Norepinephrine Transporter Inhibition with Desipramine Exacerbates L-DOPA–Induced Dyskinesia: Role for Synaptic Dopamine Regulation in Denervated Nigrostriatal Terminals

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ABSTRACT

Pharmacological dopamine (DA) replacement with Levodopa [L-dihydroxyphenylalanine (L-DOPA)] is the gold standard treatment of Parkinson’s disease (PD). However, long-term L-DOPA treatment is complicated by eventual debilitating abnormal involuntary movements termed L-DOPA-induced dyskinesia (LID), a clinically significant obstacle for the majority of patients who rely on L-DOPA to alleviate PD-related motor symptoms. The manifestation of LID may in part be driven by excessive extracellular DA derived from L-DOPA, but potential involvement of DA reuptake in LID severity or expression is unknown. We recently reported that in 6-hydroxydopamine (6-OHDA)–lesioned striatum, norepinephrine transporter (NET) expression increases and may play a significant role in DA transport. Furthermore, L-DOPA preferentially inhibits DA uptake in lesioned striatum. Therefore, we hypothesized that desipramine (DMI), a NET antagonist, could affect the severity of LID in an established LID model. Whereas DMI alone elicited no dyskinetic effects in lesioned rats, DMI + L-DOPA–treated rats gradually expressed more severe dyskinesia compared with L-DOPA alone over time. At the conclusion of the study, we observed reduced NET expression and norepinephrine-mediated inhibition of DA uptake in the DMI + L-DOPA group compared with the L-DOPA–alone group in lesioned striatum. LID severity positively correlated with striatal extracellular signal-regulated protein kinase phosphorylation among the three treatment groups, with increased ppERK1/2 in DMI + L-DOPA group compared with the L-DOPA– and DMI-alone groups. Taken together, these results indicate that the combination of chronic L-DOPA and NET-mediated DA reuptake in lesioned nigrostriatal terminals may have a role in LID severity in experimental Parkinsonism.

Introduction

Parkinson’s disease (PD) is the most common neurodegenerative movement disorder, and its incidence is likely to only increase with the impending growth of the aging population (Siderowf and Stern, 2003). PD is primarily characterized by the loss of dopamine (DA) neurons in the substantia nigra pars compacta. Therapeutically, the primary loss in nigral DA necessitates its replacement through the exogenous administration of L-dihydroxyphenylalanine (L-DOPA), a DA precursor, or with DA agonists (Hornykiewicz and Kish, 1987; Steiger and Quinn, 1995). L-DOPA in particular, has remained the drug of choice for treating PD for nearly a half century (Calne and Marsden, 1994; Olanow and Koller, 1998; Ahlskog and Mueenter, 2001). Not only is the onset of dyskinesia a significant setback for the patient, its presence is often permanent, occurring with every subsequent exposure to L-DOPA, which limits the clinical efficacy of what has long been considered our gold standard in PD treatment.

Evidence suggests chronic L-DOPA leads to major adaptive molecular changes occurring within the basal ganglia that may underlie LID pathophysiology (for review, see Cenci and Konradi, 2010). Loss of nigrostriatal dopaminergic neurons not only impairs presynaptic control of DA regulation, but also leads to large variances in extracellular levels of DA that parallel L-DOPA dosing regimens (Cenci and Lundblad, 2006). It is these supraphysiologic fluctuations in extracellular DA content that are thought to underlie the induction of L-DOPA–induced dyskinesia (Chase, 1998). LID has been associated with plastic changes in postsynaptic neuronal targets within the striatum, including abnormal trafficking of the DA D1 receptor (Guigoni et al., 2007; Berthet et al., 2009). This has

ABBREVIATIONS: AIM, abnormal involuntary movement; ANOVA, analysis of variance; CNS, central nervous system; DA, dopamine; DAT, DA transporter; DMI, desipramine; ERK, extracellular signal-regulated protein kinase; L-DOPA, L-dihydroxyphenylalanine; LID, L-DOPA–induced dyskinesia; NE, norepinephrine; NET, NE transporter; PD, Parkinson’s disease; ppERK, extracellular signal-regulated protein kinase phosphorylation; SERT, serotonin terminals or transporter; TH, tyrosine hydroxylase.
established 6-OHDA LID rodent model. In an effort to further explore this possibility, we examined the role in the onset of dyskinesia, the exact mechanism of how this occurs remains conflicting and inconclusive (Cenci and Konradi, 2010).

