

**DISTINCT ROLES FOR PROTEIN KINASE C ISOFORMS IN REGULATING PLATELET
PURINERGIC RECEPTOR FUNCTION**

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RUNNING TITLE: P2Y₁ AND P2Y₁₂ ARE DIFFERENTIALLY REGULATED BY PKCS α AND δ

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Non-standard abbreviations: GPCR, G protein-coupled receptor; DMEM, Dulbecco's modified Eagle's medium; DNM, dominant negative mutant; PMA, phorbol 12-myristate 13-acetate; PBS, phosphate-buffered saline; ECL, enhanced chemiluminescence; HEPES, N-[2-hydroxyethyl] piperazine-N-[2-ethanesulphonic acid]; ACD, Acid citrate dextrose; IBMX, isobutyl methylxanthine; A3P5P, adenosine-3', 5'-diphosphate; 2MeSADP, 2-methylthioadenosine 5'-diphosphate.

ABSTRACT

ADP is a critical regulator of platelet activation, mediating its actions through two G protein-coupled receptors (GPCRs), P2Y₁ and P2Y₁₂. We had previously shown the receptors to be functionally desensitised, in a homologous manner, by distinct kinase-dependent mechanisms, where P2Y₁ is regulated by protein kinase C (PKC) and P2Y₁₂ by G protein-coupled receptor kinases (GRKs). Here we have addressed whether different PKC isoforms play different roles in regulating the trafficking and activity of these two GPCRs. Expression of PKC α and PKC δ dominant negative mutants in 1321N1 cells revealed that both isoforms regulated P2Y₁ receptor signalling and trafficking whilst only PKC δ was capable of regulating P2Y₁₂, in experiments where PKC was directly activated by the phorbol ester PMA. These results were paralleled in human platelets, where PMA reduced subsequent ADP-induced P2Y₁ and P2Y₁₂ receptor signalling. PKC isoform-selective inhibitors revealed that novel, but not classical, isoforms of PKC regulate P2Y₁₂ function, whilst both novel and classical isoforms regulate P2Y₁ activity. Importantly also we studied receptor internalisation in platelets by a radioligand binding approach, that showed that both receptors internalise rapidly in these cells. ADP-induced P2Y₁ receptor internalisation is attenuated by PKC inhibitors, whilst that of the P2Y₁₂ receptor is unaffected. Both P2Y₁ and P2Y₁₂ receptors can also undergo PMA-stimulated internalization, and here again novel but not classical PKCs regulate P2Y₁₂ whilst both novel and classical isoforms regulate P2Y₁ internalization. This study therefore is the first to reveal distinct roles for PKC isoforms in the regulation of platelet P2Y receptor function and trafficking.

INTRODUCTION

Activation of platelets occurs through a complex series of reactions in response to vessel injury and plays an essential role in thrombosis. One agonist, adenosine diphosphate (ADP), plays a central role in platelet activation by acting as a cofactor in the platelet responses to physiological agonists, including thromboxane A₂, collagen, and thrombin. ADP activates two surface expressed GPCRs, P2Y₁ and P2Y₁₂ (Gachet, 2005; Kunapuli et al., 2003). The combined stimulation of P2Y₁ receptor (coupled to G_q and PLC β) and P2Y₁₂ receptor (negatively coupled to adenylyl cyclase through G_i) is necessary for the full platelet aggregation response to ADP with platelet activation initiated by the P2Y₁ receptor and amplified by P2Y₁₂ (Gachet, 2005).

The attenuation of receptor-stimulated signal output upon sustained or recurrent agonist stimulation, a process known as desensitisation, is a crucial physiological mechanism of adaptation observed for many GPCRs. Since ADP plays a crucial role in platelet activation it likely that the responsiveness of P2Y₁ and P2Y₁₂ receptors is tightly regulated. Recently we showed that both P2Y₁ and P2Y₁₂ receptor responses desensitise in human platelets (Hardy et al., 2005) which may underlie the observed desensitisation of platelet responses following prolonged exposure to ADP (Baurand et al., 2000; Poole et al., 1993). Mechanisms underlying desensitization are complex and can involve phosphorylation of the receptor, uncoupling from G proteins, internalization, and ultimately intracellular down-regulation (Ferguson, 2001; von Zastrow, 2003). We have recently discovered that ADP pre-treatment promotes P2Y₁ and P2Y₁₂ receptor desensitization by different kinase-dependent mechanisms. P2Y₁, but not P2Y₁₂, desensitization is mediated by protein kinase C (PKC) (Hardy et al., 2005). In contrast, agonist-induced desensitization of the P2Y₁₂ receptor, but not P2Y₁, is largely dependent on G protein-receptor coupled kinases (GRKs) activity.

It is now important to address several important questions that are raised by our previous work: (i) since platelets express multiple isoforms of PKC (Buensuceso et al., 2005; Crosby and Poole, 2002; Murugappan et al., 2004), which isoforms are responsible for mediating homologous desensitisation of the P2Y₁ receptor? (ii) given the fact that platelets express multiple G_q-coupled receptors, each leading to

activation of PKC, can activation of PKC cause 'heterologous' regulation of P2Y₁₂, and if so which isoforms of PKC are involved?, (iii) do the receptors themselves become phosphorylated by PKC? Although our previous work had addressed only the roles of kinases in regulating functional desensitisation of the receptors, it is now also important to address their role in receptor internalisation and trafficking. In this study we address these questions and reveal distinct roles for PKC isoforms in the regulation of platelet P2Y receptor function and trafficking.

MATERIALS AND METHODS

Materials - Dulbecco's modified Eagle's medium (DMEM), Lipofectamine 2000 and fetal bovine serum were obtained from Life Technologies Inc. Radiochemicals were from Perkin Elmer Life Sciences. Complete protease inhibitor tablets were from Roche. Anti-HA-monoclonal antibody (HA-11), goat anti-mouse fluorescein-conjugated secondary antibody (1:200), rhodamine-conjugated transferrin and lysotracker red were purchased from Molecular Probes. The bisindolylmaleimide GF109203X (2-[1-(3-dimethylaminopropyl)-1H-indol-3-yl]-3-(1H-indol-3-yl)-maleimide), Gö6976 [12-(2-Cyanoethyl)-6,7,12,13-tetrahydro-13-methyl-5-oxo-5H-indolo(2,3-a)pyrrolo(3,4-c)-carbazole] and rottlerin were from Calbiochem (Merck Biosciences Ltd, Nottingham, UK). All other reagents were from Sigma.

Adenovirus infection and cell culture - Expression vectors for dominant negative mutant (DNM) PKC α (rabbit) and PKC δ (mouse) in adenoviruses were kind gifts from Professors Motoi Ohba and Toshio Kuroki, and have been described previously (Mitsutake et al., 2001; Ohba et al., 1998). Adenoviruses were amplified in HEK293 cells and harvested after 72 hours, purified and viral titers estimated as previously described (Kanegae et al., 1994). 1321N1 human astrocytoma cells stably expressing either hemagglutinin (HA)-tagged human P2Y₁ or P2Y₁₂ receptor were generated as previously described (Hardy et al., 2005). Cells were maintained in DMEM supplemented with 10 % fetal bovine serum, 100 units ml⁻¹ penicillin G, 100 μ g ml⁻¹ streptomycin sulfate and 400 μ g/ml geneticin at 37 °C supplemented with in a humidified atmosphere of 95 % air, 5 % CO₂. For adenoviral infection, 1321N1 cells were infected at between 50-200 pfu/cell in serum-free DMEM for 2 hours. Infection with empty vector β -gal adenovirus was used as control.

Receptor phosphorylation - Stably transfected cells in six well dishes were twice with phosphate-free DMEM and incubated for 90 min at 37 °C in the same media supplemented with 0.2 mCi/ml [³²P]

orthophosphate and 0.2 unit / ml apyrase. After incubation with the PKC inhibitor GF109203X (1 μ M; 15 min) or vehicle alone cells were treated with either ADP (10 μ M; 5 min) or the protein kinase C activator phorbol 12-myristate 13-acetate (PMA; 1 μ M; 15 min). Following drug treatment, reactions were terminated by placing the cells on ice and washing twice with ice cold PBS. All subsequent procedures were performed at 4 °C unless otherwise stated. Cells were subsequently lysed and HA-tagged receptor immunoprecipitated using a monoclonal anti-HA antibody (HA-11) as previously described (Mundell et al., 2004). Immune complexes were isolated by brief centrifugation, washed three times with immunoprecipitation buffer and eluted from beads by the addition of 20 μ l electrophoresis sample buffer. After fractionation by SDS-PAGE and transfer to a nitrocellulose membrane, phosphoproteins were visualised by autoradiography for 24-72 hr at -80 °C. Receptor immunoprecipitation was determined by reprobing membranes with a polyclonal anti-HA antibody / horse radish peroxidase-conjugated antirabbit IgG and visualization by enhanced chemiluminescence (ECL). The extent of receptor phosphorylation was quantified by densitometric analysis of resulting autoradiographs.

Western Blotting - Protein expression in 1321N1 cells and platelets was determined by western blotting. Briefly, cells were lysed into ice-cold lysis buffer (HEPES 20 mM, pH 7.4, NaCl 200 mM, EDTA 10 mM, 1% Triton X-100, supplemented with Complete protease inhibitors), insoluble material discarded by centrifugation (13,000g, 5 min) and sodium dodecyl sulphate (SDS) loading buffer (63 mM Tris, pH 6.5, 100 mM dithiothreitol, 1% SDS, 11.6% glycerol and 0.02% bromophenol blue) added to the cell lysates. Proteins were separated by SDS-PAGE. Gels were transferred to nitrocellulose membranes and blotted with PKC isoform specific antibodies for PKC α , β , (Zhang et al., 2001), δ and θ (Blass et al., 2002). Proteins were detected by enhanced chemiluminescence (ECL).

