

## **Controls of Tonic and Phasic Dopamine Transmission in the Dorsal and Ventral Striatum**

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**ABBREVIATIONS:**  $\beta 2^*$ ,  $\beta 2$  subunit containing; DA, dopamine; DAT, dopamine transporter; DH $\beta$ E, dihydro- $\beta$ -erythroidine; nAChR, nicotinic acetylcholine receptors; NAc, nucleus accumbens; SNc, substantia nigra pars compacta; VTA, ventral tegmental area;

## ABSTRACT

Dopamine (DA) release varies within subregions and local environments of the striatum, suggesting that controls intrinsic and extrinsic to the DA fibers and terminals regulate release. While applying fast-scan cyclic voltammetry and using tonic and phasic stimulus trains, we investigated regulation of DA release in the dorsolateral to ventral striatum. The ratio of phasic to tonic evoked DA signals varied with the average ongoing firing frequency, and the ratio was generally higher in the nucleus accumbens (NAc) compared to the dorsolateral striatum. At the normal average firing frequency, burst stimulation produces a larger increase in the DA response in the NAc than the dorsolateral striatum. This finding was comparable whether the DA measurements were made using *in vitro* brain slices or were recorded *in vivo* from freely-moving rodents. Blockade of the dopamine transporters and dopamine D<sub>2</sub> receptors particularly enhanced the tonic DA signals. Conversely, blockade of nicotinic acetylcholine receptors (nAChRs) containing the  $\beta_2$  subunit ( $\beta_2^*$ ) predominantly suppressed tonic DA signals. The suppression of tonic DA release increased the contrast between phasic and tonic DA signals and that made the frequency dependent DA dynamics between the dorsolateral striatum and NAc more similar. The results indicate that intrinsic differences in the DA fibers that innervate specific regions of the striatum combine with (at least) DA transporters, DA receptors, and nAChRs to regulate the frequency dependence of DA release. A combination of mechanisms provides specific local control of DA release that underlies pathway specific information associated with motor and reward related functions.

## Introduction

Dopamine (DA) neurons operate in distinct tonic and phasic timescales to differentiate behaviorally relevant information (Schultz, 2007). DA neurons discharge tonically at low frequencies that consist of individual action potentials without bursts (Grace and Bunney, 1984). Periodically, DA neurons fire in phasic bursts of near 20 Hz and greater (Hyland et al., 2002; Robinson et al., 2004). Evidence indicates that phasic or burst firing induces greater extracellular DA release compared to tonic, single-spike firing activity (Floresco et al., 2003; Gonon, 1988; Grace, 1991). Those tonic and phasic signals arise from midbrain DA neurons of the substantia nigra pars compacta (SNc) and ventral tegmental area (VTA) that innervate the whole dorsal to ventral extent of the striatum. Although the DA neurons that project to the prefrontal cortex may have higher discharge rates (Chiodo et al., 1984; Lammel et al., 2008), many midbrain DA neurons often exhibit similar overall firing properties (Clark and Chiodo, 1988; Gariano et al., 1989; Robinson et al., 2004; Schultz, 1986). Reward-related sensory input, however, such as that initiated by an addictive drug, enhances DA release to varying degrees depending on the dopaminergic pathway and target region (Di Chiara et al., 2004; Janhunen and Ahtee, 2007; Nisell et al., 1997; Pontieri et al., 1996; Shi et al., 2000). These findings suggest that there are local processes that regulate the decoding of reward related DA impulses and, thereby, modulate DA release locally.

The relationship between stimulus frequency and DA release varies depending on the local area of interest (Cragg, 2003; Garris and Wightman, 1994; Montague et al., 2004; Wu et al., 2002; Zhang and Sulzer, 2004). One possible explanation for this variability is that DA neurons with similar firing properties have different frequency-dependent DA release depending

on the region where they project, which in turn correlates with their anatomical location in the VTA or SNc. Indeed, regional differences in DA release probability and short-term plasticity within the striatum have been reported (Cragg, 2003; Exley et al., 2008). In this study, we analyze tonic and phasic DA release along the dorsal to ventral extent of the striatum while investigating the regulation of release by the dopamine transporter (DAT), the dopamine D<sub>2</sub> autoreceptor, and the  $\beta_2$ -containing ( $\beta_2^*$ ) nicotinic acetylcholine receptor (nAChR).

Dopamine influences glutamatergic afferents and striatal medium spiny neuron efferents, thereby, modulating striatal output and behavioral consequences (Horvitz, 2002; Nicola et al., 2004). This study was spurred by the hypothesis that the contrast between phasic and tonic DA signals contributes to the functional distinctions between the dorsolateral and ventral striatum. We examined regional differences in the dynamics between tonic and phasic DA signals, and examined how the activity of endogenous factors influences DA signaling dynamics. Those factors included the DATs, the dopamine D<sub>2</sub>-type receptors, and the  $\beta_2^*$  nAChRs. The onset, severity, and progression of neurological and psychiatric disorders such as Parkinson's disease and schizophrenia depend on DA signaling. Differences in the regulation of DA release in the dorsal striatum versus the NAc suggest that specific targets for therapeutic drug development may allow mesostriatal and mesolimbic DA signaling to be manipulated separately to aid different patient populations.

## Materials and Methods

***In Vitro* Brain Slice Voltammetry Experiments.** Wild-type C57BL/6J (Jackson Laboratory, Bar Harbor, Maine) and nAChR  $\beta_2$ -subunit null mice were used at 3 to 6 months of age. The  $\beta_2$ -subunit null mice were produced and characterized previously (Xu et al., 1999).

Mice were housed and handled in accordance with the guidelines set forth by the animal care committee at Baylor College of Medicine. Under deep anesthesia (a combination of ketamine, xylazine, and acepromazine), mice were decapitated and the brains were rapidly dissected out. Sagittal or horizontal slices (350  $\mu\text{m}$ ) were cut on a vibratome, incubated at  $32 \pm 0.5$  °C for 30 min, then held at room temperature for >30 min, and studied at  $34 \pm 1$  °C in 125 mM NaCl, 2.5 mM KCl, 1.3 mM MgCl<sub>2</sub>, 2.5 mM CaCl<sub>2</sub>, 26 mM Na<sub>2</sub>HPO<sub>3</sub>, 1.25 mM NaHCO<sub>3</sub> and 10 mM glucose saturated with 95% O<sub>2</sub> and 5% CO<sub>2</sub>.

Fast-scan cyclic voltammetry was performed using homemade carbon-fiber microelectrodes (10  $\mu\text{m}$  diameter and approximately 100  $\mu\text{m}$  exposed length; P55s, Amoco Polymers, Greenville, SC) that were placed in the striatum. This study focused on the dorsolateral striatum and the NAc core and shell. The carbon-fiber electrode potential was linearly scanned (12 ms duration, 10 Hz) from 0 to -400 to 1000 to -400 to 0 mV against a silver/silver chloride reference electrode at a rate of 300 mV/ms. An Axopatch 200B amplifier, a Digidata 1320 interface, and a pClamp 8 system (Axon Instruments Inc., Union City, CA) were used to acquire and analyze data. The voltammograms were sampled at 50 kHz, and the background current was subtracted digitally. The peak oxidation currents for DA in each voltammogram (at approximately 600 mV) were converted into concentration from a post-experiment calibration against fresh solutions of 0.5 to 10  $\mu\text{M}$  dopamine.

Intrastriatal stimuli were delivered using a bipolar tungsten electrode. The two tips of the stimulating electrode were about 150  $\mu\text{m}$  away from each other. The tip of the carbon-fiber recording electrode was 100-200  $\mu\text{m}$  away from the two tips of the stimulating electrode. The average intraburst firing rate of DA neurons is approximately 20 Hz in rodents (Benoit-Marand et al., 2001; Hyland et al., 2002; Zhang et al., 2009), and thus the relationship between the

number of pulses (1 to 20) within a stimulus train and the DA release concentration was commonly determined using a 20 Hz stimulation frequency. Each pulse was 0.5–1 ms in duration. After a stable control recording ( $\geq 30$  min), the slices were exposed to a single concentration of GBR 12909 (2  $\mu\text{M}$ ), sulpiride (2  $\mu\text{M}$ ), or dihydro- $\beta$ -erythroidine (DH $\beta$ E, 0.1  $\mu\text{M}$ ) for 20 min before data were collected for drug-induced changes in DA release.

