dependent activation of the extrinsic pathway plays a significant functional role in the antileukemic activity of this regimen.

**Resveratrol Induces S-Phase Accumulation and Sensitizes Leukemia Cells to HDACIs.** Resveratrol has been shown to induce S-phase arrest in various tumor cell types, including leukemia cells (Bernhard et al., 2000). Therefore, the effects of resveratrol with or without HDACIs on cell-cycle progression were examined. As reported previously (Bernhard et al., 2000), exposure to resveratrol induced a marked increase in the S-phase cell population, in a time-dependent manner (Fig. 6A). In contrast, exposure to HDACIs (e.g., vorinostat) resulted in modest increases in the population of cells in the G0/G1 phase, with slight decreases in the S-phase population, but yielded little increase in the subdiploid population. Coadministration of vorinostat virtually eliminated the S-phase accumulation of cells induced by resveratrol, with a clear increase in the subdiploid (sub-G1) population (Fig. 6A; Supplemental Fig. 2C). In accord with these findings, flow cytometric analysis of EdU incorporation, which reflects DNA synthesis, demonstrated a pronounced increase in the EdU-positive S-phase cell population after exposure to resveratrol (e.g., 72%, compared with 46% in untreated cells), whereas coadministration of vorinostat with resveratrol led to the virtual disappearance of EdU-positive cells (e.g., to 4%) (Fig. 6B). Double-staining with EdU-Alexa Fluor 488 and annexin V-allophycocyanin demonstrated that administration of resveratrol with either LBH-589 or vorinostat induced apoptosis in both the EdU-positive (e.g., 58% or 44% for resveratrol plus LBH-589 or vorinostat, respectively, compared with 15% for resveratrol alone) (Fig. 6C) and EdU-negative populations. Together, these findings raise the possibility that resveratrol may arrest leukemia cells in the S-phase and that such...
cells may be particularly susceptible to HDACI-mediated lethality.

Discussion

Although resveratrol has traditionally been viewed as a chemopreventive agent (Baur and Sinclair, 2006), more-recent studies highlighted its capacity to induce cell death in neoplastic cells, including leukemia cells (Puissant et al., 2010). As for numerous natural products, the mechanisms through which resveratrol triggers transformed cell death are likely to be multifactorial (Athar et al., 2009). Resveratrol acts as an agonist of Sirt1 (Park et al., 2012), a member of the class III HDAC subfamily that pan-HDACIs fail to target (Xu et al., 2007), and exerts inhibitory effects on NF-κB, which contribute to its anticancer activity (Yeung et al., 2004; Dai et al., 2005). Resveratrol also was shown to prevent NF-κB activation and NF-κB-dependent gene expression through its inhibitory effects on IKKs (Holmes-McNary and Baldwin, 2000). In transformed cells, HDACIs activate NF-κB through RelA (Ser536) phosphorylation (Dai et al., 2011a) with an ataxia telangiectasia-mutated/NF-κB essential modifier-dependent mechanism (Rosato et al., 2010). Moreover, HDACIs induce RelA/p65 acetylation, which prevents nuclear export, while promoting DNA binding and transactivation (Chen et
These events lead to up-regulation of several antiapoptotic and antioxidant proteins, which decrease HDACI lethality, as well as activation of the stress-related c-Jun N-terminal kinase pathway (Dai et al., 2005). In this context, agents that prevent NF-κB activation, such as IKK (Dai et al., 2010) and proteasome inhibitors (Dai et al., 2011b), were shown to increase HDACI lethality markedly in malignant human hematopoietic cells, including leukemia cells. We hypothesized that resveratrol, like NF-κB–inhibitory agents, might enhance HDACI-mediated anti-leukemia activity. Consistent with this hypothesis, resveratrol clearly decreased p65 acetylation (e.g., K310) and NF-κB activation in HDACI-treated leukemia cells, with a sharp increase in cell death.

