Modeling the Effects of β₁-Adrenergic Receptor Blockers and Polymorphisms on Cardiac Myocyte Ca²⁺ Handling

Robert K. Amanfu and Jeffrey J. Saucerman

Department of Biomedical Engineering and the Robert M. Berne Cardiovascular Research Center, University of Virginia

Received November 23, 2013; accepted May 27, 2014

ABSTRACT

β-Adrenergic receptor blockers (β-blockers) are commonly used to treat heart failure, but the biologic mechanisms governing their efficacy are still poorly understood. The complexity of β-adrenergic signaling coupled with the influence of receptor polymorphisms makes it difficult to intuit the effect of β-blockers on cardiac physiology. While some studies indicate that β-blockers are efficacious by inhibiting β-adrenergic signaling, other studies suggest that they work by maintaining β-adrenergic responsiveness. Here, we use a systems pharmacology approach to test the hypothesis that in ventricular myocytes, these two apparently conflicting mechanisms for β-blocker efficacy can occur concurrently. We extended a computational model of the β₁-adrenergic pathway and excitation-contraction coupling to include detailed receptor interactions for 19 ligands. Model predictions, validated with Ca²⁺ and Förster resonance energy transfer imaging of adult rat ventricular myocytes, surprisingly suggest that β-blockers can both inhibit and maintain signaling depending on the magnitude of receptor stimulation. The balance of inhibition and maintenance of β₁-adrenergic signaling is predicted to depend on the specific β-blocker (with greater responsiveness for metoprolol than carvedilol) and β₁-adrenergic receptor Arg389Gly polymorphisms.

Introduction

β-Adrenergic receptor blockers (β-blockers) are front-line therapies for the treatment of heart failure, yet the biologic mechanism governing their success is still poorly understood (Krum, 2003; Tilley and Rockman, 2006; El-Armouche and Eschenhagen, 2009). The β₁-adrenergic receptor pathway has a dominant role in the regulation of heart contractility (Saucerman and McCulloch, 2006). One of the hallmarks of heart failure is elevated catecholamine release, which desensitizes the β-adrenergic pathway, causing an inability to increase contractility and cardiac output in response to acute stress (Ungere et al., 1994). Two apparently conflicting theories commonly postulated are that β-blockers are effective in heart failure by either inhibiting the harmful consequences of sustained adrenergic stimulation or maintaining the beneficial aspects of β₁-adrenergic receptor pathway activation (Lohe et al., 2003). The inhibition hypothesis is supported by clinical and experimental evidence that β-blockers help prevent or reverse the cardiac remodeling that leads to heart failure (Lowes et al., 1999). Conversely, the maintenance hypothesis is given credence by clinical evidence that β-blockers increase β₁-adrenergic receptor levels (Michel et al., 1988) and exercise tolerance (Engelmeier et al., 1985).

The ability of different β-blockers to either inhibit or maintain signaling is varied, causing controversy about which β-blocker is more effective in heart failure (Metra et al., 2006). Among the 17 US Food and Drug Administration-approved β-blockers, a variety of pharmacologic properties beyond receptor specificity alone may contribute to these differences (Mason et al., 2009). For example, some β-blockers are inverse agonists (Metra et al., 2006), reducing signaling below basal levels (Parra and Bond, 2007). Yet the importance of inverse agonism in determining clinical outcome during β-blocker treatment is unclear.

Genetic differences among patients also impact β-blocker efficacy (Krum, 2003). In vitro experiments in cell-expression systems show that the common β₁AR-Arg389Gly single-nucleotide polymorphism has a higher fold increase in adenyl cyclase activity after receptor stimulation but is more desensitized (Mason et al., 1999; Rathz et al., 2003). Carvedilol and metoprolol have similar affinities for both receptor variants in vitro (Joseph et al., 2004), but carvedilol has a larger effect on receptor conformation of the β₁-Arg389 variant (Rochais et al., 2007). Thus, there may be compound-specific phenotypes for β₁-adrenergic receptor polymorphisms (Dorn and Liggett, 2009).