Measurement of cytosolic free calcium ($[Ca^{2+}]_i$) in 1321N1 astrocytoma cells - The cytosolic free Ca^{2+} concentration was determined using the fluorescent Ca^{2+} indicator fura-2-acetoxymethyl ester (fura-2/AM)

as previously reported (Hardy et al., 2005). Briefly transfected cells were grown on poly-L-lysine coated glass coverslips and used at ~ 60% confluence. Cells were washed twice with Locke's solution (154 mM NaCl, 5.6 mM KCL, 1.2mM MgCl₂, 2.2 mM CaCl₂, 5 mM HEPES, 10 mM glucose, pH 7.4) and incubated with fura-2/AM (3 μM) at 37 ° C for 60 mins. Glass coverslips were mounted into a quartz cuvette and placed into a thermostatically-controlled cell holder at 37 °C. Cells were continuously perfused with Lockes solution. Fluorescence was measured at 340 and 380 nM excitation and 510 nM emission. ADP (0.1 –1 μM) was perfused onto cell monolayers as required. [Ca²⁺]_i was determined from ratiometric data as previously described (Grynkiewicz et al., 1985).

Measurement of cAMP accumulation in 1321N1 astrocytoma cells - Cells infected with PKC-DNM adenoviruses, or β-gal alone controls, as described above were grown to 80% confluency and exposed to a desensitising dose of ADP (1 nM; 15 min) or PMA (1 μM; 15 min) in the presence of the phosphodiesterase inhibitor Ro201724 (250 μM). Apyrase (0.2 unit /ml) was then added directly to each well and incubated for 1 min at 37 °C to remove the desensitising ADP. Cells were then washed and forskolin (1 μM) added in the absence or presence of ADP and plates incubated at 37°C for 10 minutes. Endogenous β₂ adrenoceptor responses were also examined in 1321N1 cells by measuring isoproterenol (1 μM)-stimulated cAMP accumulation. Cyclic AMP accumulation was terminated by addition of ice cold 100 % trichloroacetic acid and supernatant neutralized with 1 M NaOH and TE buffer. Cyclic AMP levels were determined as previously described (Mundell et al., 1997). Data are expressed as cAMP production (pmol cAMP/well) or as % inhibition of forskolin stimulated adenylyl cyclase.

Internalization of HA-P2Y₁ and HA-P2Y₁₂ in 1321N1 cells - HA-tagged surface receptor loss was assessed by ELISA as described previously (Daunt et al., 1997; Mundell et al., 2000). Cells were split into 24-well tissue culture dishes coated with 0.1 mg ml⁻¹ poly-L-lysine. Twenty-four hours later, cells were incubated with DMEM containing apyrase (0.1 unit/ ml) for 1 hour at 37 °C, washed and then pre-treated with the

PKC inhibitor GF109203X (1 μ M; 15 min). Cells were then challenged with DMEM containing ADP (10 μ M; 15 min) or PMA (1 μ M; 15 min) at 37 °C. Changes in surface receptor expression were subsequently determined by an immunosorbent assay (ELISA) taking advantage of the HA-epitope tag (Daunt et al., 1997; Mundell et al., 2000), and expressed as either % surface receptor or % loss of surface receptor with the background signal from controls subtracted.

Preparation of human platelets - Human blood was drawn from healthy, drug-free volunteers on the day of the experiment. Acid citrate dextrose (ACD: 120 mM sodium citrate, 110 mM glucose, 80 mM citric acid, used at 1:7 vol/vol) was used as anticoagulant. Platelet rich plasma (PRP) was prepared by centrifugation at 200g, for 17 min and platelets were then isolated by centrifugation for 10 min at 1000g, in the presence of 0.02 U/ml apyrase and prostaglandin E₁ (PGE₁; 140 nM) for all assays other than measurement of intracellular cyclic AMP (cAMP) where PGE₁ was omitted. The pellet was resuspended to a density of 4x10⁸ platelets/ml in a modified Tyrodes-HEPES buffer (145 mM NaCl, 2.9 mM KCl, 10 mM HEPES, 1 mM MgCl₂, 5 mM glucose, pH 7.3). To this platelet suspension 10 μ M indomethacin and 0.02 U/ml apyrase were added, and a 30 min resting period was allowed before stimulation.

Measurement of cytosolic free calcium ([Ca²⁺]_i) in platelets - Measurement of cytosolic calcium was performed as previously described (Poole et al., 1995). Briefly, 3 μ M Fura-2-AM was added to platelet rich plasma, and incubated at 37 °C for 45 min in the presence of 10 μ M indomethacin. Platelets were centrifuged and re-suspended in modified Tyrodes. Platelets were treated for 15 min with the PKC inhibitors GF109203X (2 μ M), Gö6976 (1 μ M) Rottlerin (10 μ M) or vehicle alone. ADP (10 μ M)-induced calcium responses were subsequently measured at 37 °C in PMA (1 μ M; 15 min) and non PMA-treated platelets using a Hitachi F-4500 spectrofluorimeter with fluorescence excitation made at 340 nm and 380 nm and emission at 510 nm.

Measurement of cAMP levels in platelets - Platelets were treated for 15 min with the PKC inhibitors GF109203X (2 μ M), Gö6976 (1 μ M), rottlerin (10 μ M) or vehicle alone. PMA (1 μ M; 15 min) and non PMA-treated platelets were stimulated in the presence of the phosphodiesterase inhibitor IBMX (100 μ M) \pm forskolin (1 μ M) in the absence or presence of ADP (10 μ M) for 5 min at 37 °C. Cyclic AMP accumulation was terminated by addition of ice cold 100 % trichloroacetic acid (TCA) and samples were left to lyse on ice for 1-2 hrs. The resulting samples were spun at 4000 g for 5 min and the cAMP-containing supernatant neutralized with 1 M NaOH and TE buffer (50 mM Tris-HCl, 4 mM EDTA, pH 7.4). Cyclic AMP levels were subsequently determined in each sample using a binding assay as previously described (Mundell et al., 1997). Data are presented as either % inhibition of forskolin-stimulated adenylyl cyclase.

Radioligand binding in human platelets - In experiments assessing receptor internalization platelets were pre-treated with the PKC inhibitors with GF109203X (2 μ M), Gö6976 (1 μ M) Rottlerin (10 μ M) or vehicle alone. Platelets were subsequently stimulated with ADP (10 μ M 0-30 min), PMA (1 μ M 15 min) or vehicle alone. ADP was then removed by addition of 0.2 U/ml apyrase for 3 min prior to fixing platelets. Platelets were fixed by continuous rotation for 25 minutes in the presence of 4% formaldehyde. Platelets were then isolated by centrifugation at 1000g (10 min) and then resuspended in binding buffer (20 mM HEPES, 1 mM MgCl₂) to a density of 4x10⁸ platelets/ml. Aliquots of platelet suspension were incubated with [³H]-2MeSADP (3 Ci/mmol; 0.01nM-1 μ M) and specific binding determined in the presence of either unlabelled ligand (10 μ M), the P2Y₁ receptor antagonist A3P5P (1 μ M – 3.33 mM) or the P2Y₁₂ receptor antagonist AR-C69931MX (1 nM- 10 μ M). Following incubation for 20 minutes at room temperature reactions were terminated by addition of ice cold binding buffer and rapid filtration through Whatman GF/C glass fibre filters under vacuum. Radioactivity bound to the filters was measured by scintillation counting.

Experimental design and statistics - Data were analysed by the iterative fitting program GraphPAD Prism (GraphPAD Software). Log concentration-effect curves were fitted to logistic expressions for single-site analysis, whilst $t_{0.5}$ values for agonist-induced internalization were obtained by fitting data to single exponential curves. Where appropriate, statistical significance was assessed by Mann-Whitney-U test or by two-way ANOVA.

RESULTS

PKC-dependent P2Y₁ and P2Y₁₂ receptor phosphorylation - No studies to date have demonstrated the direct phosphorylation of either the P2Y₁ or P2Y₁₂ purinergic receptor although both contain multiple potential phosphorylation sites with their C-terminal domain. At this time the lack of specific P2Y₁ and P2Y₁₂ receptor antibodies able to consistently identify and immunoprecipitate receptor from platelets precluded studies of endogenous purinergic receptor phosphorylation. Therefore, we examined the phosphorylation of heterologously expressed HA epitope-tagged P2Y₁ and P2Y₁₂ receptors in P2Y null 1321N1 cells. A polyclonal anti-HA antibody recognised specific immunoreactive bands in membranes from both P2Y₁ and P2Y₁₂ receptor expressing cells (Fig. 1). These bands at 45kDa (P2Y₁) or 70 kDa (P2Y₁₂) were not present in vector alone pcNEO transfected controls, and run at apparent molecular weights consistent with other reports (Moran-Jimenez and Matute, 2000; Zhong et al., 2004). P2Y₁₂ receptor runs at a considerably higher apparent molecular weight than may be predicted from its primary amino acid sequence, due to extensive N-linked glycosylation (Zhong et al., 2004). P2Y₁ receptor has been shown to have an apparent molecular weight of 40-50 kDa by electrophoretic mobility (Moran-Jimenez and Matute, 2000). Phosphorylation studies revealed that both P2Y₁ and P2Y₁₂ receptors exist as phosphoproteins under basal conditions and that addition of either ADP (10 μ M; 5 min) or the PKC activator phorbol 12-myristate 13-acetate (PMA; 1 μ M; 15 min) significantly increased receptor phosphorylation. Pre-treatment with the PKC inhibitor GF109203X (1 μ M; 15 min) significantly attenuated ADP and PMA promoted P2Y₁ receptor phosphorylation. Therefore PKC can directly phosphorylate either the agonist occupied or unoccupied P2Y₁ receptor. In contrast GF109203X only attenuated PMA promoted P2Y₁₂ receptor phosphorylation. Therefore agonist-induced P2Y₁₂ receptor phosphorylation is not regulated by PKC although this kinase can directly phosphorylate the agonist unoccupied receptor.