In PD, the loss of the DA transporter (DAT) most likely impairs its ability to maintain DA bioavailability, yet most studies indicate motor symptoms are seen only when ~70% of striatal DAT is lost (Bernheimer et al., 1973; Bezard et al., 2001), indicating an alternate mechanism through which DA may still be effective to produce normal locomotion. Indeed, many studies have concluded that serotonergic terminals may transport L-DOPA or DA, and subsequently release DA to maintain DA signaling, albeit in a dysregulated fashion (Arai et al., 1995; Miller and Abercrombie, 1999; Tanaka et al., 1999; Kannari et al., 2001; Carta et al., 2007). However, it is a comparatively neglected observation that in sparsely dopaminergic innervated regions, such as the frontal cortex, the norepinephrine (NE) transporter (NET) can also transport DA (Moron et al., 2002) and NE uptake inhibitors, which can result in increased extracellular DA levels within the prefrontal cortex (Carboni et al., 1990; Chiara et al., 1992; Gresch et al., 1995; Tanda et al., 1997; Yamamoto and Novotney, 1998; Wayment et al., 2001; Masana et al., 2012). In line with these observations, we have recently reported that DA transport still occurs in the 6-OHDA-lesioned striatum, despite major loss of DAT, and that a potential role for NET in DA uptake is evident (Chotibut et al., 2012). In these experiments, L-DOPA preferentially inhibits DA reuptake when striatal DAT loss exceeds the level necessary for PD symptom appearance, and preferentially inhibits NE over DA uptake in striatal tissue. Therefore, based upon these observations, chronic L-DOPA could exacerbate the L-DOPA–induced dysregulation of synaptic DA by its influence on a NET-dominated regulation of DA uptake that evolves with the gradual loss of DA terminals.

In an effort to further explore this possibility, we examined how the blockade of NET in a chronic desipramine (DMI) paradigm would affect dyskinesia expression, NET expression, DA uptake, and an established post-translational event in striatum associated with dyskinesia, extracellular signal-regulated protein kinase phosphorylation (ppERK), in an established 6-OHDA LID rodent model.

Materials and Methods

Animals. Male Sprague Dawley rats purchased from Charles River (Wilmington, MA) were used in all experiments. A total of 21 test subjects was used at the start of the study, was 4–8 months of age, and was housed 1 per cage and under controlled lighting conditions (12:12 light:dark cycle) with food and water available ad libitum. All animals were used in compliance with federal and institutional Animal Care and Use Committee guidelines at Louisiana State University Health Sciences Center (Shreveport, LA). All behavioral testing was performed between 10:00 and 16:00 hours. Behavioral data obtained from test subjects that did not have >70% tyrosine hydroxylase (TH) loss, as discovered after completion of LID assessments and tissue analysis, were excluded from the results. We note that protein recovery was limited in some rats due to the fact that we analyzed DA uptake, TH, NET, total ERK, and ppERK all from the same striatal tissues and, as such, the treatment group numbers are not of equal size.

In all experiments, rats were rendered briefly unconscious with isoflurane and immediately decapitated on day 40 of the experiment (1 day post-last behavioral test and treatment (L-DOPA + DMI, L-DOPA, or vehicle)) for dissection of striatal tissues. An overview of the experimental design to be described below is depicted in Figure 1.

6-OHDA Lesions. Rats were anesthetized with 40 mg/kg Nembutal i.p. (pentobarbital; Lundbeck, Deerfield, IL) with supplement of 9.0, 0.6, and 0.3 mg/kg ketamine, xylazine, and acepromazine, respectively. Each animal then underwent survival surgery to deliver the neurotoxin 6-OHDA unilaterally to the medial forebrain bundle while immobilized in a stereotaxic frame (coordinates ML +1.5, AP –3.8, DV –8.0 relative to Bregma). All stereotaxic coordinates are cited according to the stereotaxic atlas of Paxinos and Watson rat brain atlas (Paxinos and Watson, 1998). A total of 16 μg 6-OHDA in a total of 4 μl in 0.02% ascorbic acid (concentration of 4 mg/ml) was infused unilaterally at a rate of 1 μl/min. The contralateral medial forebrain bundle was infused with vehicle (0.02% ascorbic acid) and infused at the same rate and coordinates. The syringe was left in place for 10 minutes before removal to allow for maximal diffusion of drug and to avoid further mechanical damage to the tissue. Body temperature was maintained at 37°C during surgery using a temperature monitor with probe and heating pad (FHC, Bowdoinham, ME). Animals were kept warm after surgery and monitored closely after anesthesia. In our hands (Chotibut et al., 2012), we have previously shown that, after 6-OHDA infusion, there was no significant difference in lesioned striatal tissue NE content compared with contralateral striatum and, as such, have based these experiments from these observations. As such, we did not pretreat these rats with DMI.

Amphetamine Testing. The extent of the lesion was evaluated 7 days postsurgery based on the net ipsilateral rotations measured over a 60-minute period following an injection of 2.5 mg/kg D-amphetamine i.p. (in 0.9% saline) (Ungerstedt and Arbuthnott, 1970). One hundred net full turns on the ipsilateral side to the lesion were necessary to be included in the study. In our hands, we have determined this time frame as enough to consistently catch >60% of TH loss (Chotibut et al., 2012, 2014; Salvatore, 2014). Ultimately, however, postmortem verification of the lesion severity was determined by assessment of TH protein, as previously reported (Chotibut et al., 2012, 2014). Thus, any further exclusions were due to rats that did not also have >70% TH loss after Western blot analysis at the end of the study. The behavioral AIMS data reflect test subjects with >70% TH protein loss.