The complexity of the β-adrenergic receptor pathway, coupled with the influence of receptor polymorphisms, makes it difficult to intuit the effect of β-blockers on observed cardiac physiology. Here we use a systems pharmacology approach (Sorger and Schoeberl, 2012), extending our previous computational models of β₁-adrenergic signaling and excitation-contraction coupling (Saucerman et al., 2003, 2004) to investigate the apparently conflicting mechanisms by which β-blockers may inhibit or maintain β-adrenergic signaling.

This work was supported by the American Heart Association [Grant 0830470N] and the National Institutes of Health National Heart, Lung, and Blood Institute [Grants HL094476 and HL05242 (to J.J.S)].

dx.doi.org/10.1124/mol.113.090951.

This article has supplemental material available at molpharm.aspetjournals.org.

Supplemental material to this article can be found at: http://molpharm.aspetjournals.org/content/suppl/2014/05/27/mol.113.090951.DC1

MOLECULAR PHARMACOLOGY Mol Pharmacol 86:222–230, August 2014

ABBREVIATIONS: β₁-AR, β₁-adrenergic receptor; β-blockers; β-adrenergic receptor blockers; ETCM, extended ternary complex model; FRET, Förster resonance energy transfer; IBMX, 3-isobutyl-1-methyloxanthine; MEM, minimum Eagle’s medium; PKA, protein kinase A.
We tested the hypothesis that in normal ventricular myocytes, both proposed mechanisms for β-blocker efficacy can occur concurrently. To do this, a previous computational model of the β₁-adrenergic receptor pathway was extended to include detailed receptor interactions for 19 ligands. Model predictions, validated with Ca^{2+} and Förster resonance energy transfer (FRET) imaging of isolated adult ventricular myocytes, surprisingly suggest that β-blockers can both inhibit and maintain signaling depending on the magnitude of receptor stimulation. In addition, the model predicted β-blocker-specific effects of receptor polymorphisms.

### Materials and Methods

**Computational Model of β-Blockers and β-Adrenergic Signaling.** A computational model was previously developed that integrates β₁-adrenergic receptor signaling with excitation-contraction coupling in rat cardiac myocytes and is based on mass action kinetics (Saucerman et al., 2003). The receptor module was previously described by a ternary complex model (De Lean et al., 1980). To better model the inverse agonism of some β-blockers seen in in vitro experiments (Varma et al., 1999), the receptor module of our original β₁-adrenergic receptor signaling model was replaced with the extended ternary complex model (ETCM) (Samama et al., 1993). The ETCM (Fig. 1) proposes two receptor states: active and inactive, and appropriately describes the constitutive activity of β-adrenergic receptors. The existence of these receptor states has been recently confirmed by determination of the crystal structure of the β₁-adrenergic receptor (Rosenbaum et al., 2011). Parameters for the ETCM and detailed calibration procedures are described in the Supplemental Methods and Supplemental Table 1. The expanded model has 49 algebraic and differential equations and is constrained by 102 parameters. Sensitivity analysis was used to determine ETCM parameters with distinct effects on model prediction before sequential parameter estimation (Supplemental Figs. 1 and 2). In descriptions comparing model predictions and experimental data, the terms calibration and fitting are used to describe instances where model parameters were used to better fit those data, while the term validation is used to describe instances where model parameters were not adjusted to fit those data.

**Isolation and Culture of Rat Cardiac Myocytes.** All procedures were performed in accordance with the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health and approved by the University of Virginia Institutional Animal Care and Use Committee. Adult rat ventricular myocytes were isolated similar to Bers et al. (Bers et al., 1990) from adult male Sprague-Dawley rats (200–250 g) Sprague-Dawley rats. Briefly, rats were anesthetized with ketamine/xylazine and hearts quickly excised before being Langendorff-perfused with collagenase (Cellutron Life Technologies, Baltimore, MD). Ventricular tissue was removed, mechanically dispersed, and filtered; and myocyte suspensions were rinsed and filtered (Förster resonance energy transfer (FRET) imaging of isolated adult ventricular myocytes, surprisingly suggest that β-blockers can both inhibit and maintain signaling depending on the magnitude of receptor stimulation. In addition, the model predicted β-blocker-specific effects of receptor polymorphisms.