PKC-dependent desensitization of P2Y₁ and P2Y₁₂ receptor activity in 1321N1 cells - To identify more accurately the PKC isoforms that may regulate P2Y₁ and P2Y₁₂ receptor function we examined their expression in both 1321N1 cells and human platelets. Western blotting revealed that although platelets expressed PKCs α , β , δ and θ , 1321N1 cells expressed only PKC α and PKC δ (Fig. 2A). We therefore subsequently over-expressed dominant negative catalytically inactive mutants of both of these isoforms in 1321N1 cells, by an adenoviral infection approach (Fig. 2B, where immunoblotting for both PKC α and PKC δ is shown), and examined effects of expression upon receptor signalling and internalisation. P2Y₁ receptor desensitisation was examined by monitoring the cytosolic calcium response to ADP, as we have previously shown (Hardy et al., 2005). Over-expression of either isoform-specific dominant negative mutant (DNM) did not significantly attenuate agonist-induced desensitisation (Fig. 3A). A reduction in P2Y₁ receptor desensitisation was only apparent in cells co-expressing both PKC α - and PKC δ -DNM (Fig. 3A). Pre-treatment with PMA (1 μ M; 15 min) significantly attenuated subsequent agonist-induced P2Y₁ receptor activity. As with the homologous desensitisation of P2Y₁ receptor activity, heterologous PMA-induced desensitisation was only attenuated when both PKC α - and PKC δ -DNM were co-expressed. Studies examining the G_i-coupled P2Y₁₂ purinergic receptor revealed that ADP-induced inhibition of forskolin-stimulated adenylyl cyclase (ADP induced an 80% inhibition of response, consistent with previous studies (Hardy et al., 2005)) was reduced following PMA (1 μ M; 15 min) pre-treatment to a similar extent to that found in ADP (10 nM; 15 min) desensitised cells (Fig. 3B). Actual numbers are shown in Fig. 3B(i) and normalised data is shown in Fig. 3B(ii). Data from Fig. 3B(i) also shows that PMA is not able to induce activation of adenylyl cyclase, nor inhibit forskolin-induced activity of the cyclase. Therefore PKC can regulate the agonist unoccupied P2Y₁₂ purinergic receptor in a heterologous manner. Over-expression of DNM-PKC δ , and not DNM-PKC α selectively attenuated PMA-induced P2Y₁₂ purinergic receptor desensitisation whilst ADP-induced desensitisation was unaffected by either DNM PKC.

PKC-dependent internalization of P2Y₁ and P2Y₁₂ receptors in 1321N1 cells - Using 1321N1 cells stably expressing N-terminal HA-epitope tagged versions of either receptor we are able to quantify agonist-induced surface receptor loss by ELISA (Mundell et al., 2004). Stimulation with either ADP (10 μ M; 5 min) or PMA (1 μ M; 15 min) induced internalization of both P2Y₁ and P2Y₁₂ receptors (Fig. 4). The rate of internalisation is shown in Fig. 4 A & B for P2Y₁ and P2Y₁₂ receptors respectively. Importantly also we show that carbachol (1 mM), operating through endogenously expressed muscarinic M₃ receptors, is able heterologously to induce internalisation of both P2Y₁ and P2Y₁₂ receptors. Pre-treatment with GF109203X (1 μ M; 15 min) selectively attenuated ADP-induced P2Y₁ receptor internalization whilst that of the P2Y₁₂ receptor was unaffected (Fig. 4C). As expected inhibition of PKC with GF109203X reversed internalisation of both P2Y₁ and P2Y₁₂ receptors induced by PMA. ADP and PMA-induced P2Y₁ receptor internalisation was partially inhibited by expression of either PKC α - or PKC δ -DNM, with co-expression of both DNMs producing a more robust inhibition of receptor internalisation (Fig. 4D). As with GF109203X pre-treatment, expression of PKC α - or PKC δ -DNM did not attenuate ADP-induced P2Y₁₂ receptor internalisation (Fig. 4D). Interestingly, consistent with their effects upon functional desensitisation, expression of DNM-PKC δ but not DNM-PKC α attenuated PMA-induced P2Y₁₂ receptor internalisation (Fig. 4D).

Regulation of P2Y₁ and P2Y₁₂ receptor desensitisation by PKC in human platelets - Since our studies in 1321N1 cells revealed that PKC could regulate P2Y₁ and P2Y₁₂ receptor function we sought to determine if this was also the case in human platelets. Recently we reported that, in response to ADP, PKC can regulate the desensitisation of P2Y₁ purinergic receptor responses in human platelets (Hardy et al., 2005). Therefore as expected pre-treatment with PMA resulted in a significant reduction in subsequent ADP-promoted P2Y₁ receptor-mediated calcium response (Fig. 5A). Inhibition of PKC with GF109203X (2 μ M), a non-selective PKC inhibitor significantly attenuated P2Y₁ receptor desensitisation. Pre-treatment with Gö6976 (1 μ M), which inhibits calcium-dependent classical PKC isoforms (Martiny-Baron et al.,

1993), including PKC α , and rottlerin (10 μ M), a PKC δ isoform selective inhibitor (Gschwendt et al., 1994), were unable to attenuate PMA promoted P2Y₁-receptor desensitisation (Fig. 5A). Importantly however, pre-treatment with both Gö6976 and rottlerin was able to partially attenuate the desensitisation of P2Y₁ receptor activity. Stimulation of PKC with PMA also decreased ADP-induced P2Y₁₂ receptor-mediated inhibition of forskolin stimulated adenylyl cyclase (Fig. 5B(i) shows actual numbers and Fig. 5B(ii) shows normalised data) to a level comparable to that induced by ADP pre-treatment (Hardy et al., 2005). Pre-treatment with either GF109203X or rottlerin attenuated PMA-induced P2Y₁₂ receptor desensitisation whilst Gö6976 had no effect on desensitisation of the P2Y₁₂ response (Fig. 5B). Therefore stimulation of PKC activity can promote the heterologous desensitisation of both P2Y₁ and P2Y₁₂ receptor responses in human platelets. Finally, in order to demonstrate that, in the platelet system the Gq-coupled P2Y₁ receptor may regulate P2Y₁₂, we chose to investigate the effect of P2Y₁ receptor blockade upon ADP-induced desensitisation of the P2Y₁₂ receptor response. Figure 6 shows that pre-treatment of platelets with the P2Y₁ receptor antagonist A3P5P (1 mM) does not attenuate P2Y₁₂-mediated inhibition of adenylyl cyclase, but that in the presence of A3P5P the ADP-induced desensitisation of the P2Y₁₂ receptor is reduced, indicating partial heterologous regulation of P2Y₁₂ by the P2Y₁ receptor.

Internalization of P2Y₁ and P2Y₁₂ receptors in human platelets - In order to study the internalization of purinergic receptors in human platelets we made use of the non-specific P2 ligand 2MeSADP (Takasaki et al., 2001). Importantly we chose to use formaldehyde-fixed platelets for our study. The reason for doing so was to be able to avoid complications to the binding assay produced by released ADP and ATP, which would compete with radioligand for binding sites. This has been a problem in other studies, and the use of fixed platelets to overcome this problem has been validated previously (Agarwal et al., 1989; Jefferson et al., 1988). Saturation binding experiments measuring [³H]-2MeSADP binding to fixed platelets in the presence and absence of unlabelled radioligand (10 μ M) indicated that there were 901 ± 41 [³H]-2MeSADP binding sites per platelet with an affinity of 4.9 ± 0.3 nM. Further saturation experiments using the P2Y₁ receptor antagonist A3P5P (1 mM) or the P2Y₁₂ receptor antagonist AR-C69931MX (1

μM) revealed two distinct binding populations of 184 ± 27 and 644 ± 11 [^3H]-2MeSADP binding sites per platelet, which represent the P2Y_1 and P2Y_{12} receptors respectively. Interestingly experiments using combined P2Y_1 and P2Y_{12} receptor antagonists estimated the number of binding sites to be 844 ± 57 , a number not significantly different to that obtained with unlabelled 2MeSADP. Further experiments using a fixed concentration of [^3H]-2MeSADP (100 nM) revealed that the concentrations of A3P5P and AR-C69931MX used in the saturation analysis studies were indeed maximal (data not shown), and that the respective IC_{50}s for these antagonists were determined to be 19.9 ± 1.2 mM and 45.0 ± 2.4 nM, values which correlate well with those reported by Takasaki et al. (Takasaki et al., 2001).

Therefore in subsequent experiments (Fig. 7) examining receptor internalization, platelets were incubated with [^3H]-2MeSADP (100 nM) in the presence of A3P5P (1 mM) or AR-C69931MX (1 μM) to give an estimate of either the P2Y_1 or P2Y_{12} surface binding sites. Following pre-treatment with ADP (10 μM ; 10 min), its subsequent removal with apyrase (0.2 unit /ml; 3 min) and platelet fixation, there was a clear reduction in [^3H]-2MeSADP binding to both P2Y_1 and P2Y_{12} receptors compared with non-pretreated or apyrase alone treated controls (Fig. 7A). Further studies showed that the P2Y_{12} receptor internalised much more rapidly than the P2Y_1 receptor although by 30 minutes the relative surface expression of each receptor was comparable (Fig. 7B). Therefore, as in 1321N1 cells, both the P2Y_1 and P2Y_{12} purinergic receptors internalise in a rapid agonist-dependent manner in human platelets.

PKC-dependent regulation of P2Y_1 and P2Y_{12} receptor surface receptor expression and internalization in human platelets - In order to address the role PKC may play in this process we used the PKC inhibitors, GF109203X, Gö6976 and rottlerin described above. Interestingly pre-treatment of platelets with any of these inhibitors significantly increased the number of P2Y_1 binding sites, with GF109203X the most potent (Fig. 7C). Pre-treatment with GF109203X or rottlerin also significantly increased the number of P2Y_{12} receptor binding sites whilst Gö6976 had no significant effect (Fig. 7C). Inhibition of PKC activity with GF109203X, rottlerin or Gö6976 selectively attenuated ADP (10 μM ; 5 min)-induced P2Y_1 receptor

internalisation whilst that of the P2Y₁₂ was unaffected (Fig. 7D) in agreement with our studies in 1321N1 cells (Fig. 4C). Stimulation of platelets with PMA (1 μ M; 15 min) significantly enhanced the internalisation of both the P2Y₁ and P2Y₁₂ receptor (Fig. 7D). Pre-treatment with each of the PKC inhibitors reduced PMA-promoted P2Y₁ receptor internalisation. Again only GF109203X or rottlerin significantly reduced PMA-promoted P2Y₁₂ surface receptor loss. Therefore, PKC can regulate the surface expression and internalisation of both P2Y₁ and P2Y₁₂ receptors in human platelets.