DMI and L-DOPA Administration. Nine days postsurgery, rats were randomly divided into three groups. Two groups received a treatment of DMI (Tocris, Bristol, UK; catalogue 3067) (12 mg/kg), and one group received vehicle (0.9% saline 12 mg/kg) i.p. for 30 consecutive days. Then, at 19 days postsurgery, an additional treatment of either L-DOPA (12 mg/kg and benserazide-hydrochloride (15 mg/kg) or vehicle (0.9% saline) was given once daily for 20 consecutive days (Fig. 2). In summary, three treatment groups were created, as follows: 1) DMI pretreatment + L-DOPA/benserazide; 2) vehicle pretreatment + L-DOPA/benserazide; and 3) DMI pretreatment + vehicle (0.9% NaCl, saline; Hospira, Lake Forest, IL). With 6-OHDA lesion present in each group, these three groups were used to evaluate the impact of DMI on LID behavior caused by chronic L-DOPA, with the DMI-alone group controlling for whether NET blockade alone could produce LID.

Behavioral AIMS Ratings. L-DOPA–induced abnormal involuntary movements (AIMs) were then rated at six discrete time points...
Fig. 1. Timeline of treatment paradigm. Male Sprague Dawley rats were lesioned unilaterally with 6-OHDA and randomly assigned a treatment group. Seven days postlesion, rats were tested with amphetamine (2.5 mg/kg i.p.), and a number of turns ipsilateral to lesion turns were assessed to confirm for a successful lesion. Treatment began on day 9 with 3 treatments: 1) desipramine + L-DOPA; 2) desipramine + vehicle; and 3) vehicle + L-DOPA. Beginning on day 19, behavioral AIMs were assessed at six discrete time points for the remainder of the study (days 19, 23, 27, 31, 35, 39). On day 39, rats were sacrificed and dopamine uptake and biochemical markers were assessed.

During L-DOPA administration, rats were placed in separate cages and individual dystonic behaviors were quantified based on severity and frequency during a 1-minute observation period performed every 20 minutes over a period of 160 minutes. AIMs were classified into three subtypes, as follows: axial AIMs (dystonic posturing of the upper trunk toward the side contralateral to the lesion), limb AIMs (hypermotoric or jerk movements of the forelimb contralateral to the lesion), and orolingual AIMs (abnormal jaw movements, facial twitching, and tongue protrusion). Each subtype was scored based on severity and frequency from 0 to 4, with 4 being the most severe and/or occurring continuously during the entire 1-minute observation period (for review, see Cenci et al., 1998 and Lundblad et al., 2004). AIMs scores were calculated by multiplying the severity score and/or occurring continuously during the entire 1-minute observation period. Theoretically, the highest AIMs score achievable in 1 behavioral test day (with 8 observation periods) would be 384.

Preparation of Synaptosomes. To ascertain uptake properties in conjunction with protein and phosphorylation expression levels present in the rats immediately following the last LID assessment, synaptosomes were prepared according to the protocol previously described (Salvatore et al., 2003) with the following modifications: tissue dissected from dorsal striatum was homogenized in 5 mL 0.32 M sucrose solution using a Teflon/glass homogenizing wand (Glas-Col, Terre Haute, IN). All tissue was kept on ice or at 4°C from the moment of brain excision until the uptake assay took place. Background determinations were subtracted in the same manner as in the DA uptake determinations of NET protein from the 6-OHDA–lesioned and contralateral (control) striatum. The supernatant was aspirated and resuspended in 1 mL Kreb’s buffer (118 mM NaCl, 4.7 mM KCl, 1.2 mM KH2PO4, 25 mM NaHCO3, 1.0 mM Na2 EDTA, 1.7 mM CaCl2, 10 mM glucose, 100 μM parglyline, and 100 μM ascorbic acid). Protein concentration was determined using a bicinchoninic acid colormetric assay (Thermo Scientific, Rockford, IL). All tissue was kept on ice or at 4°C from the moment of brain excision until the uptake assay took place.

[^1H]DA Uptake into Synaptosomes. Synaptosomes were distributed in ice-cold test tubes to prepare for DA uptake. The determination of[^1H]DA uptake in the crude synaptosomes from dorsal striatum harvested from the contralateral and 6-OHDA–infused hemispheres was conducted simultaneously and included assessments of DA uptake capacity in the presence of unlabeled 1 μM NE and DA. We previously determined that this concentration of NE or DA had differential impact on DA uptake (Chotibut et al., 2012) and consistent with previous observations. Each determination was done in triplicate for each assay condition, and uptake was determined comparing the lesioned striatum with the contralateral control striatum. Nonspecific uptake was determined by counts obtained in synaptosomes incubated with 500 nM DA (all as labeled DA) on ice during the time period of uptake. Background was determined and subtracted in the same manner as in the DA uptake studies.