Because stimulus trains of varying length and frequency were used, the release of DA was often spread in time during the length of the stimulus train. To be able to compare the diverse DA release and uptake patterns arising from these stimulus trains, we often quantified the DA signal by using the area under the curve of the DA amplitude plotted against time ( $\mu\text{M}\cdot\text{s}$ ). The relative DA signal was calculated by comparing the burst evoked DA signal to the single-pulse evoked DA signal. The voltammetric recordings from the dorsolateral striatum and NAc shell in the same mouse were used to determine the ratio of DA release for the two subregions. For comparison, some experiments also were performed using the NAc core, which displayed DA signaling characteristics intermediate between the dorsolateral striatum and the NAc shell. All data are means  $\pm$  SEM. Comparisons between differences in means were assessed by paired *t*-tests or one-way ANOVA.

***In Vivo* Fast-scan Cyclic Voltammetry Experiments.** Rats were implanted with carbon fiber microelectrodes for *in vivo* fast-scan cyclic voltammetry. Surgical preparation used an aseptic technique and anesthesia following the University of Washington Institutional Animal Care and Use Committee guidelines. Briefly, rats were anesthetized with isoflurane, administered the long-acting, non-steroidal anti-inflammatory, carprofen (5 mg/kg, subcutaneously) and placed in a stereotaxic frame. The scalp was swabbed with 10% povidone iodine, bathed with a mixture of lidocaine (0.5 mg/kg) and bupivacaine (0.5 mg/kg), and incised



to expose the cranium. Holes were drilled and cleared of dura mater above the NAc (1.3 mm lateral and 1.3 mm rostral from bregma) and the dorsolateral striatum (3.0 mm lateral and 1.3 mm rostral from bregma) and at a convenient location for a reference electrode. Carbon-fiber recording microelectrodes were lowered into the target regions (7.0 mm and 4.5 mm ventral of dura mater for the NAc core and the dorsolateral striatum respectively). Placement of the microelectrodes was verified by post-experiment histology.

After recovery from surgery, rats were food restricted and maintained at 90% of free-feeding body weight for the duration of the experiment. Electrochemical recordings were made in standard operant chambers (Med Associates) equipped with a food receptacle located at the center of the side wall. Food-evoked DA release was obtained by delivering an uncued food pellet to the receptacle.

Changes in DA concentration were assessed by fast-scan cyclic voltammetry in awake, behaving rats (Phillips et al., 2003). DA was recorded concurrently from the NAc and the dorsolateral striatum using the chronically implanted carbon-fiber microelectrodes. The potential at a carbon fiber microelectrode was held at -0.4 V versus the reference electrode and, then, ramped to +1.3 V and back to -0.4 V (400 V/s) every 100 ms (10 Hz). The cyclic voltammograms provided a chemical signature that enabled identification of DA relative to standards. Waveform generation, data collection, and analysis were carried out on a data acquisition system (National Instruments, Austin, TX) coupled to a miniaturized head-mounted voltammetric amplifier.

## Results

### **Regional Differences in DA Release in response to Tonic and Phasic Stimulation.**

In mouse brain slices, DA signals were evoked by a single-pulse stimulus (1p) or by a stimulus train of 5 pulses (5p) or 20 pulses (20p) delivered at 20 Hz. The amplitude of the DA signal evoked by 1p decreased as the voltammetry measurements moved down ventrally from the dorsolateral striatum (Fig. 1a), to the NAc core (Fig. 1b), to the NAc shell (Fig. 1c). The average DA signals to 1p are shown in Fig. 1d. The location with the smallest DA signal to 1p showed the greatest increase in the DA signal to 20p at 20 Hz (Fig. 1a,b,c at bottom). The degree of frequency dependent facilitation was represented as the ratio of the DA signal evoked by 20p delivered at 20 Hz to the DA signal evoked by 1p,  $[DA]_{20p} / [DA]_{1p}$  (Fig. 1e). That ratio was inversely related to the DA signal to 1p (Fig. 1d). In the NAc shell, burst stimulation (20p at 20 Hz) caused a rapid increase in the DA release that was larger than in the dorsolateral striatum:  $4.7 \pm 0.7 \mu\text{M}\cdot\text{s}$  in NAc shell and  $2.9 \pm 0.5 \mu\text{M}\cdot\text{s}$  in dorsolateral striatum ( $n = 18\text{--}19/\text{group}$ ,  $p < 0.05$ ). This finding indicates that the NAc shell has greater intrinsic ability to respond to burst firing activity (Cragg, 2003; Exley et al., 2008).

To explore tonic and phasic DA neuron firing activity in greater detail, we applied various stimulus trains. To mimic very slow tonic signaling, we applied stimuli at 0.2 Hz. After the train approached a pseudo steady state, 3 of those pulses are shown applied to the dorsolateral striatum (Fig. 2a) or the NAc shell (Fig. 2b). The pulses at 0.2 Hz (tonic stimulus) are followed by a phasic stimulus (5p at 20 Hz). With this slow tonic stimulus frequency, there was greater DA release evoked by each stimulus in the dorsolateral striatum (Fig. 2a). However, the difference between the tonic and phasic evoked release was greater in the NAc shell (Fig. 2b). That result is reflected in the statistically greater ratio of phasic to tonic evoked DA release,

$[DA]_{5p}/[DA]_{1p}$ , in the NAc shell (Fig. 2c). Other stimulus patterns, however, produced other effects.

The known average background firing frequency of DA neurons is about 4 Hz (Clark and Chiodo, 1988; Grace and Bunney, 1984; Hyland et al., 2002; Zhang et al., 2009). We matched the average firing frequency with a tonic stimulus (applied at 3.3 Hz) followed by a phasic stimulus (5p at 20 Hz). After the tonic train approached a pseudo steady state, 7 of the 3.3 Hz pulses are shown applied to the dorsolateral striatum (Fig. 3a), the NAc core (Fig. 3b), or the NAc shell (Fig. 3c). The pulses at 3.3 Hz (tonic stimuli) were followed by a phasic stimulus (5 p at 20 Hz). With this more biologically relevant tonic stimulus frequency, the response to the tonic stimuli began to blur, giving rise to a background DA level. When the phasic burst was applied, the magnitude of the DA signal increased as the recording was moved from the dorsolateral striatum, to the NAc core, to the NAc shell (Fig. 3d).

The qualitative shape of the DA signal under this biologically relevant stimulus pattern is comparable to that measured with fast-scan cyclic voltammetry applied *in vivo* to a freely-moving rodent (Fig. 3e). Unexpected rewards are known to produce brief burst firing by DA neurons (Schultz et al., 1997). When an unexpected food pellet was given, it caused a greater *in vivo* DA response in the NAc than in the dorsolateral striatum (Fig. 3e), consistent with the *in vitro* data (Fig. 3d). Measured as the area under the curve, the *in vivo* DA response in the NAc,  $0.27 \pm 0.05 \mu\text{M}\cdot\text{s}$ , was significantly larger than that measured from the dorsolateral striatum,  $0.03 \pm 0.01 \mu\text{M}\cdot\text{s}$  ( $n = 10, p < 0.01$ ).