The observation that most HDACIs do not target class III HDACs, including Sirt1, provides a rationale for combining Sirt1 agonists such as resveratrol with HDACIs for cancer treatment. Although the Sirt1 agonist activity of resveratrol may involve an indirect mechanism (Pacholec et al., 2010), it was demonstrated that this agent exerts its effects by activating Sirt1, which negatively regulates p65 acetylation (an event essential for sustained NF-κB activation) (Chen et al., 2001). It was anticipated that, if Sirt1 activation by resveratrol (Milne et al., 2007) was responsible for potentiating HDACI lethality, then knockdown of Sirt1 would attenuate HDACI-mediated cell death, contrary to the effects of a Sirt1 agonist. Unexpectedly, Sirt1 knockdown failed to protect leukemia cells from HDACI-mediated lethality, which raises several possibilities. Other resveratrol actions (e.g., pro-oxidant activity or disruption of the cell cycle) (Athar et al., 2009) may be primarily responsible for the potentiation of HDACI lethality. Alternatively, aside from deacetylation of p65, Sirt1 actions may play predominantly cytoprotective roles (Chen et al., 2009). For example, Sirt1 is known to mediate deacetylation of numerous nonhistone substrates (e.g., Forkhead box proteins, p53, NF-κB, peroxisome prolif-
erator-activated receptor γ, and peroxisome proliferator-activated receptor γ coactivator 1α). In support of this notion, pharmacological Sirt1 antagonists were shown recently to promote chronic myelogenous leukemia stem cell death by increasing the acetylation and transcriptional activity of p53 (Li et al., 2012).

Although the chemopreventive actions of resveratrol may result from its antioxidant properties (de la Lastra and Villegas, 2007), resveratrol was shown to be a potent inducer of tumor cell oxidative injury (Chandra, 2009). In leukemia cells, oxidative injury also was shown to represent an important mechanism of HDACI lethality (Ruefli et al., 2001; Rosato et al., 2003; Petrucelli et al., 2011). The observation that ROS generation played a critical functional role in the actions of this combination regimen is not surprising. Resveratrol administered alone induced a pronounced sustained increase in ROS levels, but this was not accompanied by marked DNA damage (expression of γH2A.X or formation of comet tails) or cell death. Although HDACIs did not further increase ROS production in resveratrol-treated cells, coadministration sharply increased DNA damage and cell death. Given evidence that HDACIs interfere with DNA repair processes by hyperacetylating DNA repair proteins such as Ku70 (Subramanian et al., 2005) and down-regulating others (e.g., Rad50 or MRE11) (Lee et al., 2010), it is tempting to speculate that HDACIs amplify the lethal consequences of resveratrol-mediated oxidative injury and the resulting DNA damage (Subramanian et al., 2005).

Coadministration of resveratrol and HDACIs triggered the activation of caspases, particularly caspase-8, which indicates activation of the extrinsic apoptotic cascade (Scaffidi et al., 1998). In this context, HDACIs are known to up-regulate death receptors in human leukemia cells (Inginga et al., 2005), and resveratrol was shown to up-regulate death receptors, including DR5, in lymphoma cells (Hussain et al., 2011). Consistent with these observations, exposure of leukemia cells to resveratrol and HDACIs increased DR5 expression, compared with the effects of each agent administered individually. This finding provides a possible explanation for the marked activation of the extrinsic pathway produced by the resveratrol/HDACI regimen. Blockade of the extrinsic pathway with dominant-negative caspase-8 substantially decreased resveratrol/HDACI lethality, which demonstrates a significant functional role for the activation of this pathway in the antileukemia activity of this strategy. Both up-regulation of DR5 and activation of the extrinsic pathway were essentially abrogated by the ROS scavenger MnTBAP. It is interesting to note that similar phenomena were observed in human lymphoma (diffuse large B-cell lymphoma) cells exposed to resveratrol alone (Hussain et al., 2011). Together, these findings suggest that ROS-dependent induction of death receptors (e.g., DR5) and the resulting activation of the extrinsic apoptotic pathway represent an important mechanism underlying antileukemic synergism between these agents. Finally, although the ability of ML171 to protect cells from resveratrol/HDACI lethality implicates NOXs in this phenomenon, additional studies will be required to identify more definitively the sources of the ROS generated.