The extended ternary complex model of the β₁-adrenergic receptor, coupled with the β₁-adrenergic pathway and ventricular myocyte excitation-contraction coupling, was developed with the Myopacer (Ionoptix, Milton, MA) at a frequency of 1 Hz with bipolar pulse duration of 4 milliseconds at a voltage of 10 V. All measurements were performed at room temperature.

**Camera-Based Ca^{2+} Imaging of Myocytes.** Ca^{2+} was measured using fluo-4 as described previously (Amanfu et al., 2011). Myocytes were imaged on an Olympus IX-81 inverted microscope (Olympus, Center Valley, PA) with a Hamamatsu C9300 charge-coupled device camera (Bridgewater, NJ) and automated stage (Prior Scientific, Rockland, MA) at a sampling frequency of 67 Hz using Metamorph (Molecular Devices, Sunnyvale, CA). To minimize photobleaching and phototoxicity, cells were imaged intermittently for 10 seconds after every minute. Automated cell segmentation using Otsu’s method identified regions of interest from which Ca^{2+} transients for each cell were extracted at each time point. Raw fluorescence values were background-subtracted and normalized to yield fold change in fluo-4 intensity:

\[
\text{fluor} = \frac{\text{fluor}_{t} - \text{fluor}_{t=0}}{\text{fluor}_{t=0}}
\]

Average fluo-4-fold change was calculated by averaging five to seven consecutive transients at specific time points. All segmentation and feature extraction was implemented in MATLAB. Code for these
analyses and example movies are freely available at http://bme.virginia.edu/saucerman.

**FRET Imaging of Cardiac Myocytes.** Adenovirus was constructed from plasmid DNA of AKAR3 protein kinase–A reporter (Allen and Zhang, 2006). Myocytes were infected with adenovirus immediately after isolation in serum MEM media for 1 hour. Cells were then cultured for 24 hours in serum-free MEM media. Myocytes were preincubated in solutions of 0.1 μM isoproterenol with and without 0.1 μM propranolol. Cells were placed in a slotted bath with Tyrodes perfusate and paced at 10 Hz. Expressing myocytes were imaged on an Olympus IX-81 inverted microscope with a Hamamatsu C9300 charge-coupled device camera. A cocktail of 10 μM forskolin (Tocris) and 100 μM 3-isobutyl-1-methylxanthine (Sigma-Aldrich, St. Louis, MO) was used as positive control at the end of each experiment. Automated cell segmentation and FRET computation (using the precision FRET [PFRET] algorithm) (Chen and Periasamy, 2006) were performed in MATLAB. FRET response was normalized to positive control.

**Results**

**Calibration and Validation of the β1-Adrenergic Model with the Extended Ternary Complex Receptor Model.** To quantitatively investigate how β-blockers modulate β-adrenergic signaling in cardiac myocytes, a computational model of the β1-adrenergic receptor pathway was developed that includes detailed interactions between ligand, receptor, and G-protein in the form of the extended ternary complex model (Fig. 1). The integrated model describes stimulation of the β1-adrenergic receptor, activation of receptor intermediates, production of cAMP, activation of protein kinase A (PKA), phosphorylation of downstream PKA targets, and the effect on Ca²⁺ transients. Receptor desensitization by both the β-adrenergic receptor kinase and PKA is also included.

Model predictions were compared with a range of experimental data from the literature and the current study (Fig. 2). The shift in agonist binding to the receptor in the presence of guanosine 5′-[β,γ-imido] triphosphate (which displaces the G protein) was validated (Fig. 2A). The model validates reasonably well against measured kinetics of cAMP (Fig. 2B) and PKA activity (Fig. 2D) in response to isoproterenol. The model is calibrated to have appropriate basal and maximally stimulated cAMP levels in cardiac myocytes, with validation of the sensitivity to isoproterenol (Fig. 2C). The EC₅₀ of isoproterenol for phosphorylation of phospholamban by PKA is also accurately validated (Fig. 2E). The model was calibrated to have an appropriate EC₅₀ of isoproterenol for Ca²⁺ transients to isoproterenol, as measured with fluo-4 by our group and others (Fig. 2F). In addition, we validated model predictions of Ca²⁺ transient responses to increasing propranolol concentration in the presence of 0.1 μM isoproterenol (Supplemental Fig. 4). A summary of all calibrations and validations is provided in Supplemental Table 2. These results indicate that the updated model is consistent with experimental data at multiple levels of the β₁-adrenergic receptor pathway, providing confidence in the utility of the model for testing hypotheses regarding β-blocker efficacy.