**Cellular Mechanisms involved in regulating Frequency-dependent DA Signals.** The contrast between tonic and phasic DA release within the striatum led us to examine factors that locally regulate those signals. The dorsolateral striatum was stimulated with 1 and 5 pulses at

frequencies ranging from 5 to 80 Hz. In control dorsolateral slices, there was a modest increase in DA release evoked at higher frequencies (Fig. 4a, control). The DA signal peaked at approximately 20 Hz ( $140 \pm 6\%$ ,  $n = 19$ ). Inhibition of the dopamine transporter (DAT) with GBR12909 (2  $\mu\text{M}$ ) enhanced the frequency dependent DA signal especially at the lower frequencies ( $\leq 10$  Hz) (Fig. 4a, GBR, and Fig 4b open diamonds). Inhibition of the DATs shifted the peak of the DA signal to 10 Hz ( $182 \pm 14\%$ ,  $n = 10$ ,  $p < 0.01$ , relative to the DA signal evoked by a single pulse). Inhibition of the dopamine D<sub>2</sub>-type receptors with sulpiride (2  $\mu\text{M}$ ) also enhanced the frequency dependence at lower frequencies (Fig. 4a, sulpiride, and Fig. 4b open triangles), and the DA signal peaked at approximately 5 Hz ( $176 \pm 12\%$ ,  $n = 11$ ,  $p < 0.05$ ).

Nicotinic acetylcholine receptors (nAChRs) also modulated the frequency dependence of DA signaling (Rice and Cragg, 2004; Zhang and Sulzer, 2004; Zhou et al., 2001) in a way that was quite distinct from the modulation caused by DATs or D<sub>2</sub> receptors. Inhibition of  $\beta_2^*$  nAChRs with dihydro- $\beta$ -erythroidine (DH $\beta$ E, 0.1  $\mu\text{M}$ ) decreased DA release (Fig. 4a, DH $\beta$ E), particularly DA release evoked by a signal pulse:  $80 \pm 5\%$  inhibition from control,  $n = 8$ ,  $p < 0.01$ . However, the decrease was inversely related to the stimulus frequency, which resulted (after normalization) in a frequency dependent increase of DA release in DH $\beta$ E-treated slices, particularly above 10 Hz (Fig. 4a, DH $\beta$ E, and Fig. 4c open circles). Simultaneous inhibition of DATs (by GBR), DA D<sub>2</sub>-type receptors (by sulpiride), and  $\beta_2^*$  nAChRs (by DH $\beta$ E) resulted in larger but comparably sized DA signals across all frequencies in the dorsolateral striatum (Fig. 4a bottom and Fig. 4d).

Under our experimental conditions, 1p-evoked DA release was not significantly changed by the inhibition of GABA<sub>A</sub> receptors with bicuculline (data not shown), and muscarinic and glutamatergic receptors were not systematically examined. Taken together, these data suggest

different roles for the DATs, D<sub>2</sub>-type receptors, and nAChRs in modulating tonic and phasic DA release in the dorsolateral striatum.

**Modulation of the DA Signals evoked by Bursts of Varying Length.** The length of phasic bursts (i.e., number of spikes per burst) alters DA release (Floresco et al., 2003; Garris et al., 1994). Although longer bursts are expected to increase DA release throughout the striatum, the increase was much larger in the NAc shell than in the dorsolateral striatum (Fig. 1). We hypothesized that DAT, D<sub>2</sub> receptor, and nAChR activity influence the differences in DA release associated with increased burst length. To test this possibility, bursts of differing length (1 – 10p) were applied at 20 Hz. In the dorsolateral striatum, increasing the burst length enhanced the DA signal only moderately relative to 1p stimulation (control, Fig. 5a, filled squares). However, in the NAc shell the DA signal increased much more as the burst length increased (Fig. 5a, open circles), demonstrating a burst-length dependent facilitation during phasic activity. The DA signal from the NAc core displayed intermediate characteristics, falling between the dorsolateral striatum and NAc shell (*Supplementary Fig. 1*).

When normalized to 1p, bath application of GBR (2 μM) and sulpiride (2 μM) did not alter the relative burst-length dependence of DA release in dorsolateral striatum and NAc shell significantly (Fig. 5b,c). The difference between the dorsolateral striatum and NAc shell remained large (Fig. 5b,c) and comparable to control (Fig. 5a). In contrast, β<sub>2</sub>\* nAChR inhibition by DHβE (0.1 μM) significantly increased the relative DA release evoked by bursts, particularly in the dorsolateral striatum (Fig. 5d). This change made the frequency dependence of the DA signaling of the two regions more similar (Fig. 5d).

The effect of DHβE on the relative DA signal occurred because β<sub>2</sub>\* nAChR inhibition disproportionately decreased tonic DA release evoked by a 1p stimulus as compared to phasic

DA release evoked by a stimulus train. For example, in the dorsolateral striatum, the relative inhibition with DH $\beta$ E was ~80% for a 1p stimulus and ~50% for a stimulus train (n = 8, p < 0.01) (Fig. 6a,b). Furthermore, the inhibitory effect of DH $\beta$ E on tonic DA release was significantly greater in the dorsolateral striatum than in the NAc shell (80% vs. 60%, p < 0.05). That result is represented by showing the DH $\beta$ E inhibition of normalized DA signals to 1p in the dorsolateral striatum and NAc shell (Fig. 6c). In this case, the DA signal to 1p is scaled to be the same size in the dorsolateral striatum and the NAc shell to show that DH $\beta$ E inhibition of nAChRs is less effective in the NAc shell. That finding is also seen when the remain amplitude of the DA signal is shown in a bar graph after DH $\beta$ E inhibition (Fig. 6d).

**DAT, D<sub>2</sub> Receptor, and  $\beta_2^*$  nAChR Modulation of Tonic DA Signals.** We found that low DA release to a single pulse (1p) correlates with stronger burst facilitation (Fig. 1), and we examined how D<sub>2</sub>-type receptors, DATs, and  $\beta_2^*$  nAChRs modulate 1p evoked DA release in the dorsolateral striatum compared to the NAc shell (Fig. 7). Examples of DA release to 1p in the dorsolateral striatum and NAc shell are shown in Fig. 7a. The DA release evoked by 1p ([DA]<sub>1p</sub>) expressed as a ratio of the dorsolateral striatum to the NAc shell indicates greater release to 1p in the dorsolateral striatum (control ratio =  $5.1 \pm 0.63$ , n = 14) (ctrl, Fig. 7a,b). The magnitude of the DA signal, given as area under the curve (Fig. 7c), confirms that the DA signal to 1p is greater in the dorsolateral striatum (ctrl) than the NAc shell (Fig. 7c, NAc, light gray bar).

D<sub>2</sub>-type receptor inhibition (sulpiride, 2  $\mu$ M) increased the 1p-evoked DA release (Fig. 7a, sul) in the dorsolateral striatum ( $131 \pm 11\%$ , n = 11) and the NAc shell ( $122 \pm 6\%$ , n = 7) calculated as area under the curve, without significantly changing the [DA]<sub>1p</sub> ratio between the dorsolateral striatum and the NAc shell (Fig. 7b). Similarly, DAT inhibition (GBR, 2  $\mu$ M) strongly increased the DA signal in the dorsolateral striatum ( $1032 \pm 267\%$ , n = 10) and the NAc

shell ( $762 \pm 153\%$ ,  $n = 8$ ), but did not significantly change the  $[DA]_{1p}$  ratio (Fig. 7b) between the subregions ( $n = 6$ ,  $p > 0.05$ ). In contrast, inhibition of  $\beta_2^*$  nAChRs (DH $\beta$ E,  $0.1 \mu\text{M}$ ) decreased 1p-evoked DA release in both subregions, and the  $[DA]_{1p}$  ratio decreased significantly ( $2.4 \pm 0.49$ ,  $n = 6$ ,  $p < 0.01$  vs. control) (Fig. 7a, b). Similar results were obtained using mutant mice in which the nAChR  $\beta_2$  subunit was knocked out (Fig. 7c, compare bars DH $\beta$ E and  $\beta_2$ ). These results indicate that there are regional differences in  $\beta_2^*$  nAChR modulation of DA release.