Synaptosomes (30 μg protein per replicate) were added to 4°C oxygenated Kreb’s buffer and test ligand (if indicated) to reach a total volume of 100 μL. The synaptosomes were then warmed to 35°C for 5 minutes, and then 100 μL prewarmed 1μM[^1H]DA, prepared from ViTrax, [7-, 8-[^1H]DA], specific activity of 25 Ci/mmol, was added to the synaptosome preparations (giving a 500 nM final[^1H]DA cure of TH protein standard (Salvatore et al., 2009). We have determined in previous experiments that this fraction is sufficient for precise assessment of cytosolic proteins (Chotibut et al., 2012, 2014), reflecting the relative quantities recovered in fresh-frozen preparations (Salvatore et al., 2009). An aliquot of P2 fraction was saved for determination of TH protein, ppERK, and total ex-
concentration), allowed to incubate for uptake, and terminated after 120 seconds with an excess volume of ice-cold Kreb’s buffer and reimmersing the tubes in the ice bath. The uptake time for DA was chosen to be as close as technically and practically possible to the approximately 2-minute uptake time of striatal DA observed in vivo, as seen by Sabeti et al. (2002), and where differences in uptake capacity between lesioned and intact striatum have been previously reported (Chotibut et al., 2012). Synaptosomes were washed extensively to remove excess labeled DA with equal osmolarity phosphate-buffered saline buffer through a Brandel M24-TI (Gaithersburg, MD) cell harvester using Brandel GF/C filter paper pretreated with a 2% polyethylenimine solution to reduce nonspecific binding of label. The filter paper containing the rinsed synaptosomes was transferred into scintillation vials containing 5 mL biodegradable scintillation cocktail (Research Products International, Mount Prospect, IL) and counted with a Beckman Coulter LS6500 scintillation counter (Brea, CA).

Calculating DA Uptake. To determine the quantity of DA uptake, the percentage of [3H]DA recovered in the synaptosomes against the total amount of [3H]DA added during the uptake experiment was first determined. The total pmol of recovered [3H]DA was then determined based upon the percentage of [3H]DA recovery in the synaptosomes after subtracting the nonspecific binding value, and

Fig. 2. Abnormal movements induced by the chronic treatment with L-DOPA in animals lesioned with 6-OHDA. The time course of changes in dyskinesia evaluated from the product of the frequency and amplitude behavior (orolingual, axial, forelimb) induced by a 20-day treatment with L-DOPA (6 mg/kg plus benserazide 12 mg/kg, i.p.). (A) Day 19 (first day of L-DOPA) and (B) day 23 showed minimal difference at time points in dyskinesia severity. (C) Day 27, (D) day 31, and (E) day 35, approximately 1 week after treatment initiation, show DMI + L-DOPA treatment is worsening dyskinesia severity at later time points in the observation period. (F) By the end of the study on day 39, DMI + L-DOPA treatment worsens dyskinesia severity at all time points compared with L-DOPA alone. In all observation periods, DMI alone did not elicit any dyskinesia. Data analyzed by repeated ANOVA followed by Bonferroni post hoc test (time course). Significant difference from L-DOPA-only treatment: *P< .05, **P< .01, ***P< .001.
the result was normalized to synaptosome protein and expressed as pmol DA per mg protein per minute.

**Analyses of Proteins and ERK Phosphorylation.** Synaptosome fraction (P1) and the processed preparatory sample (P2) were sonicated in a 1% sodium dodecyl sulfate solution (pH ~8) using a Branson Sonifier 150 (Danbury, CT). Protein concentration was determined using the bicinchoninic acid colorimetric assay. Following gel electrophoresis, proteins were transferred for 500 V hours in a Tris/glycine/methanol buffer onto nitrocellulose membranes (Bio-Rad Laboratories, Hercules, CA).

The nitrocellulose membrane was stained with Ponceau S to further normalize staining in each sample lane. These lanes were scanned and quantified by Image J to normalize protein in each sample. This relative total level then served as an additional normalizing value to determine the quantity of each protein assayed (Salvatore et al., 2012). To continue processing, the membranes were blocked in PVP (Millipore; catalogue M8159). Protein loads for linear detection were 50 μg total protein for NET, 10 μg for TH and NET (Thermo Scientific, Santa Clara, CA) and ppERK1/2 (Sigma-Aldrich, St. Louis, MO; catalogue 437034). Protein loads for linear detection were 50 μg total protein for NET, 10 μg for TH and NET (Thermo Scientific, Santa Clara, CA) and ppERK1/2 (Sigma-Aldrich, St. Louis, MO; catalogue 437034). Protein loads for linear detection were 50 μg total protein for NET, 10 μg for TH and NET (Thermo Scientific, Santa Clara, CA) and ppERK1/2 (Sigma-Aldrich, St. Louis, MO; catalogue 437034).