**Fig. 2.** Experimental validation of coupled β₁-adrenergic signaling and excitation-contraction coupling model. (A) Model reproduces shift in agonist binding affinity in the presence of guanosine 5′-[β,γ-imido] triphosphate (GPP), which displaces G from the receptor. (B) Kinetics of cAMP stimulation by 10 nM isoproterenol (ISO). (C) Model dose response to ISO. (D) PKA activity measured by FRET reporter AKAR3. (E) Phospholamban phosphorylation in response to ISO. (F) Ca²⁺ dose response to ISO. Results in (A), (C), (E), and (F) show direct comparison with published experimental data (Mason et al., 1999), (Vila Petroff et al., 2001), (De Arcangelis et al., 2010), (Vittone et al., 1998), and (Collins and Rodrigo, 2010), whereas data in (D) and (F) were acquired in the current study.
Propranolol Inhibits and Maintains the \(\beta\)-Adrenergic Response Depending on the Magnitude of Receptor Stimulation. While inhibition and maintenance of \(\beta\)-adrenergic responsiveness are typically thought to be incompatible explanations of \(\beta\)-blocker efficacy, we hypothesized that both could occur depending on the magnitude of receptor stimulation. To test this hypothesis in silico, we simulated low (0.1 \(\mu\)M) and then high (10 \(\mu\)M) levels of isoproterenol in the absence and presence of the first-generation \(\beta\)-blocker propranolol. We used 0.1 \(\mu\)M propranolol because this was the lowest dose that suppressed Ca\(^{2+}\) transients at 0.1 \(\mu\)M isoproterenol (Supplemental Fig. 3). Low and high doses of isoproterenol are analogous to chronically elevated levels of catecholamines in heart failure and acutely elevated levels in exercise, respectively. In the absence of propranolol, the model predicts that low receptor stimulation increases Ca\(^{2+}\) amplitude (Fig. 3A), with no further sensitivity to subsequent high levels of isoproterenol (Fig. 3B). In the presence of propranolol, responsiveness to low receptor stimulation is suppressed, but the pathway maintains sensitivity of Ca\(^{2+}\) transients to high isoproterenol (Fig. 3C). Independent experiments imaging Ca\(^{2+}\) dynamics in isolated adult rat ventricular myocytes qualitatively validated these model predictions (Fig. 3, D–F). These simulations and experiments indicate that the apparently conflicting roles of the \(\beta\)-blocker propranolol to inhibit signaling and maintain responsiveness are in fact compatible.

To experimentally investigate whether these effects persist with chronic receptor stimulation, cells were pretreated with a low dose of isoproterenol for 24 hours before subsequent stimulation with high isoproterenol (Fig. 4). In the absence of propranolol, Ca\(^{2+}\) transient amplitude in pretreated cells was not further sensitive to high isoproterenol (Fig. 4C), as Ca\(^{2+}\) transients were already elevated. In contrast, cells pretreated with propranolol maintained sensitivity to high-dose isoproterenol in the presence of propranolol, similar to model predictions and the acute experiments (Fig. 3). Using a FRET reporter for PKA activity, we found that PKA (upstream of Ca\(^{2+}\) in the \(\beta\)-adrenergic pathway) also maintains sensitivity to high isoproterenol in the presence of propranolol (Fig. 4B), again validating model predictions (Supplemental Fig. 8).

\(\beta\)-Blockers Differ in Their Ability to Inhibit and Maintain \(\beta\)-Adrenergic Responsiveness. To examine whether the dual role of propranolol in inhibiting and maintaining \(\beta\)-adrenergic responsiveness may be generalized to other \(\beta\)-blockers, we extended the model to 17 additional