We next analyzed the influence of  $D_2$ -type receptors, DATs, and  $\beta_2^*$  nAChRs on the ratio of phasic-to-tonic DA signals. The  $[DA]_{1p}$  was increased by sulpiride or GBR application, but the ratio of DA release evoked by a 5p at 20 Hz versus that evoked by 1p,  $[DA]_{5p}/[DA]_{1p}$ , did not change in the dorsolateral striatum (Fig. 7d). Greater frequency dependent facilitation only occurred when  $[DA]_{1p}$  was reduced by blocking  $\beta_2^*$  nAChR activity (DH $\beta$ E) or using  $\beta_2$ -subunit null mice. The NAc shell (Fig. 7d, NAc, light gray bar) has an intrinsically low  $[DA]_{1p}$  and displays a large control  $[DA]_{5p}/[DA]_{1p}$  ratio. These results indicate that inhibition of nAChRs reduced the DA signal to 1p,  $[DA]_{1p}$ , and switched the pattern of DA release to favor phasic facilitation in the dorsolateral striatum. With nAChRs inhibited, the dorsolateral striatum displays phasic facilitation, represented by  $[DA]_{5p}/[DA]_{1p}$  that is comparable to that seen in the NAc shell (Fig. 7d, compare the last 3 bars)

**Impact of nAChR  $\beta_2$ -subunit Deletion on the Modulation of DA Release.** To confirm the pharmacological effects of  $\beta_2^*$  nAChR inhibition, we evoked DA release in  $\beta_2$ -subunit null mice (Xu et al., 1999). Using the inhibitor, DH $\beta$ E, has the advantage that it is reversible upon washout (Zhou et al., 2001), but the  $\beta_2$ -subunit knockout mouse ensures that there are no residual influences. Evoked DA release (1p) was decreased in the dorsolateral striatum of  $\beta_2^*$  nAChR (-/-) mice ( $0.16 \pm 0.02 \mu\text{M}\cdot\text{s}$ ,  $n = 15$ ) compared to wild-type mice ( $1.03 \pm 0.09 \mu\text{M}\cdot\text{s}$ ,  $n =$

19,  $p < 0.01$ ) and in the NAc shell ( $0.08 \pm 0.02 \mu\text{M}\cdot\text{s}$ ,  $n = 12$ , null;  $0.25 \pm 0.03 \mu\text{M}\cdot\text{s}$ ,  $n = 21$ , wild-type;  $p < 0.01$ ) (Fig. 8a,b). The difference in the  $[\text{DA}]_{1p}$  ratio of the dorsolateral striatum to the NAc shell was also reduced in the null mice (ratio  $2.0 \pm 0.3$ ,  $n = 12$ ) compared to wild-type mice ( $5.1 \pm 0.6$ ,  $n = 14$ ,  $p < 0.01$ ).

The frequency dependent facilitation of DA release was more pronounced in  $\beta_2$ -subunit null mice compared to controls (Fig. 8c,d), where the  $1p$  DA signal is used to normalize the signal to  $5p$  at different intraburst frequencies. In  $\beta_2^*$  nAChR (-/-) mice, burst stimuli elicited greater frequency dependent facilitation in the dorsolateral striatum (Fig. 8a,c). This change made the frequency dependent DA response between the subregions more similar. These results are consistent with the pharmacological effects of  $\beta_2^*$  nAChR inhibition on DA dynamics.

## Discussion

Under control conditions, the NAc shell shows much greater phasic facilitation of DA release to stimulus trains than the dorsolateral striatum (Cragg, 2003; Exley et al., 2008; Thomson, 2000; Zhang et al., 2009). Compared to the DA release evoked by a single pulse, the DA signal arising from a high frequency burst is much greater in the NAc shell than the dorsolateral striatum. The difference in the frequency dependence of DA signaling has been hypothesized to arise from the difference in dopaminergic innervation, with regions that receive ascending fibers from the VTA (A10 cell group) showing more phasic facilitation than those regions predominantly innervated by the SNc (A9 cell group) (Cragg, 2003; Davidson and Stamford, 1993).



Using single pulses or stimulus trains to mimic tonic and phasic DA neuron activity, we found that  $\beta_2^*$  nAChR inhibition disproportionately suppressed tonic DA release compared to phasic release. It should be noted that the nAChRs on DA neurons contain the  $\beta_2$  subunit in various combinations with other subunits, especially  $\alpha_4$  and  $\alpha_6$  (Grady et al., 2007; Salminen et al., 2004). The DA signaling change caused by nAChR inhibition increased the contrast between phasic and tonic DA signals. In that way, nAChR inhibition increased the relative frequency dependence of DA release. Moreover, the effect of  $\beta_2^*$  nAChR inhibition on DA dynamics was more apparent in the dorsolateral striatum than the NAc shell. These pharmacological results were corroborated in nAChR  $\beta_2$ -subunit null mice, which displayed smaller DA signals evoked by single pulses and displayed similar frequency dependent patterns of DA release in the dorsolateral striatum and NAc shell.

We also showed that inhibiting D<sub>2</sub>-type receptors and DATs increased evoked DA release without significantly changing the ratio of phasic to tonic DA responses. Inhibition of D<sub>2</sub>-type receptors and DATs also did not significantly alter the general differences in the DA dynamics between the dorsolateral striatum and NAc shell. Because our direct experimental readout is DA release, we anticipated that the main influence of D<sub>2</sub>-type receptor inhibition with sulpiride was to prevent DA feedback inhibition onto the D<sub>2</sub> autoreceptors on DA terminals. That expectation is consistent with the larger DA release we observed after inhibition (see Figs. 4 and 7). However, sulpiride will inhibit both presynaptic and postsynaptic DA D<sub>2</sub>-type receptors. Because there are differences in function and expression of DATs and D<sub>2</sub> receptors along the dorsal to ventral extent of the striatum (Centonze et al., 2003; Delle Donne et al., 1997), there is potential for varied distributions and roles of D<sub>2</sub> receptors. Despite that potential, the first order influence of sulpiride is likely inhibition of D<sub>2</sub> autoreceptors that regulate DA release.

The present study also demonstrates that  $\beta_2^*$  nAChRs potently regulate the contrast between phasic and tonic DA signals (Zhang et al., 2009), and the results suggest a stronger local functional role of  $\beta_2^*$  nAChRs in the dorsolateral striatum than in the NAc.

**Significance of  $\beta_2^*$  nAChR, DAT, and D<sub>2</sub> Receptor Modulation of Tonic DA Signals.** Much research has shown that the striatum and the midbrain DA neurons that innervate the striatum are important sites of action for nicotine (Dani and Harris, 2005; Grenhoff et al., 1986; Mansvelder et al., 2003; Picciotto and Corrigall, 2002; Pontieri et al., 1996; Rice and Cragg, 2004; Zhang and Sulzer, 2004; Zhang et al., 2009; Zhou et al., 2001). Our findings showed that  $\beta_2^*$  nAChRs have an important role in the modulation of tonic and phasic DA signals within the striatum. We hypothesize that if the probability of DA release is normally elevated in the dorsolateral striatum (via ongoing  $\beta_2^*$  nAChR activity), then higher frequency stimulation cannot boost the release much higher. Thus, the facilitation of DA release caused by phasic stimulation is not normally as apparent in the dorsolateral striatum compared to the NAc shell.

Various nAChR subtypes that contain the  $\beta_2$ -subunit are involved in the modulation of DA release. For example, the  $\alpha_6$ -subunit (in various subunit combinations with  $\beta_2$ ) is an important constituent of the nAChRs with high sensitivity to nicotine that regulates DA release (Exley et al., 2008; Grady et al., 2007; Salminen et al., 2004). Additionally, nicotine inactivates striatal nAChRs through desensitization (Wooltorton et al., 2003), which reduces local DA transmission with a similar overall effect as a nicotinic antagonist. Thus, the relative enhancement of the phasic-to-tonic DA signaling caused by nAChR inhibition could also underlie the type of nicotinic regulation implicated in sensory gating processes or during pauses

in cholinergic activity within the striatum (Cragg, 2006; Zhang and Sulzer, 2004; Zhang et al., 2009).