**Statistical Analysis.** Data were analyzed using SPSS (Chicago, IL) with correction for multiple comparisons and P values < 0.05 considered significant. A two-way analysis of variance (ANOVA) (time × treatment) was used to analyze AIM scores over time within each session. Total AIM scores (axial + limb + oral) summed across the 60-minute sessions were analyzed by two-way ANOVA. One-way ANOVA and t-paired Student tests were used in instances with more than three or more groups or two groups, respectively. One-way ANOVA analyses were followed by post hoc Tukey multicomparison test.

Surgery were performed in triplicates with the assumption that any irregularities in efficacy of the 6-OHDA lesion may occur in all animals lesioned with the same 6-OHDA on that day. Specifically, the area under the curve reflects the accumulative LID score attained between the two groups. L-DOPA + DMI elicited greater dyskinesia than L-DOPA alone over the course of the 20-day treatment regimen (Fig. 3A). however, on day 19, when the first dose of L-DOPA was administered, DMI significantly attenuated LID severity (Fig. 4A). This attenuation of LID was not evident after day 5 of L-DOPA (or 23 days postlesion) (Fig. 4B).

**Fig. 3. Dyskinesia over time with L-DOPA versus L-DOPA + DMI treatment.** (A) Area under the curve (AUC) of dyskinesia elicited between day 4 until end of 20-day treatment regimen between L-DOPA and L-DOPA + DMI (paired Student t test, n = 5, *P < .05). (B) Linear regression of dyskinesia and time with L-DOPA and L-DOPA + DMI treatment (L-DOPA, r² = .30, P = .24; L-DOPA + DMI, r² = .82, P = .0028). Percentages reflect the difference in the AUC score between the two treatment groups as a function of L-DOPA administrations.

**Results**

**LID Severity Increases with DMI Pretreatment + L-DOPA Compared with L-DOPA Alone.** L-DOPA-induced AIMs were rated at six discrete time points (days 19, 23, 27, 31, 35, 39 post-6-OHDA lesion, or days 1, 5, 9, 13, 17, 21 of daily L-DOPA) during the 20-day administration of L-DOPA starting on day 1 (19 days post–6-OHDA lesion). As L-DOPA treatment duration increased, the NET inhibitor, DMI, exacerbated dyskinesia expression in 6-OHDA–lesioned male Sprague Dawley rats compared with L-DOPA alone beginning on day 27 and through day 39 post–6-OHDA lesion induction (Fig. 4). However, on day 19, when the first dose of L-DOPA was administered, DMI significantly attenuated LID severity (Fig. 4A). However, this attenuation of LID was not evident after day 5 of L-DOPA (or 23 days postlesion) (Fig. 4B).

**TH Loss in Treatment Groups.** There were no significant differences in TH loss among the three treatment groups as a function of L-DOPA administration. Thereafter, DMI exacerbated LID severity (Fig. 4, C–F). By the end of the study on day 39, DMI pretreatment with L-DOPA significantly exacerbated LID at all time periods of observation compared with L-DOPA treatment alone.

DMI alone (no chronic L-DOPA, 6-OHDA lesion only) did not elicit dyskinesia, supporting previous findings that DA replacement (with its precursor, L-DOPA) is a necessary component of L-DOPA–induced dyskinesia, despite lesion severity. The area under the curve reflects the accumulative LID score attained between the two groups. L-DOPA + DMI elicited greater dyskinesia than L-DOPA alone over the course of the 20-day treatment regimen (Fig. 3A). Because dyskinesia was not observed with DMI alone (Fig. 4, average AIM score = 0), a comparison of L-DOPA + DMI versus L-DOPA alone was made.

Additionally, LID severity did not change over time in the L-DOPA–alone group. However, in the L-DOPA + DMI group, there was a significant increase in dyskinesia over the 20-day course of treatment with L-DOPA + DMI, with the severity increasing as the study progressed (Fig. 3B).

**TH Loss in Treatment Groups.** There were no significant differences in TH loss among the three treatment groups as function of L-DOPA administration. Therefore, the exacerbation of LID severity in DMI + L-DOPA–treated rats most likely was not due to increased dopaminergic terminal destruction or lesion severity (L-DOPA, average TH loss 80.2% ± 4.9; DMI, 79% ± 5.3; L-DOPA + DMI, 77% ± 4.1).
Relative Abundance of NET: Lesioned versus Contralateral Striatum. Either DMI alone or DMI + L-DOPA treatment significantly decreased NET abundance in the lesioned striatum compared with contralateral control striatum in the respective groups (Fig. 6, A, B, and D). No difference in NET expression between control and lesioned striatum was observed in the L-DOPA group (Fig. 6, C and D).