---

**Fig. 3.** Propranolol both inhibits and maintains \(\beta\)-adrenergic-mediated regulation of Ca\(^{2+}\) transients. (A) Model-predicted individual Ca\(^{2+}\) transients in response to increasing concentration of isoproterenol (ISO). (B) Ca\(^{2+}\) concentration increased in response to 0.1 \(\mu\)M ISO, with no further response to subsequent stimulation with 10 \(\mu\)M ISO. (C) The model predicted that propranolol (PRO) inhibits response to 0.1 \(\mu\)M ISO, but the responsiveness to 10 \(\mu\)M ISO is maintained (large sensitivity). (D) Individual Ca\(^{2+}\) transients as measured by fluo-4 from rat ventricular myocytes exposed to increasing ISO; scale bar 20 \(\mu\)m. (E) Similar to model predictions, myocytes were not responsive to further stimulation with 10 \(\mu\)M ISO. (F) PRO inhibited response to 0.1 \(\mu\)M ISO, but myocytes were responsive to further stimulation with 10 \(\mu\)M ISO. Sensitivity was quantified as the increase in Ca\(^{2+}\) transient magnitude when increasing from 0.1 \(\mu\)M ISO (analogous to chronically elevated catecholamines in heart failure) to 10 \(\mu\)M ISO (analogous to exercise).
Metoprolol and Carvedilol Differ in Their Ability to Maintain β-Adrenergic Responsiveness. An interesting prediction of the in silico screen is that carvedilol and metoprolol (two clinically prescribed β-blockers) differ significantly in their ability to enhance cAMP sensitivity. This difference indicates that the two drugs may have distinct effects (inhibition or maintenance) on the β1-adrenergic response. To experimentally validate model predictions for carvedilol and metoprolol, Ca2+ imaging experiments in adult rat ventricular myocytes were performed. As with propranolol, we empirically selected doses for carvedilol and metoprolol that were just sufficient to suppress responses to 0.1 μM isoproterenol (Supplemental Fig. 4). Sensitivity to high-dose isoproterenol was maintained in cardiac myocytes treated with 1 μM metoprolol (Fig. 6A) but suppressed with 1 μM carvedilol (Fig. 6B), qualitatively validating our model predictions (summarized in Fig. 6, C and D).

To test the robustness of this result to β-blocker concentration, we further simulated how cAMP sensitivity is affected by propranolol, metoprolol, and carvedilol doses. The model predicted that propranolol and metoprolol robustly maintained β-adrenergic responsiveness, but that at doses lower than we had previously examined, carvedilol may also maintain β-adrenergic responsiveness (Supplemental Fig. 5). To test this prediction, we performed subsequent experiments with 0.3 μM carvedilol (Supplemental Fig. 6), which showed that lower carvedilol still suppressed the responsiveness to high-dose isoproterenol. The robust suppression seen in the carvedilol experiments can be explained by an alternative receptor model accounting for binding of carvedilol to an allosteric site on the β1-adrenergic receptor (Supplemental Fig. 7), as supported by previous experiments by Kindermann et al. (Kindermann et al., 2004).

Receptor Polymorphisms Are Differentially Modulated by Diverse β-Blockers. Genetic differences among patients also impact β-blocker efficacy (Shin and Johnson, 2010). Patients with the β1-Arg389 variant have a better prognosis after β-blocker administration compared with patients with the β1-Gly389 polymorphism. Increased G-protein binding is observed experimentally for the β1-Arg389 variant, causing higher constitutive activity. This behavior
was modeled by altering $K_G$, the ETCM model parameter that affects binding of the active receptor to G-protein. $K_G$ in the $\beta_1$-Arg389 model was manually calibrated to 0.7 $\mu$M to replicate the shift in agonist binding in the presence of guanosine 5'-$\beta,\gamma$-imido triphosphate (Fig. 7A) and the higher constitutive activity of the Arg389 variant (Fig. 7B).

$\beta_1$-Arg389 and $\beta_1$-Gly389 polymorphisms were predicted to have varying responses to propranolol. Propranolol had more of an effect inhibiting the low-dose isoproterenol cAMP and Ca$^{2+}$ responses in the Arg389 variant (Fig. 7C), but it also enhanced sensitivity to high-dose isoproterenol. The 19 ligands were predicted to have varying effects on cAMP sensitivity (Fig. 7D), similar to the $\beta_1$-Gly389 variant (Fig. 5A). However, there are some significant differences in the responses to particular ligands between the receptor polymorphisms (Fig. 7E). For example, atenolol was predicted to be less effective at maintaining $\beta_1$-adrenergic responsiveness for the Arg389 variant compared with the Gly389 variant. This diversity of responses indicates that computational models may be useful for predicting pharmacogenetic interactions.