The DA transporter (DAT) and the DA D<sub>2</sub> autoreceptor both regulate the net extracellular DA concentration and modulate DA transients (Cragg and Rice, 2004). Inhibition of DATs or D<sub>2</sub>-type receptors enhanced DA signals by slowing reuptake and preventing auto-inhibition respectively. Although their inhibition had more potent effects at low frequencies, DAT and D<sub>2</sub> receptor inhibition also has influence over phasic signals. These results are in general agree with previous studies suggesting that the functional efficacy of the DAT and D<sub>2</sub> receptor in regulating DA release declines as the firing frequency increases (Benoit-Marand et al., 2001; Chergui et al., 1994). Although evidence has shown that DAT density is lower in NAc shell than in the dorsal striatum (Coulter et al., 1996), our results demonstrate that inhibition of DATs elicited meaningful changes in evoked DA concentration in both regions.

#### **Significance of the DA Source and the Target-specific Variation in DA Release.**

Microdialysis and voltammetry measurements have shown that the extracellular DA concentration is lower in the NAc than in the dorsal striatum (Cragg, 2003; Fadda et al., 2005; Jones et al., 1995; Keck et al., 2002; Melendez et al., 2003). We found that lower basal DA release (i.e., DA release evoked by 1p) correlated with stronger facilitation of DA release in the NAc shell during phasic stimulation. By contrast in the dorsolateral striatum, which had higher basal DA release, burst-evoked DA release was relatively constant over a wide range of frequencies (5–80 Hz) and burst lengths (1–20 spikes/burst). These results are in agreement with previous studies that demonstrate an inverse relationship between action potential evoked release probability and degree of frequency dependent facilitation (Cragg, 2003; Exley et al., 2008; Thomson, 2000; Zhang et al., 2009).

A larger and rapid transient extracellular DA signal in the NAc shell compared to the dorsolateral striatum indicates greater intrinsic response to phasic midbrain firing activity, which is consistent with the idea that mesolimbic DA neurons encode salient unpredicted sensory events (Schultz et al., 1997). Conversely, the relative stability of dorsolateral striatal DA release in response to changes in firing rate is consistent with the hypothesis that DA release in the dorsal striatum enables the initiation of cognitive and motor responses but does not necessarily correlate with the induction of movement (Romo and Schultz, 1990). These results may explain previous studies showing that dorsal striatal DA release is not linearly dependent on firing activity (Chergui et al., 1994; Montague et al., 2004). High frequency signals cannot boost release linearly because the probability of DA release is initially closer to its maximum value in the dorsal striatum. In contrast, the lower release probability in the ventral striatum allows for the amplification of the DA signal across a wide range of firing frequencies, resulting in a continuing positive dependence of release as the burst length increases (i.e., more spikes per burst).

Based on these findings, it is plausible that the postsynaptic elements in the NAc shell (i.e., glutamatergic afferents and medium spiny neurons) experience a larger contrast between the tonic and phasic DA signals after salient or reward-predictive signals arrive compared to the dorsolateral striatum. Dopamine and glutamate terminals often synapse upon common dendritic spines on striatal neurons (Bouyer et al., 1984). It is theorized that tonic and phasic DA input onto medium spiny neurons could be a part of the mechanism underlying the dopamine-mediated increase in the signal-to-noise ratio of the glutamatergic input signal (Horvitz, 2002; Nicola et al., 2004). Our results demonstrate that the differences in the contrast between tonic and phasic DA release are intrinsic to each striatal subregion, as is the difference in the origin of the DA afferents. The ventral tegmental area (VTA) provides the main dopaminergic innervation to the

NAc shell; whereas the substantia nigra compacta (SNc) provides the main innervation of the dorsolateral striatum. The areas innervated by the VTA, including the medial axis of the striatum and the ventral striatum, show a higher phasic-to-tonic ratio compared to the areas innervated by the SNc, including the dorsolateral striatum. Changes in DA neuron burst firing are particularly important in the ventral striatum where fast glutamatergic processing is required for attention and behavioral selection (Grace et al., 2007). The contrast between the tonic and phasic DA signals provides a key neuronal signal that modulates the postsynaptic targets. Thus, lower tonic DA release in the ventral striatum optimizes the contrast between the phasic and tonic signals.

In summary, these results indicate that there are inherent and local regional differences in the dopaminergic afferents and DA signaling within the striatum (Cragg, 2003; Davidson and Stamford, 1993). We suggest that contrast between phasic and tonic DA signals is a key functional feature that varies within the striatum, and that feature can be endogenously and exogenously modulated by a number of factors. Because neurological and psychiatric disorders such as Parkinson's disease and schizophrenia depend on DA signaling, differences in DA release dynamics and regulation offer an entry target for therapeutic drug development. Separately regulating mesocorticolimbic versus mesostriatal DA signaling could selectively aid schizophrenic and parkinsonian patients respectively. One potential difference between the mesocorticolimbic and mesostriatal DA pathways is the nAChR subtypes that regulate DA release (Janhunen and Ahtee, 2007). By capitalizing on those differences, DA signaling may be therapeutically manipulated to aid different patient populations depending on their DA signaling needs.

## Reference List

- Benoit-Marand M, Borrelli E and Gonon F (2001) Inhibition of dopamine release via presynaptic D2 receptors: time course and functional characteristics in vivo. *J Neurosci* **21**(23):9134-9141.
- Bouyer JJ, Park DH, Joh TH and Pickel VM (1984) Chemical and structural analysis of the relation between cortical inputs and tyrosine hydroxylase-containing terminals in rat neostriatum. *Brain Res* **302**(2):267-275.
- Centonze D, Grande C, Usiello A, Gubellini P, Erbs E, Martin AB, Pisani A, Tognazzi N, Bernardi G, Moratalla R, Borrelli E and Calabresi P (2003) Receptor subtypes involved in the presynaptic and postsynaptic actions of dopamine on striatal interneurons. *J Neurosci* **23**(15):6245-6254.
- Chergui K, Suaud-Chagny MF and Gonon F (1994) Nonlinear relationship between impulse flow, dopamine release and dopamine elimination in the rat brain in vivo. *Neuroscience* **62**(3):641-645.
- Chiodo LA, Bannon MJ, Grace AA, Roth RH and Bunney BS (1984) Evidence for the absence of impulse-regulating somatodendritic and synthesis-modulating nerve terminal autoreceptors on subpopulations of mesocortical dopamine neurons. *Neuroscience* **12**(1):1-16.
- Clark D and Chiodo LA (1988) Electrophysiological and pharmacological characterization of identified nigrostriatal and mesoaccumbens dopamine neurons in the rat. *Synapse* **2**(5):474-485.

- Coulter CL, Happe HK and Murrin LC (1996) Postnatal development of the dopamine transporter: a quantitative autoradiographic study. *Brain Res Dev Brain Res* **92**(2):172-181.
- Cragg SJ (2003) Variable dopamine release probability and short-term plasticity between functional domains of the primate striatum. *J Neurosci* **23**(10):4378-4385.
- Cragg SJ (2006) Meaningful silences: how dopamine listens to the ACh pause. *Trends Neurosci* **29**(3):125-131.
- Cragg SJ and Rice ME (2004) DANCING past the DAT at a DA synapse. *Trends Neurosci* **27**(5):270-277.
- Dani JA and Harris RA (2005) Nicotine addiction and comorbidity with alcohol abuse and mental illness. *Nat Neurosci* **8**(11):1465-1470.
- Davidson C and Stamford JA (1993) Neurochemical evidence of functional A10 dopamine terminals innervating the ventromedial axis of the neostriatum: in vitro voltammetric data in rat brain slices. *Brain Res* **615**(2):229-239.
- Delle Donne KT, Sesack SR and Pickel VM (1997) Ultrastructural immunocytochemical localization of the dopamine D2 receptor within GABAergic neurons of the rat striatum. *Brain Res* **746**(1-2):239-255.
- Di Chiara G, Bassareo V, Fenu S, De Luca MA, Spina L, Cadoni C, Acquas E, Carboni E, Valentini V and Lecca D (2004) Dopamine and drug addiction: the nucleus accumbens shell connection. *Neuropharmacology* **47 Suppl 1**:227-241.
- Exley R, Clements MA, Hartung H, McIntosh JM and Cragg SJ (2008) Alpha6-containing nicotinic acetylcholine receptors dominate the nicotine control of dopamine neurotransmission in nucleus accumbens. *Neuropsychopharmacology* **33**(9):2158-2166.