ERK Phosphorylation in Treatment Groups. Levels of phosphorylated and total ERK1/2 were assessed in tissue fractions prepared from the 6-OHDA–lesioned striatum from rats from each treatment group. Both antibodies revealed two bands with the expected mol. wt. of ERK1 (44 kDa) and ERK2 (42 kDa). Accounting for ppERK1/2 in lesioned striatum, when compared with DMI alone, L-DOPA and DMI + L-DOPA treatment groups showed increased ppERK1/2 and ppERK/ERK2 (Fig. 7, A and B, respectively; representative Western blot, Fig. 7E). Interestingly, these changes were blunted in the control striatum (Fig. 7, C and D). ppERK/ERK2 was increased only in L-DOPA–treated rats compared with DMI (Fig. 7D). No differences were observed in total ERK between treatments (data not shown).

ERK Phosphorylation and Dyskinesia Severity. The levels of ppERK1 are increased in L-DOPA preclinical studies (Santini et al., 2007; Westin et al., 2007). We also observed a correlation between the severity of dyskinesia (as seen in the L-DOPA + DMI treatment group) and the ppERK1 signal, as measured in the lesioned striatum in each treatment group (Fig. 1), indicating that NET blockade in combination with L-DOPA may increase DA signaling associated with LID manifestation, as observed at the end of the study (day 21 of L-DOPA). Specifically, simple regression of phospho-ERK1 levels on the cumulative axial, limb, and orolingual global AIM scores recorded from chronically L-DOPA–, DMI + L-DOPA–, and DMI-treated rats had a positive correlation with dyskinesia severity (Fig. 1).

DA Uptake and NE Inhibition. DA uptake was determined in synaptosomes from lesioned versus control striatal tissue in the presence of a concentration of DA or NE (1 μM) previously shown to differentially inhibit uptake of labeled [3H]-DA (Chotibut et al., 2012). In the L-DOPA group, we did not observe a significant difference in inhibition [3H]-DA uptake by unlabeled DA between the lesioned and control striatum (percent inhibition in control ~70%) (Fig. 8A). However, NE inhibited DA uptake in lesioned striatum to a greater extent compared with that in control striatum (percent inhibition in control ~50%) (Fig. 8B). In the DMI + L-DOPA group, unlabeled DA did not differentially inhibit [3H]-DA uptake.
between lesioned and control striatum (percent inhibition in control ~70%) (Fig. 8C). However, NE was significantly less effective in inhibiting DA uptake in lesioned compared with control striatum (percent inhibition in control ~70%) (Fig. 8D).

NE also inhibited DA uptake to a significantly greater extent in lesioned striatum in the DMI-alone group (data not shown).

Discussion

Dysregulation of extracellular (DA) in the DA-denervated Parkinsonian striatum is associated with LID (Carta et al., 2006, 2007; Cenci and Lundblad, 2006). However, the relative contribution of DA uptake in LID onset or severity has not been established. The 6-OHDA lesion may increase striatal NET expression, and NET may affect DA uptake therein (Chotibut et al., 2012). In intact central nervous system (CNS) tissue, such as prefrontal cortex, there is evidence for NET-mediated DA reuptake (Carboni et al., 1990; Tanda et al., 1997; Moron et al., 2002). Accordingly, in lesioned striatum, the tissue content level of NE is comparable to that of DA (Chotibut et al., 2012), suggesting that remaining DAT and inherent NET are potentially in comparable abundance and...
NET may therefore assume a greater role in DA uptake. Our results are limited to striatal analysis because of the relative abundance of DA neuropil (although lesioned) over other DA regions like substantia nigra. DA uptake in the substantia nigra could also affect LID severity (Navailles et al., 2014), given significant noradrenergic innervation in this region. However, there is comparatively less DAT protein and DA uptake in the substantia nigra, which precluded us from examining this possibility.

L-DOPA may inhibit DA uptake in lesioned striatum (Hashimoto et al., 2005; Chotibut et al., 2012), which could lead to accumulation of extracellular DA, and therefore affect LID onset or severity. Increased extracellular DA in DA-denervated striatum is observed following DMI (Arai et al., 2008). This finding is congruent with our observations that reuptake of DA, derived from L-DOPA, may be modulated by NET. Thus, reducing NET function, either by L-DOPA blockade or reduced NET expression, could reduce DA clearance and exacerbate LID severity. Accordingly, we found that NET inhibition (with DMI) and L-DOPA gradually exacerbated LID severity over the 20-day course of L-DOPA administration, compared with L-DOPA alone (Fig. 3). These differences in LID severity were unrelated to differences in lesion severity (Fig. 5). Our previous finding that striatal NET expression increases at a comparatively earlier time point in lesion progression (Chotibut et al., 2012), coupled with the present findings, gives credence to involvement of NET in DA uptake and regulating LID expression or severity.