**Discussion**

**Mechanisms of $\beta$-Blocker Efficacy in Heart Failure.**

A key feature of heart failure is the modest chronic elevation of circulating catecholamines (e.g., epinephrine) which desensitizes the $\beta_1$-adrenergic receptor signaling pathway, rendering patients incapable of increasing cardiac output in response to intense acute stress (e.g., exercise). Crucial alterations to the signaling pathway in this chronically activated state include reduced $\beta_1$-adrenergic receptor density (Bristow et al., 1982) and Ca$^{2+}$ (Harding et al., 1992) in response to adrenergic stimulation. Sustained stimulation has detrimental long-term consequences, including apoptosis and hypertrophy (Communal et al., 1998; Taimor et al., 2001). Maintenance of signaling in cardiomyopathy by adenylyl cyclase overexpression (Roth et al., 1999) or G-protein receptor kinase 2 inhibition (Reinkober et al., 2012) has improved cardiac function in in vitro murine models. Previous studies of mechanisms governing $\beta$-blocker efficacy have focused exclusively on one of two mechanisms: i.e., the inhibition (Lowes et al., 1999) or maintenance of $\beta_1$-adrenergic receptor signaling (Engelmeier et al., 1985; Michel et al., 1988). With evidence supporting both theories, it is unclear how these two contradictory mechanisms can explain the same biologic phenomena or the appropriate context where one mechanism dominates. This study provides evidence that at least in normal isolated adult ventricular myocytes, both mechanisms can occur concurrently dependent on the magnitude of receptor stimulation.

Complexities at the receptor level and the influence of receptor polymorphisms complicate attempts to infer these mechanisms. Computational modeling is highly suited for this task by allowing the unbiased comparison of clinically available $\beta$-blockers. Previous computational models of the $\beta_1$-adrenergic receptor pathway have used simplified receptor kinetic models (Saucerman et al., 2003, 2004). Although sufficient to describe the activation of the signaling pathway by agonists, these pathway models do not have the mechanistic detail of receptor kinetics needed to adequately model
the inverse agonism of β-blockers. Detailed receptor models have been developed, but these models have been evaluated in isolation from downstream signaling pathways (Samama et al., 1993). To model β-blockers, detailed models of receptor kinetics were linked to the cardiac β₁-adrenergic receptor pathway and excitation contraction coupling. Computational model simulations indicate that both inhibition and maintenance of signaling are compatible, dependent on the magnitude of receptor stimulation. Propranolol inhibited low-dose isoproterenol (analogous to chronic levels of catecholamine seen in heart failure) but enabled sensitivity to high-dose isoproterenol (analogous to acute catecholamine levels during exercise). Fluo-4 and FRET imaging of isolated cardiac myocytes confirmed this prediction.

Metoprolol and Carvedilol Have Distinct Mechanisms of Action in Isolated Ventricular Myocytes. Separate clinical trials of the two β-blockers commonly used to treat heart failure show reduction in mortality. Results of the COMET trial, which aimed to compare both treatments, concluded that carvedilol had a larger effect on mortality (Poole-Wilson et al., 2003). Significant controversy surrounds this result with questions raised on the appropriate dose of each compound that merits fair comparison (Kveiborg et al., 2007). Another important clinical measure of heart failure treatment effectiveness is exercise tolerance. Studies have shown that metoprolol has a larger effect on exercise tolerance than carvedilol (Metra et al., 2000). Our computer simulations and Ca²⁺ imaging experiments confirm that metoprolol maintains β-adrenergic signaling in isolated ventricular myocytes due to its moderate binding affinity and high inverse agonism. Carvedilol, although also an inverse agonist, did not maintain isoproterenol sensitivity, due to its tight binding to the β₁-adrenergic receptor and the potential contribution from allosteric binding (Kindermann et al., 2004).