- Fadda P, Scherma M, Fresu A, Collu M and Fratta W (2005) Dopamine and serotonin release in dorsal striatum and nucleus accumbens is differentially modulated by morphine in DBA/2J and C57BL/6J mice. *Synapse* **56**(1):29-38.
- Floresco SB, West AR, Ash B, Moore H and Grace AA (2003) Afferent modulation of dopamine neuron firing differentially regulates tonic and phasic dopamine transmission. *Nat Neurosci* **6**(9):968-973.
- Gariano RF, Tepper JM, Sawyer SF, Young SJ and Groves PM (1989) Mesocortical dopaminergic neurons. 1. Electrophysiological properties and evidence for somadendritic autoreceptors. *Brain Res Bull* **22**(3):511-516.
- Garris PA, Ciolkowski EL, Pastore P and Wightman RM (1994) Efflux of dopamine from the synaptic cleft in the nucleus accumbens of the rat brain. *J Neurosci* **14**(10):6084-6093.
- Garris PA and Wightman RM (1994) Different kinetics govern dopaminergic transmission in the amygdala, prefrontal cortex, and striatum: an in vivo voltammetric study. *J Neurosci* **14**(1):442-450.
- Gonon FG (1988) Nonlinear relationship between impulse flow and dopamine released by rat midbrain dopaminergic neurons as studied by in vivo electrochemistry. *Neuroscience* **24**(1):19-28.
- Grace AA (1991) Phasic versus tonic dopamine release and the modulation of dopamine system responsivity: a hypothesis for the etiology of schizophrenia. *Neuroscience* **41**(1):1-24.
- Grace AA and Bunney BS (1984) The control of firing pattern in nigral dopamine neurons: single spike firing. *J Neurosci* **4**(11):2866-2876.
- Grace AA, Floresco SB, Goto Y and Lodge DJ (2007) Regulation of firing of dopaminergic neurons and control of goal-directed behaviors. *Trends Neurosci* **30**(5):220-227.



- Grady SR, Salminen O, Laverty DC, Whiteaker P, McIntosh JM, Collins AC and Marks MJ (2007) The subtypes of nicotinic acetylcholine receptors on dopaminergic terminals of mouse striatum. *Biochem Pharmacol* **74**(8):1235-1246.
- Grenhoff J, Aston-Jones G and Svensson TH (1986) Nicotinic effects on the firing pattern of midbrain dopamine neurons. *Acta Physiol Scand* **128**(3):351-358.
- Horvitz JC (2002) Dopamine gating of glutamatergic sensorimotor and incentive motivational input signals to the striatum. *Behav Brain Res* **137**(1-2):65-74.
- Hyland BI, Reynolds JN, Hay J, Perk CG and Miller R (2002) Firing modes of midbrain dopamine cells in the freely moving rat. *Neuroscience* **114**(2):475-492.
- Janhunen S and Ahtee L (2007) Differential nicotinic regulation of the nigrostriatal and mesolimbic dopaminergic pathways: implications for drug development. *Neurosci Biobehav Rev* **31**(3):287-314.
- Jones SR, Garris PA, Kilts CD and Wightman RM (1995) Comparison of dopamine uptake in the basolateral amygdaloid nucleus, caudate-putamen, and nucleus accumbens of the rat. *J Neurochem* **64**(6):2581-2589.
- Keck ME, Welt T, Muller MB, Erhardt A, Ohl F, Toschi N, Holsboer F and Sillaber I (2002) Repetitive transcranial magnetic stimulation increases the release of dopamine in the mesolimbic and mesostriatal system. *Neuropharmacology* **43**(1):101-109.
- Lammel S, Hetzel A, Hackel O, Jones I, Liss B and Roeper J (2008) Unique properties of mesoprefrontal neurons within a dual mesocorticolimbic dopamine system. *Neuron* **57**(5):760-773.

- Mansvelder HD, De Rover M, McGehee DS and Brussaard AB (2003) Cholinergic modulation of dopaminergic reward areas: upstream and downstream targets of nicotine addiction. *Eur J Pharmacol* **480**(1-3):117-123.
- Melendez RI, Rodd-Henricks ZA, McBride WJ and Murphy JM (2003) Alcohol stimulates the release of dopamine in the ventral pallidum but not in the globus pallidus: a dual-probe microdialysis study. *Neuropsychopharmacology* **28**(5):939-946.
- Montague PR, McClure SM, Baldwin PR, Phillips PE, Budygin EA, Stuber GD, Kilpatrick MR and Wightman RM (2004) Dynamic gain control of dopamine delivery in freely moving animals. *J Neurosci* **24**(7):1754-1759.
- Nicola SM, Woodward Hopf F and Hjelmstad GO (2004) Contrast enhancement: a physiological effect of striatal dopamine? *Cell Tissue Res* **318**(1):93-106.
- Nisell M, Marcus M, Nomikos GG and Svensson TH (1997) Differential effects of acute and chronic nicotine on dopamine output in the core and shell of the rat nucleus accumbens. *J Neural Transm* **104**(1):1-10.
- Phillips PE, Robinson DL, Stuber GD, Carelli RM and Wightman RM (2003) Real-time measurements of phasic changes in extracellular dopamine concentration in freely moving rats by fast-scan cyclic voltammetry. *Methods Mol Med* **79**:443-464.
- Picciotto MR and Corrigall WA (2002) Neuronal systems underlying behaviors related to nicotine addiction: neural circuits and molecular genetics. *J Neurosci* **22**(9):3338-3341.
- Pontieri FE, Tanda G, Orzi F and Di Chiara G (1996) Effects of nicotine on the nucleus accumbens and similarity to those of addictive drugs. *Nature* **382**(6588):255-257.
- Rice ME and Cragg SJ (2004) Nicotine amplifies reward-related dopamine signals in striatum. *Nat Neurosci* **7**(6):583-584.

- Robinson S, Smith DM, Mizumori SJ and Palmiter RD (2004) Firing properties of dopamine neurons in freely moving dopamine-deficient mice: effects of dopamine receptor activation and anesthesia. *Proc Natl Acad Sci U S A* **101**(36):13329-13334.
- Romo R and Schultz W (1990) Dopamine neurons of the monkey midbrain: contingencies of responses to active touch during self-initiated arm movements. *J Neurophysiol* **63**(3):592-606.
- Salminen O, Murphy KL, McIntosh JM, Drago J, Marks MJ, Collins AC and Grady SR (2004) Subunit composition and pharmacology of two classes of striatal presynaptic nicotinic acetylcholine receptors mediating dopamine release in mice. *Mol Pharmacol* **65**(6):1526-1535.
- Schultz W (1986) Responses of midbrain dopamine neurons to behavioral trigger stimuli in the monkey. *J Neurophysiol* **56**(5):1439-1461.
- Schultz W (2007) Multiple dopamine functions at different time courses. *Annu Rev Neurosci* **30**:259-288.
- Schultz W, Dayan P and Montague PR (1997) A neural substrate of prediction and reward. *Science* **275**(5306):1593-1599.
- Shi WX, Pun CL, Zhang XX, Jones MD and Bunney BS (2000) Dual effects of D-amphetamine on dopamine neurons mediated by dopamine and nondopamine receptors. *J Neurosci* **20**(9):3504-3511.
- Thomson AM (2000) Facilitation, augmentation and potentiation at central synapses. *Trends Neurosci* **23**(7):305-312.