Our results also indicate that ERK1 and 2 phosphorylation was increased in the DMI + L-DOPA group compared with L-DOPA–alone group (Fig. 7). Increased signaling through D1 receptors is implicated in the molecular and synaptic responses in striatal neurons associated with LID onset (Picconi et al., 2003; Konradi et al., 2004; Aubert et al., 2005; Westin et al., 2007), and enhanced ERK1/2 phosphorylation in DA denervated striatum occurs with selective agonists for D1 or D2 receptors (Cai et al., 2000; Zhen et al., 2002; Pavon et al., 2006). Thus, increased ERK1/2 activity in striatal neurons is at the very least a biochemical marker of L-DOPA–induced dyskinesia through D1 or D2 overactivation. As such, phosphorylation of ERK1/2 may provide a molecular counterpart for increased D1 activity and be involved in LID induction. Reduced ERK1/2 phosphorylation dose-dependently decreases LID and other molecular correlates causally linked to LID development (Santini et al., 2007). Thus, the observation that DMI pretreatment with L-DOPA both exacerbates LID and increases ERK1/2 phosphorylation over that of L-DOPA alone supports a dopaminergic mechanism in LID severity wherein NET-mediated DA uptake and L-DOPA together modulate LID expression. Given that chronic DMI also reduced NET expression in lesioned striatum (Fig. 6), this observation suggests that increasing NET function or expression could reduce LID severity.

Compensatory changes in DA regulation do occur during the loss of DA-regulating proteins in 6-OHDA rodent models (Snyder et al., 1990; Sarre et al., 2004; Perez et al., 2008), but the potential involvement of NET function is a relatively novel concept. We have reported increased NET expression in lesioned striatum (Chotibut et al., 2012). Increased locomotion is observed in monkeys with DAT inhibitors with high NET but low serotonin transporter, affinity in cases of severe DAT loss (80%) compared with those with moderate DAT loss (46%) (Madras et al., 2006). LID severity could also be diminished by other interactions with noradrenergic inputs to the basal ganglia (Gomez-Mancilla and Bedard, 1993; Lundblad et al., 2002; Dekundy et al., 2007). Furthermore, rats with combined noradrenergic and dopaminergic lesions have greater LID severity, compared with dopaminergic lesions alone (Fulceri et al., 2007; Shin et al., 2014). Indeed, noradrenergic lesions can produce dyskinesias through DA-mediated locomotor impairment (Donaldson et al., 1976; Rommelfanger et al., 2007). As such, NET blockade via DMI may not be the only factor in worsening LID behavior. For example, in LID pathology, NE has the ability to act as a D1 dopaminergic agonist (Kubrusly et al., 2007). Thus, it may be possible that agents that decrease NET expression could lead to increased extracellular NE, thereby increasing D1 activation and worsening LID.

Reduced NET expression in the lesioned striatum therefore appears to play one of two critical components of LID and its severity. The other component of LID relates to the impact of chronic L-DOPA. DMI alone did not produce LID, but the combination of DMI and L-DOPA gradually worsened LID compared with L-DOPA alone. The neurobiological background of nigrostriatal lesion is also an important component. Chronic DMI reduced NET expression in lesioned striatum (Fig. 6), arguably offsetting any lesion-induced increase (Chotibut et al., 2012). Chronic DMI can reduce NET expression in other CNS regions such as amygdala, striatum (Jeannotte et al., 2009), and hippocampus (Kitayama et al., 2006). We also point out that the serotonin terminals or transporter (SERT) can affect LID severity in preclinical models (Carta et al., 2007; Eskow et al., 2007; Bishop et al., 2012). However, chronic DMI is not reported to alter SERT expression or function (Hyttel, 1994; Mantovani et al., 2009), thus making it unlikely that changes in SERT expression or SERT-mediated uptake are associated with our behavioral observations. From a clinical perspective, this leads to questions as to whether an antidepressant with NET affinity could produce, hasten the onset of, or worsen the severity of LID, given the prevalence of PD-related depression and that depression commonly precedes motor manifestations (Burn, 2002; Aarsland et al., 2012; Brichta et al., 2013). DMI or tricyclics are often prescribed for this patient population. In small-scale clinical trials (17 test subjects completing each
methylphenidate (which also affects NET function (Pan et al., 1994)) tended to increase LID (Devos et al., 2007; Espay et al., 2011). Exacerbation of dyskinesia in a clinical setting may go unnoticed given the absence of baseline readings before DMI administration. Therefore, further study could answer to the possibility that NET-blocking drugs could exacerbate LID severity or its frequency.

DMI may also interfere with alternate receptors such as muscarinic, adrenergic, and histamine. There is a functional interaction between the cholinergic and dopaminergic systems (Quik and Wonnacott, 2011), and DA denervation can increase acetylcholine levels in striatum. Given that acetylcholine mediates its effects via muscarinic and nicotinic receptors and that DMI may affect the sensitivity of post- and/or presynaptic muscarinic receptors (Murugaiah and Ukponmwan, 2003), it is possible DMI may be mediating its effects through muscarinic receptors, which may contribute to development of motor signs.