DISCUSSION

The activation of P2Y₁ and P2Y₁₂ purinergic receptors by ADP is critical for normal platelet function. In order to avoid inappropriate thrombosis the sensitivity of these receptors to agonist needs to be continuously regulated. To date, the molecular mechanisms regulating platelet P2Y₁ and P2Y₁₂ purinergic receptor signalling and surface receptor expression are relatively poorly understood. In this study investigating the regulation of these two clinically important GPCRs we find both are phosphorylated following activation of PKC and demonstrate that their surface expression and activity are tightly regulated by this family of kinases. We show that specific PKC isoforms can differentially regulate P2Y₁ and P2Y₁₂ receptor function, demonstrating some redundancy of PKC isoforms for regulation of P2Y₁ but lack of redundancy of PKC isoforms for regulation of P2Y₁₂, which is heterologously controlled by PKC δ . Although P2Y₁ and P2Y₁₂ purinergic receptors play an essential role in ADP-induced platelet activation and are important pharmacological targets in the treatment of arterial thrombotic disease (Foster et al., 2001; Gachet, 2005; Kunapuli et al., 2003) minimal studies have investigated the regulation of function of these two GPCRs. In a recent study (Hardy et al., 2005) we demonstrated for the first time that both P2Y₁ and P2Y₁₂ receptors desensitise in platelets and show that these receptors desensitise by different kinase-dependent mechanisms, where GRKs regulate the P2Y₁₂ receptor and PKC regulates agonist-induced desensitization of the P2Y₁ receptor in human platelets. Protein kinase C has been shown also to phosphorylate and regulate agonist-unoccupied receptors (Hipkin et al., 2000; Mundell et al., 2002; Xiang et al., 2001). Initially we demonstrated that both P2Y₁ and P2Y₁₂ receptors underwent ADP-induced phosphorylation and that activation of PKC by PMA also promoted phosphorylation of both receptor subtypes. To our knowledge this is the first demonstration that either of these GPCRs can be phosphorylated in an agonist-dependent or independent manner. The non-selective PKC inhibitor GF109203X attenuated ADP-stimulated P2Y₁ receptor phosphorylation consistent with our recent demonstration that PKC regulates agonist-induced P2Y₁ receptor activity (Hardy et al., 2005). Although PKC can activate GRK2 and 3 isoforms, promoting their translocation to the cell membrane (Winstel et

al., 1996), these kinases are unlikely to mediate agonist-independent PMA-dependent phosphorylation since GRKs only phosphorylate agonist-occupied receptors (Penn et al., 2000; Pitcher et al., 1998). We would therefore propose, from the present data, that PKC isoforms lie directly upstream of phosphorylation of P2Y₁ and P2Y₁₂ receptors, at least in 1321N1 cells. These experiments were undertaken in P2Y receptor null 1321N1 cells stably expressing epitope tagged versions of both receptors. The lack of specific and high affinity P2Y receptor antibodies with which to isolate either receptor from platelet cell membranes prevents similar studies in human platelets at this time. There are however a number of putative PKC phosphorylation sites located within the C-terminus of both receptors, including Thr³³⁹ in the P2Y₁ receptor, which regulates PKC-dependent desensitisation (Fam et al., 2003), and may be important as such in platelets.

Since phosphorylation of agonist-unoccupied receptors has been implicated in desensitisation and internalization of many GPCRs we sought to determine its functional significance and to identify the specific PKC isoforms responsible. Interestingly, PMA pre-treatment attenuated subsequent ADP-stimulated P2Y₁ and P2Y₁₂ receptor activity and promoted agonist-independent surface receptor loss, the first demonstration of heterologous regulation of these GPCRs. Over-expression of dominant negative mutant (DNM) catalytically inactive forms of PKC α and PKC δ , the two PKC isoforms common to both platelets and 1321N1 cells, revealed that both isoforms regulated the agonist and PMA-induced desensitisation and internalization of P2Y₁ receptors since an attenuation of receptor desensitisation and surface receptor loss was only evident on co-expression of both DNMs. ADP-induced P2Y₁₂ receptor desensitisation is regulated by GRKs and for this receptor we have ruled out any contribution from PKCs since expression of DNM PKC constructs did not have any effect (Hardy et al., 2005). Interestingly, however, we found that heterologous PMA-promoted desensitisation and internalization of P2Y₁₂ receptor function was regulated by PKC δ alone. Importantly also we now show that not only receptor-independent activation of PKC, but also activation of PKC by endogenous M₃ muscarinic receptors in 1321N1 cells is able to induce internalisation of both P2Y₁ and P2Y₁₂ receptors (Fig. 4A&B). Collectively these novel findings in 1321N1 cells demonstrate that PKC-dependent phosphorylation of both P2Y₁ and P2Y₁₂

receptors can significantly decrease receptor function and promote a rapid loss of surface receptor. It is unclear at this time whether PMA-promoted loss of surface receptor plays a significant role in reduced receptor responsiveness or if PMA-stimulated receptor phosphorylation can lead to direct receptor / G protein uncoupling. Detailed studies investigating which regions of these GPCRs regulate their internalization are planned. Such studies will allow us to make internalization deficient receptor mutants and thereby determine if surface receptor loss plays a significant role in reduced signalling output.

As PKC heterologously regulates P2Y₁ and P2Y₁₂ receptor function and trafficking in 1321N1 cells we next examined these phenomena in human platelets. Interestingly, as in 1321N1 cells heterologous activation of PKC reduced subsequent P2Y₁ and P2Y₁₂ receptor responsiveness. Also importantly we showed in Fig. 6 that in platelets P2Y₁ receptors contribute partially to desensitisation of P2Y₁₂ receptors in a heterologous manner. Since platelets lack a nucleus and significant protein synthetic machinery, approaches to disrupt or reduce endogenous protein function e.g. expression of dominant negative mutants are not viable at present. Therefore in order to determine the PKC isoforms that regulate purinergic receptor function we used three PKC inhibitors, GF109203X (2 μM), a potent inhibitor of conventional and novel PKC isoforms (Toullec et al., 1991), Gö6976 (1 μM), an inhibitor with IC₅₀s in the nanomolar range for calcium-dependent PKC isoforms (Martiny-Baron et al., 1993), including PKCα, and rottlerin (10 μM) which selectively inhibits the calcium-independent PKC isoforms, inhibiting PKCδ with an IC₅₀ of approximately 5 μM; 10-30 fold higher concentrations are required to inhibit conventional PKC isoforms (Gschwendt et al., 1994). Using these selective inhibitors we found that as in 1321N1 cells, classical and novel isoforms of PKC can regulate the heterologous desensitisation of P2Y₁ receptor activity whilst only PKCδ was capable of desensitising P2Y₁₂ receptor activity in an agonist-independent manner.

In order to examine changes in purinergic receptor surface expression in human platelets we used the P2Y receptor radioligand [³H]-2MeSADP in combination with the P2Y₁ receptor antagonist A3P5P and the P2Y₁₂ receptor antagonist AR-C69931MX. Our estimates of receptor number (see Results section) are

similar to those obtained by others (Baurand et al., 2000). This is important since we had chosen to use formaldehyde-fixed platelets for our study. In our study, following stimulation with ADP the number of binding sites for both P2Y₁ and P2Y₁₂ receptors was significantly reduced. Recent investigations, in agreement with our own, indicate that agonist pre-treatment with ADPβS also reduced the number of P2Y₁ receptor binding sites in stably transfected 1321N1 cells (Baurand et al., 2000; Baurand et al., 2005). This was paralleled in the study of Baurand et al. (2005) by ADP-induced internalisation of P2Y₁ receptors in platelets. By contrast however, P2Y₁₂ receptor surface expression was reported by these authors not to change following pre-treatment of 1321N1 cells with ADPβS, and only to internalise transiently and very rapidly upon treatment of platelets with 5 μM ADP. The data relating to the P2Y₁₂ receptor are therefore in contrast to those of the present study, where we show a more sustained internalisation in platelets and 1321N1 cells. The reasons for the discrepancy between our data and those of Baurand et al. (2005) are not clear, although there are a number of methodological differences between the two studies. First, it should be noted that since ADPβS is less potent at P2Y₁₂ receptors than P2Y₁ (Takasaki et al., 2001) and is a partial agonist at the P2Y₁₂ purinergic receptor (Cusack and Hourani, 1981), it may be unable to promote full internalization of this receptor (Clark et al., 1999). Additionally, the study of Baurand et al. (2005) used GFP-tagged receptor expressed in 1321N1 cells (Baurand et al., 2005), in contrast to the HA tag used in our study. The relatively bulky GFP tag may unpredictably alter signalling and trafficking properties of the receptor. The platelet studies also differ in that Baurand et al. (2005) use an immunogold transmission electron microscopic approach whereas the present study uses a radioligand binding approach. These methodological differences may explain the different results obtained for P2Y₁₂ receptors. Interestingly, in our study, pre-treatment with PMA also reduced P2Y₁ and P2Y₁₂ receptor surface expression. This is the first demonstration that heterologous activation of PKC can promote the internalisation of P2Y receptors in human platelets.

Interestingly in the absence of agonist treatment the surface expression of both receptors was increased following pre-treatment with inhibitors of PKC (Fig. 7C) and it seems likely therefore that basal PKC

activity is directly regulating surface receptor number. In 1321N1 cells following agonist-induced internalization, both the P2Y₁ and P2Y₁₂ receptors can subsequently recycle to the cell surface (data not shown). It is therefore possible that an attenuation of agonist-independent receptor internalisation by inhibition of PKC, coupled with recycling of receptor already present in endocytic compartments back to the cell surface, together leads to increased cell surface receptor number.