Resveratrol was reported to synchronize cells in the S-phase in association with the inhibition of cdc2 (Tyagi et al., 2005) or NF-κB (Estrov et al., 2003). The results of the present study demonstrated that resveratrol exposure sharply increased the S-phase population and the proportion of cells positive for EdU (a specific S-phase marker that reflects DNA synthesis) (Kramer and Wesierska-Gadek, 2009), in a time-dependent manner. Coadministration of HDACIs prevented resveratrol-induced S-phase accumulation and DNA synthesis, with a marked increase in the subdiploid fraction, which reflects apoptosis. One possible explanation for these findings is that cells exposed to resveratrol may accumulate in the S-phase, which is characterized by active DNA synthesis, and such cells may be particularly sensitive to HDACIs, as observed for the classic S-phase synchronizer hydroxyurea (Krämer et al., 2008). Previous findings that abrogation of G2/M arrest through blockade of induction of the endogenous cyclin-dependent kinase inhibitor p21CIP1 markedly sensitized leukemia cells to HDACIs (Rosato et al., 2002) are consistent with this notion. Alternatively, the cytoprotective effects of HDACI-mediated NF-κB activation, particularly its antioxidant actions attributable to induction of proteins such as superoxide dismutase 2 (Dai et al., 2005), may be critical for the survival of S-phase cells, which are known to be particularly vulnerable to oxidative injury (Ge et al., 2006). Finally, HDACIs kill transformed cells through diverse mechanisms, including up-regulation of proapoptotic proteins such as Bim (Zhao et al., 2005), induction of death receptors (Nebbioso et al., 2005), and promotion of DNA damage (Lee et al., 2010), among numerous others. Additional studies will be required to define the relative contributions of these actions to HDACI lethality toward S-phase cells.

In summary, the present studies demonstrate that resveratrol, administered at pharmacologically achievable concentrations (Howells et al., 2011), significantly increases HDACI lethality in human AML cells through oxidative injury-mediated activation of the extrinsic apoptotic pathway. The results also indicate that, although resveratrol does block HDACI-mediated NF-κB acetylation and activation, which were previously implicated in the potentiation of HDACI-mediated leukemic cell death (Dai et al., 2005; Rosato et al., 2010), actions other than or in addition to Sirt1 activation are likely to be responsible for the observed synergism. A hypothetical model summarizing mechanisms through which these agents may interact is shown in Fig. 7. According to this model, exposure of leukemia cells to resveratrol triggers multiple interrelated actions, including activation of Sirt1 (Milne et al., 2007), induction of ROS (Chandra, 2009), and synchronization of cells in the S-phase (Estrov et al., 2003). Activation of Sirt1 decreases HDACI-mediated p65 acetylation and NF-κB activation, which are events known to promote HDACI lethality (Dai et al., 2005). Resveratrol-mediated ROS generation, in cooperation with HDACIs (Inginga et al., 2005), triggers up-regulation of death receptors (e.g., DR5) (Hussain et al., 2011), which leads to activation of the extrinsic cascade, release of mitochondrial death proteins (e.g., cytochrome c), and full engagement of the apoptotic cascade. ROS also cause DNA damage, the lethal consequences of which may be exacerbated by HDACI-mediated interference with or down-regulation of DNA repair proteins (Subramanian et al., 2005; Lee et al., 2010). S-phase-synchronized leukemia cells may be particularly susceptible to the oxidative injury (Ge et al., 2006) and DNA damage triggered by the preceding events. The net effect of these cooperative actions is a pronounced induction of cell death in association with the inhibition of death receptors (e.g., DR5) and the resulting activation of the extrinsic apoptotic pathway.
death. Given the relative lack of toxicity of resveratrol concentrations considerably higher than those used in the present study (Howells et al., 2011), a strategy combining HDACs with resveratrol warrants further attention for AML and possibly other hematological malignancies.

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Authorship Contributions

Participated in research design: Yaseen, Chen, Rosato, Dent, Dai, and Grant.

Conducted experiments: Yaseen, Chen, Hock, and Dai.

Contributed new reagents or analytic tools: Chen, Dai, and Grant.

Performed data analysis: Yaseen, Chen, and Dai.

Wrote or contributed to the writing of the manuscript: Yaseen, Dai, and Grant.

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Resveratrol Increases HDAC Inhibitor Anti-AML Activity


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