Behaviorally, we observe an initial delay in LID in the L-DOPA + DMI treatment group compared with L-DOPA, which may be related to increased NET expression with 6-OHDA lesion (Chotibut et al., 2012). Therefore, this initial increase in NET in the lesioned compared with the contralateral unlesioned striatum may explain why dyskinetic behavior was not worsened on the first day of L-DOPA (day 19 postlesion) and that subsequent L-DOPA, in conjunction with DMI treatments, offsets this increase in NET expression, revealing increased LID severity. Our DA uptake experiments (at the end of the LID assessment) also indicated that DA uptake is more inhibited by NE in the L-DOPA–alone group, but less inhibited by NE in the DMI + L-DOPA group (Fig. 8). We speculate that these differences may be related to NET expression. Whereas NET expression is not changed relative to control striatum in the L-DOPA group, it is decreased in the groups with DMI. Thus, this decrease in NE sensitivity in DMI groups could be due to decreased NET availability in lesioned striatum. We also speculate that the lack of difference in NET expression in rats treated with L-DOPA alone may be due to increased NET with 6-OHDA lesion that is offset by possibility that L-DOPA could reduce

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Fig. 8. Uptake of DA in presence of unlabeled DA or NE. DA uptake was measured in preparations of crude striatal synaptomes from the contralateral control and lesioned (>70% TH loss) striatum. Results are presented as the percent inhibition of tritiated DA uptake by 1 μM DA or NE. Note, the assessments of inhibition by unlabeled monoamine were conducted simultaneously in lesioned and control synaptosome preparations, one test subject per group at a given time. (A) DA inhibition of DA uptake in L-DOPA group. DA uptake (2 minutes) was determined by the parameters described in Materials and Methods. There was a significant increase (37%) in inhibition of DA uptake in the lesioned striatum by NE (***P < 0.05, Student’s two-tailed t test, t = 3.38; df = 4). (B) NE inhibition of DA uptake in L-DOPA group. DA uptake (2 minutes) was determined by the parameters described in Materials and Methods. There was a significant increase (37%) in inhibition of DA uptake in the lesioned striatum by NE (***P < 0.05, Student’s two-tailed t test, t = 3.38; df = 4). (C) DA inhibition of DA uptake in DMI + L-DOPA group. DA uptake (2 minutes) was determined by the parameters described in methods (N.S., Student’s two-tailed t test, t = 3.03; df = 3). (D) NE inhibition of DA uptake in DMI + L-DOPA group. DA uptake (2 minutes) was determined by the parameters described in Materials and Methods. There was a significant decrease (45%) in inhibition of DA uptake in the lesioned striatum by NE (***P < 0.01, Student’s two-tailed t test, t = 8.25; df = 3).
NET expression over time. As such, this is an important unresolved question moving forward.

The cellular sources of NET are an important point of consideration. Noradrenergic innervation of dorsal striatum is relatively sparse, although NE levels are comparable with remaining DA in lesioned striatum (Chotibut et al., 2012). In prefrontal cortex, NET transports DA and conversion of DA to NE may be slower than DA uptake via NET. As such, DA could be coreleased along with NE (Ahn and Kliman, 1989). Therefore, pharmacologically induced alteration of the noradrenergic system, such as by chronic DMI, could also influence extracellular DA levels (Devoto and Flore, 2006). Thus, our results may also be related compensatory alterations in DA release. Another possible source of NET-mediated DA transport may be glia, as glial express NET (Takeda et al., 2002). Astrocyte and microglia cell numbers may increase with DA neuron loss (Teismann and Schulz, 2004), possibly increasing NET protein abundance and DA uptake. Future studies examining alterations in DA uptake in glial preparations from dyskinetic rats may further delineate the cellular basis for NET-mediated DA uptake in LID pathophysiology.

Conclusion

In summary, our results indicate the possibility that increasing NET expression within the striatum may be a novel therapeutic avenue to attenuate LID severity. Our previous work has shown a upregulation of NET expression in lesioned striatum compared with contralateral striatum after DA denervation in the absence of L-DOPA treatment, but further work could establish whether this increase in NET is transient or alters with L-DOPA administration. Our results also indicate that L-DOPA must be present for any development of LID following 6-OHDA lesion. Our results add to a growing body of literature that NET regulates DA uptake and microglia cell numbers may increase with DA neuron loss may be glia, as glial express NET (Takeda et al., 2002). Astrocyte release. Another possible source of NET-mediated DA transport results may also be related compensatory alterations in DA renergic system, such as by chronic DMI, could also influence extracellular DA levels (Devoto and Flore, 2006). Therefore, pharmacologically induced alteration of the noradrenergic system, such as by chronic DMI, could also influence extracellular DA levels (Devoto and Flore, 2006). Thus, our results may also be related compensatory alterations in DA release.

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Norepinephrine Transporter and L-DOPA Dyskinesia


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