As with heterologous receptor desensitisation in platelets, PMA-dependent P2Y₁ receptor loss is regulated by both classical isoforms of PKC and PKC δ whilst only PKC δ regulates agonist-independent P2Y₁₂ receptor internalization. It is unclear why there is functional redundancy between PKC α and PKC δ in their ability to regulate P2Y₁ receptor function in human platelets. These two isoforms play different roles in platelet function (Crosby and Poole, 2003; Murugappan et al., 2005; Murugappan et al., 2004; Pula et al., 2005) and their different modes of activation may in turn determine their ability to phosphorylate downstream targets. It is also unclear why PKC δ specifically attenuates P2Y₁₂ receptor signalling. The activity of PKC isoforms is tightly regulated by multiple molecular mechanisms including interaction with binding-partner proteins (Poole et al., 2004). A greater understanding of the protein–protein interactions between particular PKC isoforms and membrane-associated anchoring protein, which serve to recruit the PKC isoforms to distinct subcellular compartments, in close proximity to receptor target substrates should prove useful in unravelling patterns of GPCR / PKC isoform specificity.

In conclusion, since ADP performs a pivotal role in the formation of stable platelet aggregates, the activity of purinergic receptors may maintain the delicate balance between rest and activation that underlies platelet sensitivity. Our results show for the first time that regulation of platelet purinergic receptor expression and activity by specific PKC isoforms may play a significant role in haemostatic function.

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FOOTNOTES

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FIGURE LEGENDS

Figure 1. Protein kinase C-mediated phosphorylation of HA-P2Y₁ and HA-P2Y₁₂. ³²P-labelled 1321N1 human astrocytoma cells stably expressing either HA-P2Y₁ (**A**), HA-P2Y₁₂ (**B**) or vector alone (pcNEO) were pretreated for 10 min with vehicle or 1 μM GF109203X before the addition of the PKC activator phorbol 12-myristate 13-acetate (PMA; 1 μM; 15 min), ADP (10 μM; 5 min) or vehicle alone. Phospho-HA-purinergic receptors were immunoprecipitated from membrane lysates and run on SDS-PAGE before transfer to nitrocellulose membranes. Specific phosphorylated bands at (**A**) 45 kDa (P2Y₁) or (**B**) 70 kDa (P2Y₁₂) not present in vector alone pcNEO transfected controls were subsequently identified by autoradiography. Similar amounts of receptor immunoprecipitation were confirmed (right hand panels (**A**) and (**B**)) by reprobing membranes with a polyclonal anti-HA antibody / horse radish peroxidase-conjugated antirabbit IgG and visualization by enhanced chemiluminescence (ECL).

Figure 2. PKC isoform expression in 1321N1 cells and human platelets. (**A**) Whole cell lysates from 1321N1 cells and human platelets were subjected to SDS-PAGE, followed by immunoblotting with PKC-isoform specific antibodies as detailed in “Materials and Methods”. Data shown are representative of three experiments. (**B**) 1321N1 cells were infected with βGal-PKCα-DNM, βGal -PKCδ-DNM or βGal adenovirus alone as control. Whole cell lysates from these cells were subjected to SDS-PAGE followed by immunoblotting with PKC-isoform specific antibodies as detailed in “Materials and Methods”. Bands therefore represent the sum of native wild type and heterologously expressed DNM-PKC. Data shown are representative of three experiments.

Figure 3. PKC-mediated desensitisation of P2Y₁ and P2Y₁₂ purinergic receptor responses in 1321N1 cells stably expressing each receptor. 1321N1 astrocytoma cells were infected with βGal-PKCα-DNM, βGal-PKCδ-DNM, both DNM constructs or βGal adenovirus alone (vector alone) as control. (**A**)

Desensitization of P2Y₁ receptors was assessed by comparing calcium responses to ADP (0.1 μM) before and after pre-treatment addition of ADP (1 μM; 2 min) or PMA (1 μM; 15 min) as detailed in “Materials and Methods”. Results are expressed as a % of desensitisation of response and data are mean ± S.E.M of at least three independent experiments. *Statistical significance at p<0.05 for data compared with respective vector alone control (Mann-Whitney U-Test). (B) Agonist (ADP; 10 nM)-dependent inhibition of forskolin (1 μM; 10 min)-stimulated adenylyl cyclase activity by P2Y₁₂ purinergic receptor activation after pre-treatment with ADP (10 nM; 5 min), PMA (1 μM; 15 min) or vehicle alone was subsequently determined. (B(i)) For vector alone control condition, data are presented as mean pmol cAMP mg⁻¹ protein ± S.E.M. (n≥4). (B(ii)) For all conditions including vector alone control, normalised data are presented as mean ± S.E.M of at least four independent experiments, expressed as % inhibition of forskolin-stimulated adenylyl cyclase. *Statistical significance at p<0.05 for data compared with respective non-pretreated agonist-induced inhibition of forskolin-stimulated controls (Mann-Whitney U-Test).

Figure 4. PKC-mediated internalization of P2Y₁ and P2Y₁₂ purinergic receptor responses in 1321N1 cells stably expressing each receptor.

(A) P2Y₁ receptor- or (B) P2Y₁₂ receptor-expressing 1321N1 cells were challenged with ADP (10 μM), the protein kinase C activator PMA (1 μM) or the muscarinic agonist carbachol (1 mM). Surface receptor loss was assessed by ELISA at various time points after addition of agonist. Data represent mean ± S.E.M. of five independent experiments. (C) P2Y₁ or P2Y₁₂ receptor-expressing 1321N1 cells were pretreated with the PKC inhibitor GF109203X (1 μM; 15 min) and subsequently challenged with ADP (10 μM; 30 min) or the protein kinase C activator PMA (1 μM; 30 min). (D) 1321N1 cells infected with βGal-PKCα-DNM, βGal-PKCδ-DNM, both DNМ constructs or βGal adenovirus alone as control, were subsequently challenged with ADP (10 μM; 30 min) or PMA (1 μM; 30 min). Surface receptor loss was assessed by ELISA. The data represent means ± S.E.M. of five independent experiments. *p<0.05 compared to respective controls (Mann Whitney U-test).

Figure 5. PKC-dependent desensitisation of purinergic receptor responses in human platelets.

Platelets were pretreated for 15 minutes with either the non-specific PKC inhibitor GF109203X (2 μ M), Gö6976 (1 μ M), which inhibits calcium-dependent PKC isoforms, including PKC α , rottlerin (10 μ M), a PKC δ isoform selective inhibitor and subsequently challenged with PMA (1 μ M; 15 min) or vehicle alone. **(A)** Desensitisation of P2Y₁ purinergic receptor responses was assessed by comparing peak calcium responses to 10 μ M ADP in platelets treated with PMA versus vehicle alone. Values are mean \pm S.E.M. of three independent experiments and results are expressed as the ADP response after a desensitising dose of PMA as a percentage of the control response. * $p < 0.05$ for data compared with PMA treatment without PKC inhibitor (control; Mann Whitney U-test). **(B)** Platelets were either pretreated with PKC inhibitors (as described above) or with vehicle alone (control) as indicated. Agonist (ADP; 10 μ M)-dependent inhibition of forskolin (1 μ M; 5 min)-stimulated adenylyl cyclase activity by P2Y₁₂ purinergic receptor activation after pre-treatment with vehicle alone (control) or PMA (1 μ M; 15 min) was determined. In **(B(i))**, data for control and PMA alone pre-treatment conditions are presented as mean pmol cAMP mg⁻¹ protein \pm S.E.M. (n=3). In **(B(ii))** values are shown as normalised data for all conditions including control and PMA pretreatments, and represent mean \pm S.E.M of three independent experiments expressed as % inhibition of forskolin-stimulated adenylyl cyclase. *Statistical significance at $p < 0.05$ for data compared with respective non-pretreated agonist-induced inhibition of forskolin-stimulated controls (Mann-Whitney U-Test).

Figure 6. Heterologous regulation of P2Y₁₂ receptor responses by P2Y₁ in platelets activated by

ADP. Platelets were pretreated with the P2Y₁-selective antagonist A3P5P (1 mM) or with vehicle alone (No pre-treatment) as control. Agonist (ADP; 10 μ M)-dependent inhibition of forskolin (1 μ M; 5 min)-stimulated adenylyl cyclase activity by P2Y₁₂ purinergic receptor activation was determined after pre-treatment with vehicle alone (control) or a desensitising addition of ADP (10 μ M; 5 min). In **(A)**, data are

presented as percentage inhibition of adenylyl cyclase activity induced by addition of ADP. In **(B)**, data are presented as the percentage desensitisation of the P2Y₁₂ receptor response seen after pre-treatment with ADP. Data are mean \pm S.E.M of three independent experiments. *Statistical significance at $p < 0.05$ for data compared with respective non-pretreated agonist-induced inhibition of forskolin-stimulated controls (Mann-Whitney U-Test).

Figure 7. Surface P2Y₁ and P2Y₁₂ receptor expression in human platelets is regulated by PKC. In experiments assessing receptor internalization platelets were stimulated with ADP (10 μ M) or vehicle alone for 5, 10 or 30 minutes. Platelets were incubated with [³H]-2MeSADP (100 nM) and specific receptor binding determined in the presence of either the P2Y₁ receptor antagonist A3P5P (1 mM; P2Y₁), the P2Y₁₂ receptor antagonist AR-C69931MX (1 μ M; P2Y₁₂) or a combination of both antagonists (Both). In **(A)** platelets were treated with ADP (10 μ M) for 10 minutes. Data are expressed as receptor specific [³H]-2MeSADP binding ([³H]-2MeSADP binding (absence of displacing ligand - presence of displacing ligand) DPM) and represent means \pm S.E.M. of three independent experiments. In **(B)** platelets were treated with ADP (10 μ M) for 0-30 minutes. Data are expressed as % surface receptor and represent means \pm S.E.M. of three independent experiments. In **(C and D)** platelets were pretreated for 15 minutes with either the non-specific PKC inhibitor GF109203X (2 μ M), Gö6976 (1 μ M), which inhibits calcium-dependent PKC isoforms, including PKC α , rottlerin (10 μ M), a PKC δ isoform selective inhibitor In **(C)** P2Y₁ and P2Y₁₂ surface receptor expression was compared in platelets pretreated with PKC inhibitors versus non-pretreated controls. Data are expressed as % increase in surface expression represent means \pm S.E.M. of three independent experiments. In **(D)** platelets were subsequently challenged for 15 minutes with either ADP (10 μ M), PMA (1 μ M) or vehicle alone. Data are expressed as % loss of surface receptor and represent means \pm S.E.M. of three independent experiments. * $p < 0.05$ for data compared with ADP or PMA treatment without PKC inhibitor (vehicle; Mann Whitney U-test).