- Wooltorton JR, Pidoplichko VI, Broide RS and Dani JA (2003) Differential desensitization and distribution of nicotinic acetylcholine receptor subtypes in midbrain dopamine areas. *J Neurosci* **23**(8):3176-3185.
- Wu Q, Reith ME, Walker QD, Kuhn CM, Carroll FI and Garris PA (2002) Concurrent autoreceptor-mediated control of dopamine release and uptake during neurotransmission: an in vivo voltammetric study. *J Neurosci* **22**(14):6272-6281.
- Xu W, Orr-Urtreger A, Nigro F, Gelber S, Sutcliffe CB, Armstrong D, Patrick JW, Role LW, Beaudet AL and De Biasi M (1999) Multiorgan autonomic dysfunction in mice lacking the beta2 and the beta4 subunits of neuronal nicotinic acetylcholine receptors. *J Neurosci* **19**(21):9298-9305.
- Zhang H and Sulzer D (2004) Frequency-dependent modulation of dopamine release by nicotine. *Nat Neurosci* **7**(6):581-582.
- Zhang T, Zhang L, Liang Y, Siapas AG, Zhou FM and Dani JA (2009) Dopamine signaling differences in the nucleus accumbens and dorsal striatum exploited by nicotine. *J Neurosci* **29**(13):4035-4043.
- Zhou FM, Liang Y and Dani JA (2001) Endogenous nicotinic cholinergic activity regulates dopamine release in the striatum. *Nat Neurosci* **4**(12):1224-1229.

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## Figure legends

**Fig. 1.** DA signals elicited by a signal pulse (1p) or by 5p or 20p given at 20 Hz. (a) Example traces of DA signals measured using fast-scan cyclic voltammetry in the dorsolateral striatum. Below, a comparison is given of the DA signal evoked by 1p (dotted line) and by a 20p train at 20 Hz (solid gray line). The 2 sets of different scale bars represent 0.5  $\mu$ M DA and 2 s. (b) Example traces of DA signals measured in the nucleus accumbens (NAc) core. (c) Example traces of DA signals measured in the NAc shell. (d) The average DA signal evoked by 1p, calculated as the area under the curve, is displayed for the 3 regions ( $n = 7-16$ , \*\*  $p < 0.01$ , compared to the dorsolateral striatum, DS), where DS = dorsolateral striatum. (e) The average phasic/tonic ratio of the DA signal evoked by 20p over 1p ( $[DA]_{20p} / [DA]_{1p}$ ) is displayed for the 3 regions ( $n = 7-16$ , \*\*  $p < 0.01$ , compared to DS).

**Fig. 2.** The ratio of the phasic to slow tonic DA signals within the dorsolateral striatum and the NAc shell. (a) An example DA trace is shown in response to a tonic stimulation (3 pulses at 0.2 Hz after achieving pseudo steady state) followed by a phasic stimulation (5p at 20 Hz) within the dorsolateral striatum. (b) An example DA trace is shown in response to the same tonic and phasic stimulation within the NAc shell. Scale bars represent 0.2  $\mu$ M DA, 5s. (c) The average ratio of the phasic to tonic DA signal ( $[DA]_{5p} / [DA]_{1p}$ ) showed a greater contrast in the NAc shell compared to the dorsolateral striatum ( $n = 4$ , \*\*  $p < 0.01$ ).

**Fig. 3.** To mimic biologically realistic tonic and phasic DA neuron firing activity, we applied stimulus trains of 3.3 Hz (tonic) and 20 Hz (phasic). (a) An example DA trace is shown in

response to a tonic stimulation (7 pulses at 3.3 Hz after achieving pseudo steady state) followed by a phasic stimulation (5p at 20 Hz) within the dorsolateral striatum. (b) An example DA trace is shown in response to the same tonic and phasic stimulation within the NAc core and (c) within the NAc shell. (d) DA signal (area under the curve) evoked by the phasic train was significantly larger in NAc shell and core than in the dorsolateral striatum ( $n = 11,12,10$ ;  $* p < 0.05$ ). (e) *In vivo* fast-scan cyclic voltammetry showed that an unexpected food pellet elicited a DA signal in the dorsolateral striatum that was smaller than that in the NAc ( $n = 10$ ,  $p < 0.01$ ), consistent with the *in vitro* slice data above.

**Fig. 4.** The frequency dependence of DA signals in the dorsolateral striatum in response to inhibition of DATs, dopamine D<sub>2</sub>-type receptors, and  $\beta_2^*$  nAChRs. (a) Example DA traces evoked by 1p or by 5p trains at stimulus frequencies of 5, 20, or 80 Hz. The DA signal was evoked under the following conditions: no inhibition (control), inhibition of DATs with GBR12909 (GBR, 2  $\mu$ M), inhibition of D<sub>2</sub>-type receptors with sulpiride (2  $\mu$ M), inhibition of nAChRs with DH $\beta$ E (0.1  $\mu$ M), and in the presence of all 3 antagonists (GBR+sul+DH $\beta$ E). Scale bars represent 1  $\mu$ M DA and 5s, except in GBR where the y-scale represents 2  $\mu$ M DA. (b) GBR (open diamonds) and sulpiride (open triangles) enhanced the relative DA signal (normalized to the DA signal evoked by 1p) at low frequencies compared to the control (filled squares). (c) DH $\beta$ E (open circles) elicited robust facilitation of the DA signal at higher frequencies compared to control. (d) Combining GBR, sulpiride, and DH $\beta$ E led to an increased DA signal that was comparable in size across stimulus frequencies using either 3p (open squares) or 5p (crossed squares) compared to control ( $n = 4-12$ ).

**Fig. 5.** The dorsolateral striatum and the NAc shell respond differently to stimulus trains of 5p or 10p given at 20 Hz. Comparisons of the DA signals evoked by phasic stimulus trains were obtained in the absence (control) or presence of DAT, D<sub>2</sub> receptor, and  $\beta_2^*$  nAChR antagonists. The DA signals were normalized to the response evoked by 1p stimulation under each experimental condition. (a) In the absence of antagonists (control), the DA signal in the NAc shell was more responsive to phasic stimulus trains. (b) With DATs inhibited by GBR (2  $\mu$ M, n = 6), the relationship of the 2 regions remained similar to control. (c) With D<sub>2</sub>-type receptors inhibited by sulpiride (2  $\mu$ M, n = 6), the relationship of the 2 regions remained similar to control. (d) With nAChRs inhibited by DH $\beta$ E (0.1  $\mu$ M, n = 6), the frequency dependence of both regions increased, but the increase was greater in the dorsolateral striatum. Consequently, the frequency dependence of the 2 regions became more similar after nAChR inhibition.

**Fig. 6.** Inhibition of  $\beta_2^*$  nAChRs with DH $\beta$ E (0.1  $\mu$ M) altered DA signals (a) Example DA signals evoked by 1p and by a 5p train given at 20 Hz within the dorsolateral striatum in the absence (control) or presence of DH $\beta$ E. Scale bars represent 0.5  $\mu$ M DA and 0.5 s. (b) The average DA signal evoked by 1p and 5p at 20 Hz in the presence of DH $\beta$ H normalized to the control response as 100%. (c) The DA signal evoked by 1p in the dorsolateral striatum and NAc shell in the absence (control) or presence of DH $\beta$ E. The DA signals are normalized to 1p using different y-scale bars: 0.5  $\mu$ M DA for the dorsolateral striatum and 0.1  $\mu$ M DA for the NAc shell and 0.5 s. (d) The average DA signal evoked by 1p in the dorsolateral striatum and NAc shell in the presence of DH $\beta$ H normalized to the control response as 100%.