Pharmacogenomic Targeted Treatment with β-Blockers. Another factor complicating treatment of heart failure patients is the presence of β-adrenergic receptor polymorphisms. β₁-Gly389 has been shown to couple less effectively to G-protein in expression cell systems, but the β₁-Arg389 variant provides higher risk for heart failure and differential responses to β-blockers. A recent study has shown that carvedilol exhibits enhanced inverse agonism with the β₁-Arg389 variant (Rochais et al., 2007), an example of the potential for personalized medicine. Understanding how genotype affects therapeutic response is expected to open a new era of pharmacogenomics and personalized medicine. One obstacle is that existing knowledge of β₁-adrenergic
receptor polymorphisms comes from cell lines that might function differently in healthy versus failing myocytes. We modeled β1-adrenergic receptor polymorphisms in the background of a ventricular myocyte. The model identified differences between the receptor polymorphisms’ cAMP sensitivity to high-dose isoproterenol in the presence of particular ligands. For example, atenolol was predicted to be less effective at maintaining β-adrenergic responsiveness in isolated ventricular myocytes expressing β1-Arg389 compared with the β1-Gly389 variant.

Limitations and Considerations

A critical decision in developing computational models is specifying the models’ scope. Uncertainty in parameters, and henceforth the ensuing predictions, becomes overwhelming as model scope increases. We have restricted our model to the β1-adrenergic receptor pathway and its effects on Ca2+ transients in isolated rat ventricular myocytes, because this pathway plays a central role in enhancing contractility after β-adrenergic stimulation. However, an alternative hypothesis is that other properties of β-blockers (i.e., binding to other adrenergic receptors and pharmacokinetic properties including half-life, lipid solubility, and nonspecific binding) may play a larger role than blockade of the β1-adrenergic receptors. Indeed our simulations suggest that binding of carvedilol to an allosteric site on the β1-adrenergic receptor influences its effect on β-adrenergic responsiveness. Our current computational model is not yet able to fully explore the consequence of this mechanism in vivo. Future work could couple the β1-adrenergic signaling model to whole-body pharmacokinetics or simulate crosstalk with other adrenergic receptors, including the β2-adrenergic receptor (Zamah et al., 2002).

Conclusions

Previous studies have suggested two seemingly conflicting mechanisms (inhibition or maintenance of the β-adrenergic receptor signaling pathway) to explain β-blocker efficacy. Here we show, both in pathway models and adult ventricular myocytes, that the β-blockers propranolol and metoprolol (but not carvedilol) not only block response to low isoproterenol (analogous to chronic stimulation in heart failure) but also maintain the β-adrenergic receptor response to subsequent high-dose isoproterenol (analogous to acute stimulation in exercise). Thus, both inhibition and maintenance of signaling can occur concurrently, dependent on the magnitude of receptor stimulation. Computational simulations indicate that these responses are modulated by particular receptor polymorphisms. Evaluating the mechanisms for these differences, with the help of computational models, is an important step toward designing personalized β-blocker therapies.

Acknowledgments

The authors thank Renata Polanowska-Grabowska for technical assistance.

Authorship Contributions

Participated in research design: Amanfu, Saucerman.
Conducted experiments: Amanfu.
Contributed new reagents or analytic tools: Amanfu.
Performed data analysis: Amanfu.
Wrote or contributed to the writing of the manuscript: Amanfu, Saucerman.

Fig. 7. β1AR-Arg389 polymorphism responds differently to β-blockers. (A) Model reproduces the shift in agonist binding affinity in the presence of guanosine 5’-[β,γ-imido] triphosphate for Arg389. (B) Concentration dependence of adenylyl cyclase (AC) activity to isoproterenol for Gly389 and Arg389. (C) Arg389 is predicted to have higher cAMP sensitivity and Ca2+ response versus Gly 389 in cardiac myocytes. (D) In silico screen of 19 β1-adrenergic ligands against Arg389. (E) Differential cAMP sensitivity between Arg389 and Gly389 for different β1-adrenergic ligands predicted for cardiac myocytes. Experimental data in panels (A) and (B) are from (Mason et al., 1999; Mialet Perez et al., 2003).
References


Faaza S and Bond RA (2007) Isoverse agonism from curiosity to accepted dogma, but is it clinically relevant? Curr Opin Pharmacol 7:143–150.


Address correspondence to: Dr. Jeffrey J. Saucerman, Department of Biomedical Engineering, PO Box 800759, Charlottesville, VA 22908. E-mail: jsaucerman@virginia.edu