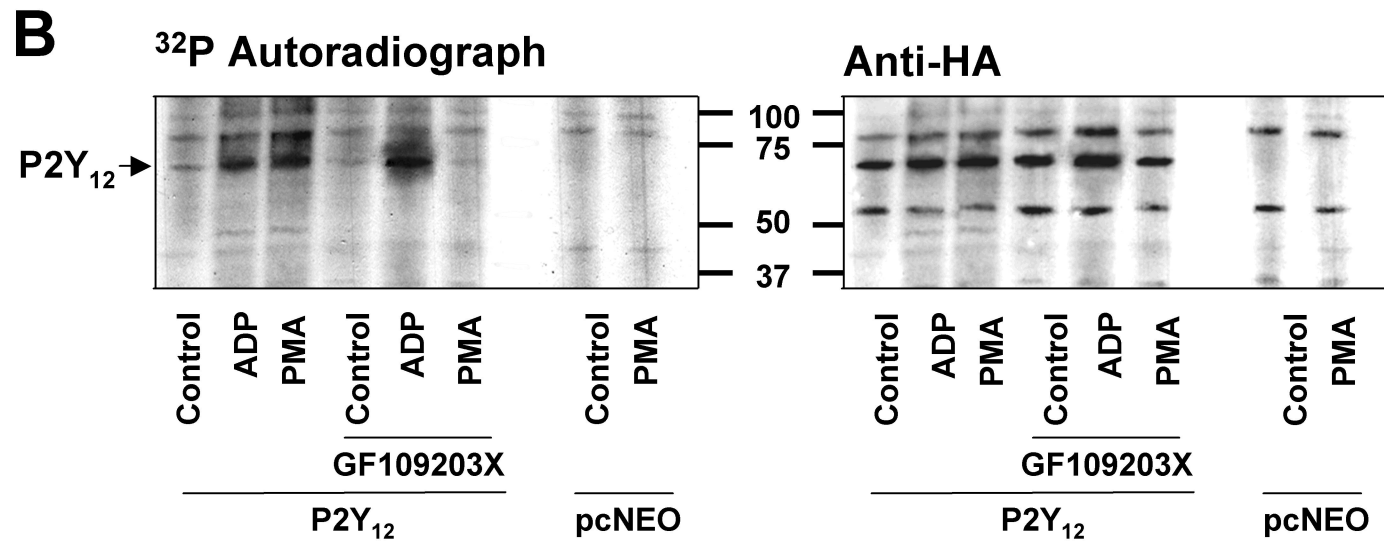
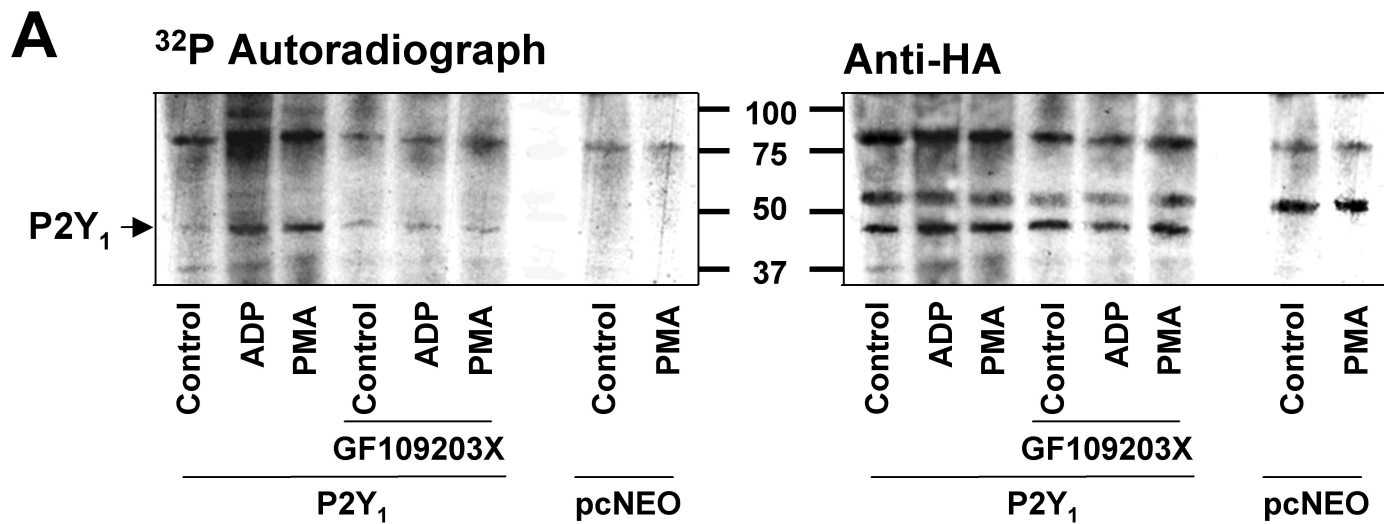
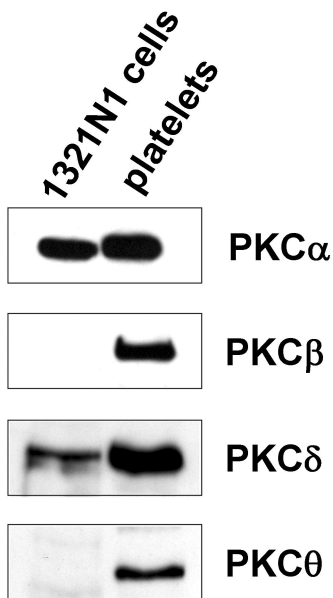
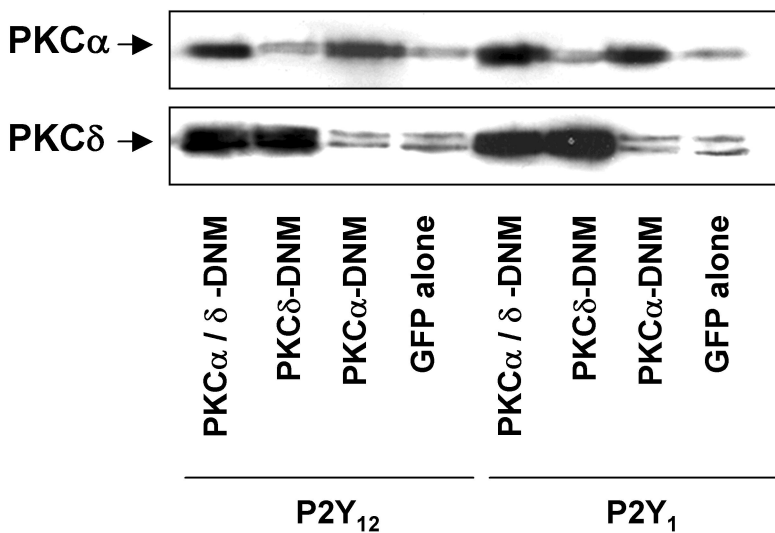
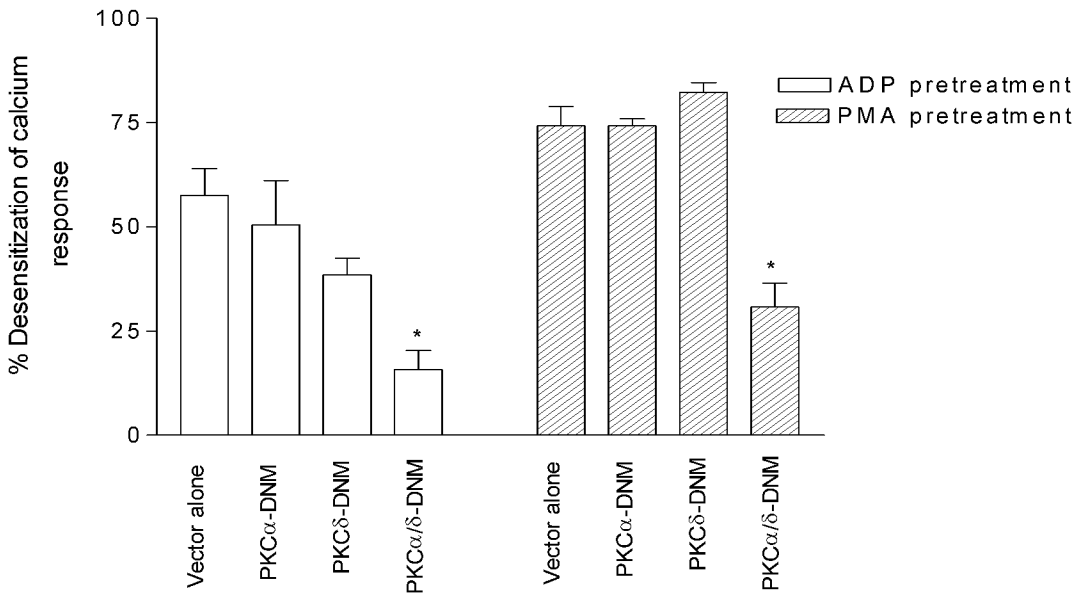
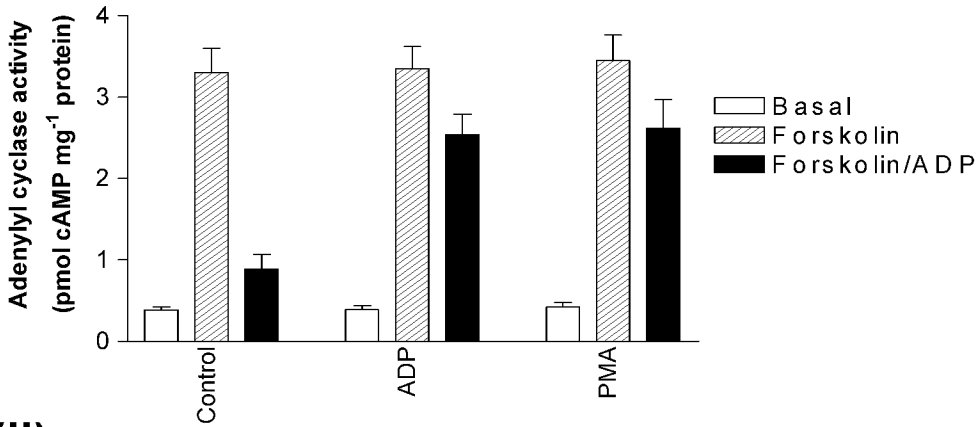
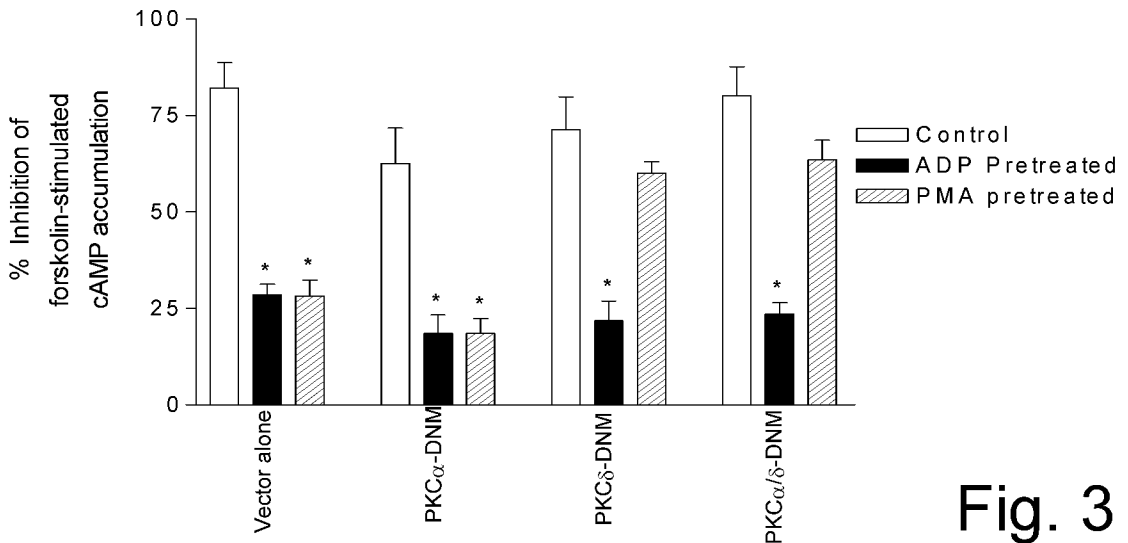