**Fig. 7.** DA signals evoked by 1p in the absence or presence of antagonists for DATs, D<sub>2</sub>-type receptors, or  $\beta_2^*$  nAChRs. (a) Representative DA signals evoked by 1p from the dorsolateral striatum (gray traces) and the NAc shell (dash traces). The DA signals were controls (ctrl) or with DATs (GBR, 2  $\mu$ M), D<sub>2</sub>-type receptors (sulpiride, 2  $\mu$ M), or  $\beta_2^*$  nAChRs (DH $\beta$ H, 0.1  $\mu$ M) inhibited. Scale bars represent 1  $\mu$ M DA and 2 s. (b) The ratio of the DA signal evoked by 1p in the dorsolateral striatum over that in the NAc shell with and without antagonists as labeled (n = 5–11). (c) The average magnitude of DA signal evoked by 1p in the dorsolateral striatum with and without antagonists as labeled (n = 6–19). The bar labeled “ $\beta_2$ ” represents data from mutant mice lacking the nAChR  $\beta_2$  subunit. The control response in the NAc shell is shown for comparison (last bar, light gray). (d) The average ratio of the DA signal evoked by 5 p at 20 Hz over that evoked by 1p,  $[DA]_{5p}/[DA]_{1p}$ , in the dorsolateral striatum with and without antagonists as labeled (n = 6–19). The control response in the NAc shell is shown for comparison (last bar, light gray). Significance is given as \* p < 0.05, \*\* p < 0.01 compared to ctrl.

**Fig. 8.** The frequency dependence of DA signals from wild-type and nAChR  $\beta_2$ -subunit null mice. (a) Representative DA signals evoked by 1p and 5p at 20 Hz from wild-type littermates ( $\beta_2$  +/+) and mutant null mice ( $\beta_2$  -/-) in the dorsolateral striatum. (b) Representative DA signals from wild-type ( $\beta_2$  +/+) and null mice ( $\beta_2$  -/-) in the NAc shell. (c) Normalized to the DA signal evoked by 1p in the dorsolateral striatum, the average DA signal is shown for 5p delivered at frequencies ranging from 10 to 80 Hz in null mice ( $\beta_2$  -/-, open squares) and wild-type mice ( $\beta_2$  +/+, filled squares) (n = 11,15 ). (d) Normalized to the DA signal evoked by 1p in the NAc shell, the average DA signal is shown for 5p at 10 to 80 Hz in null mice ( $\beta_2$  -/-, open circles) and wild-type mice ( $\beta_2$  +/+, filled circles) (n = 11,12 ).

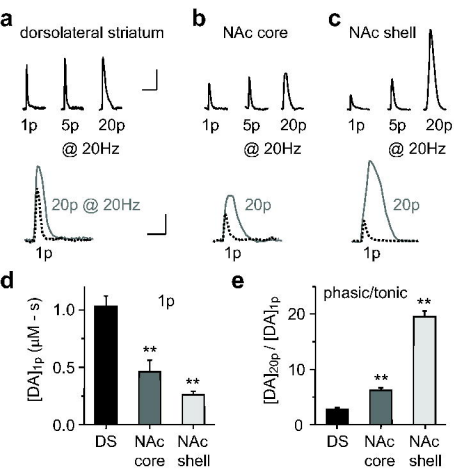


Fig. 1

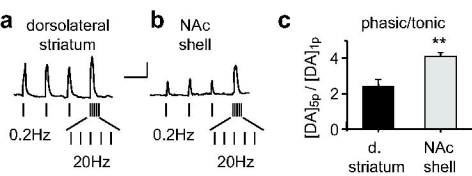


Fig. 2

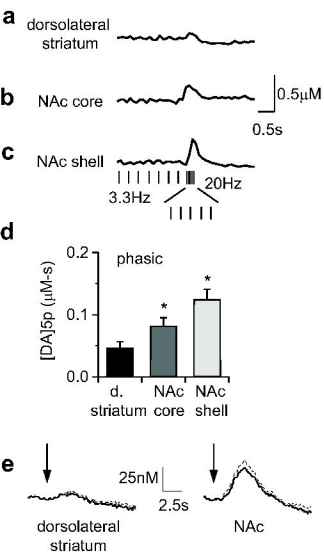


Fig. 3

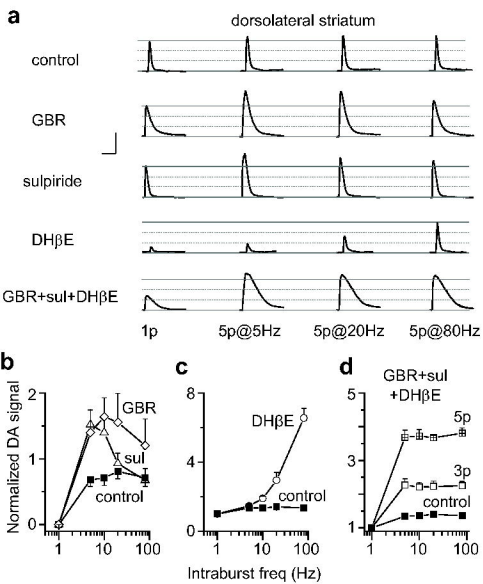


Fig. 4

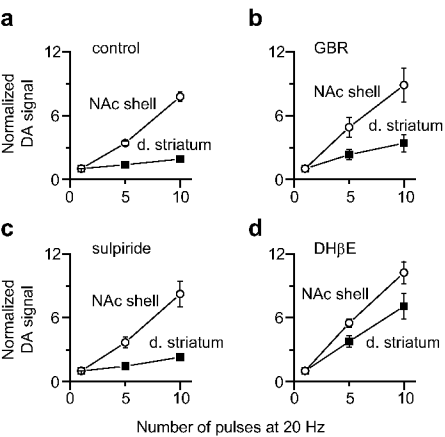


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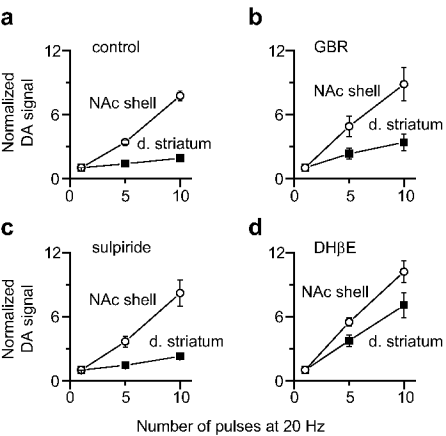


Fig. 6

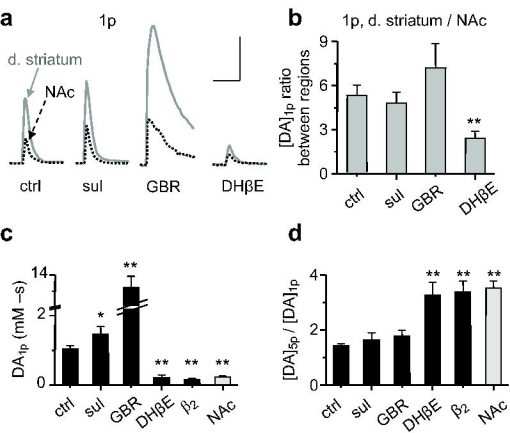


Fig. 7



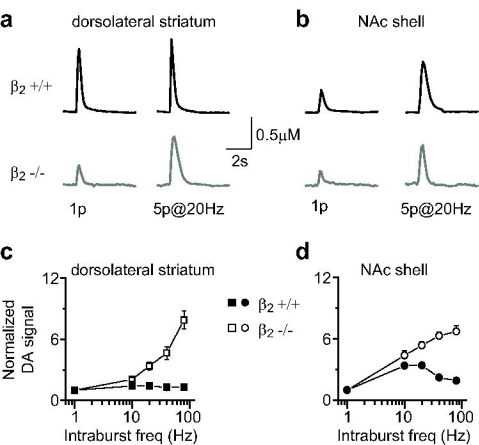


Fig. 8