Fig. 1

A**B****Fig. 2**

A**B (i)****(ii)****Fig. 3**

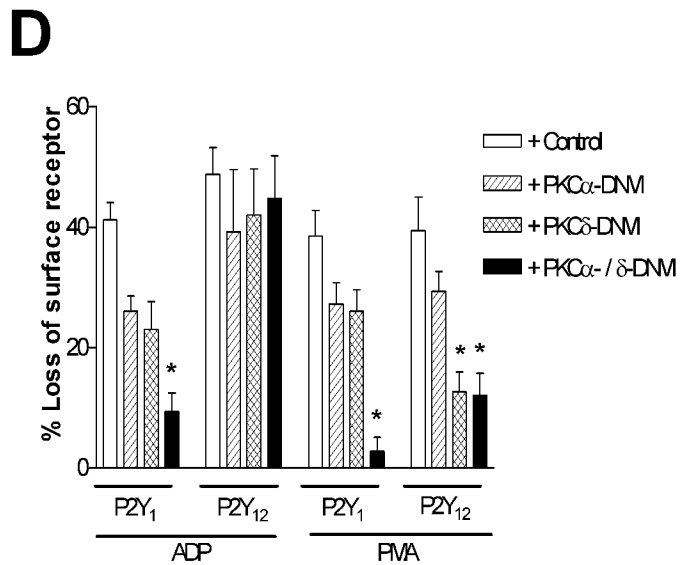
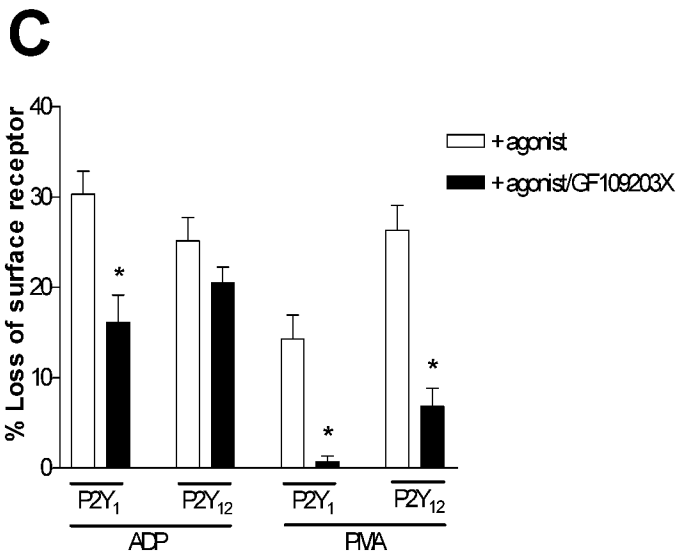
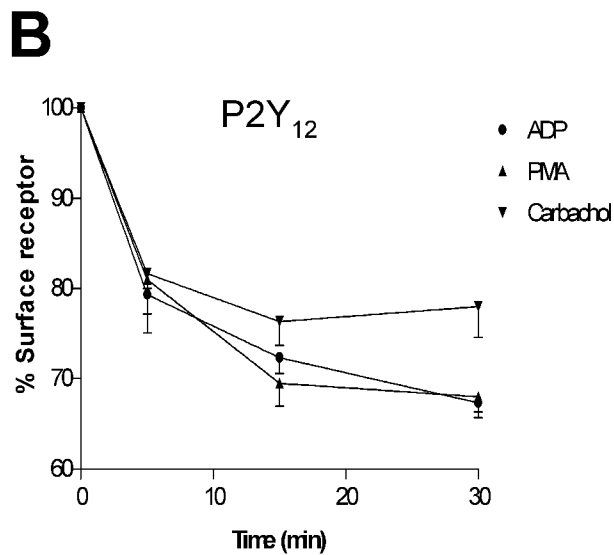
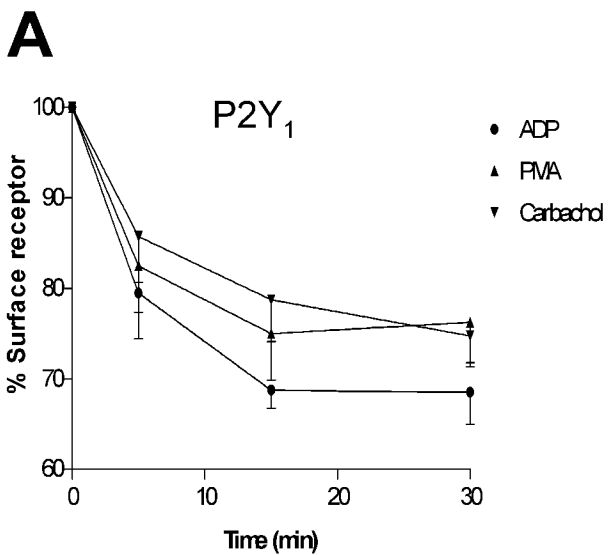
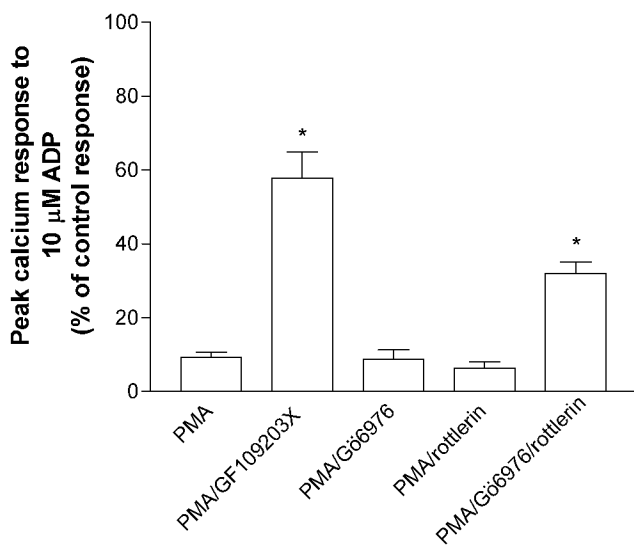
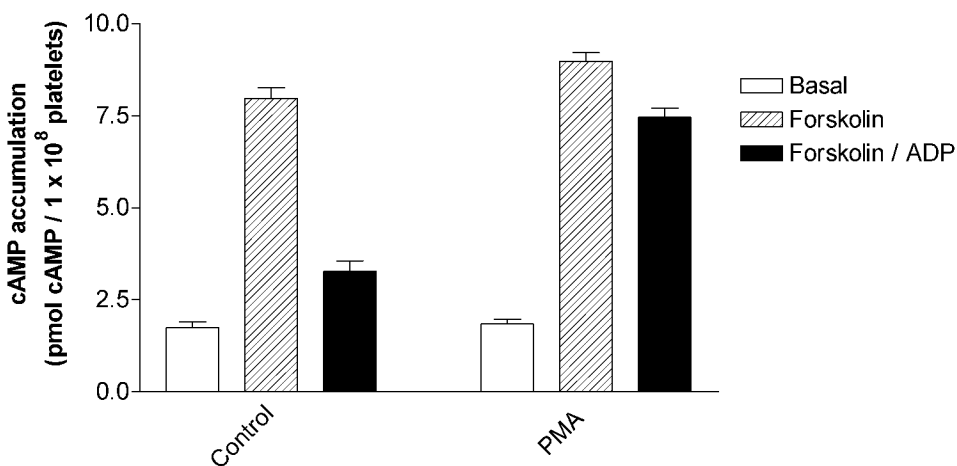
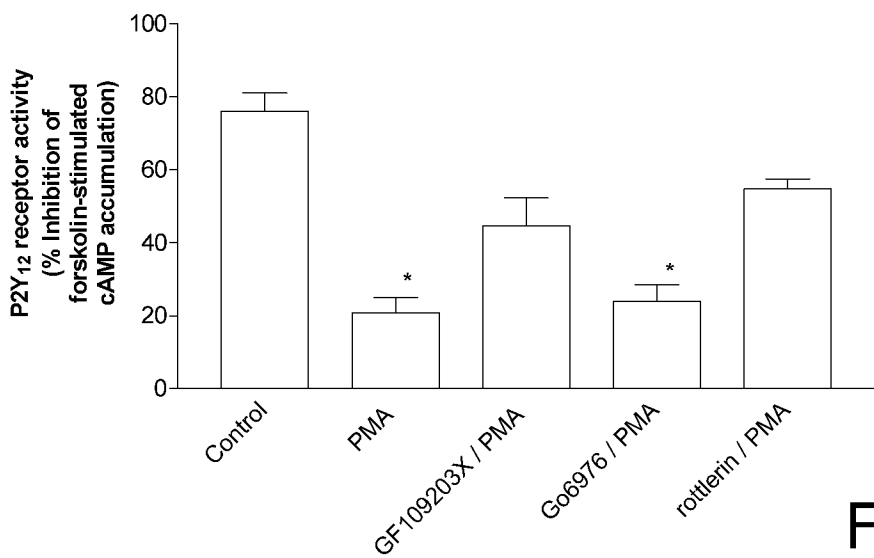
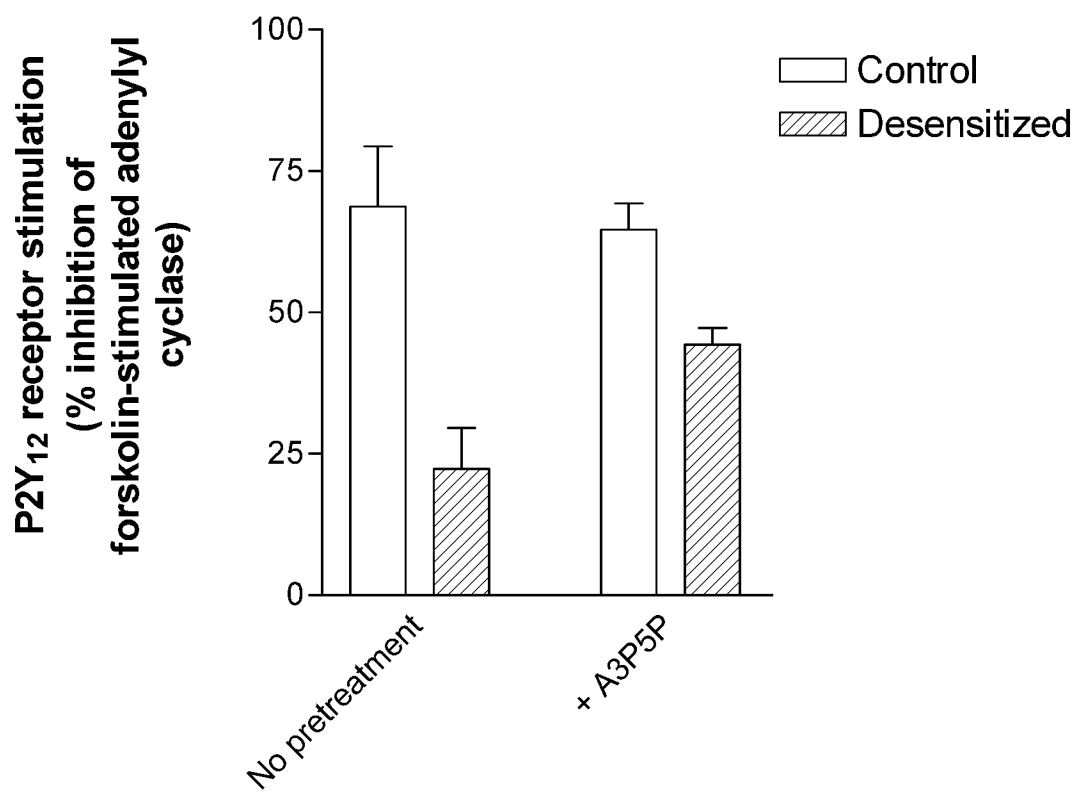
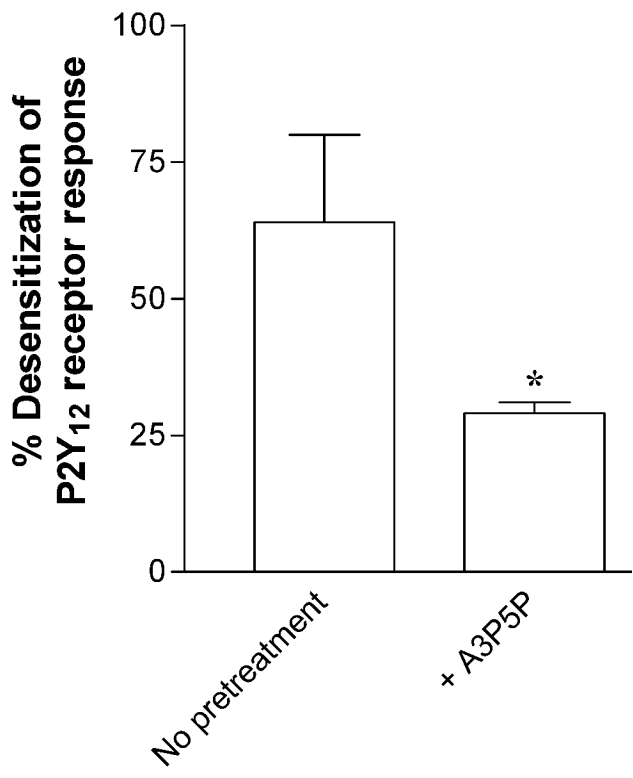
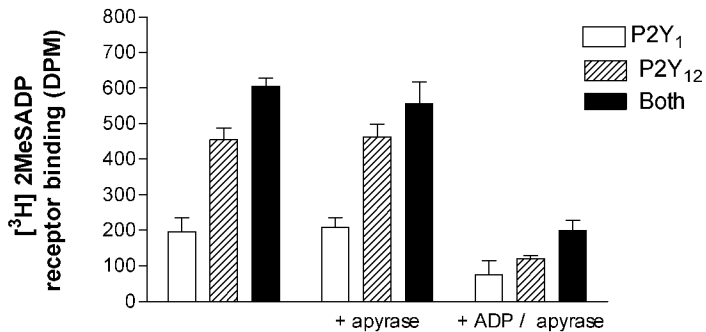
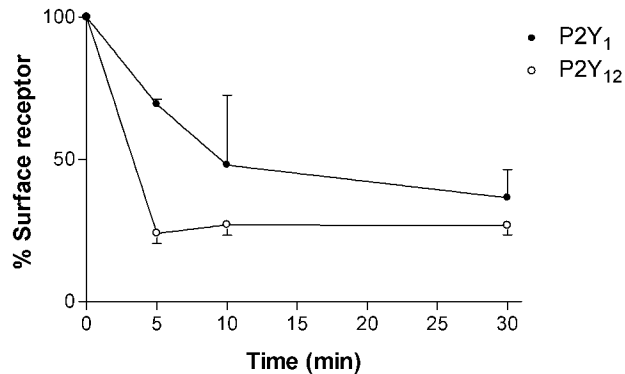
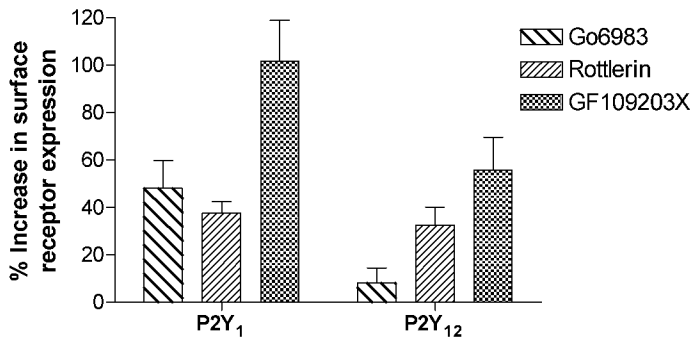
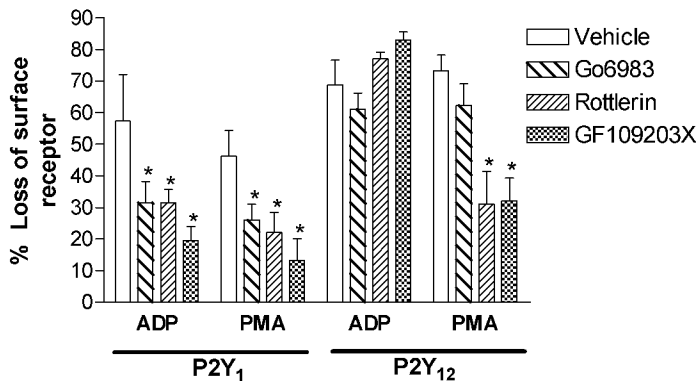


Fig. 4

A**B (i)****(ii)****Fig. 5**

A**B****Fig. 6**

A**B****C****D****Fig. 7**