SITE-DIRECTED MUTAGENESIS OF ALDEHYDE DEHYDROGENASE-2 SUGGESTS THREE DISTINCT PATHWAYS OF NITROGLYCERIN BIOTRANSFORMATION

M. Verena Wenzl, Matteo Beretta, Martina Griesberger, Michael Russwurm, Doris Koesling, Kurt Schmidt, Bernd Mayer, and Antonius C.F. Gorren

Department of Pharmacology and Toxicology, Karl-Franzens-Universität Graz, A-8010 Graz, Austria (M.V.W., M.B., M.G., K.S., B.M., A.C.F.G.)

Department of Pharmacology and Toxicology, Ruhr-Universität Bochum, D-44780 Bochum, Germany (M.R., D.K.)
Running Title: Three Pathways of GTN Biotransformation by ALDH-2

Address correspondence to:
A.C.F. Gorren
Dept. of Pharmacology & Toxicology
Karl-Franzens-Universität Graz
Universitätsplatz 2, A-8010, Austria
Tel.: +43-316-380 5569
Fax: +43-316-380 9890
e-mail: antonius.gorren@uni-graz.at

Number of pages: 30 (excluding schemes and figures)
Number of tables: 2
Number of figures: 5 (+ 2 schemes)
Number of references: 25
Number of words in Abstract: 247
Number of words in Introduction: 619
Number of words in Discussion: 1484

Abbreviations: ALDH2, mitochondrial aldehyde dehydrogenase (EC 1.2.1.3); DEA/NO, 2,2-diethyl-1-nitroso-oxyhydrazine; DTPA, diethylene triamine pentaacetic acid; GDN, glyceryl dinitrate; GTN, glyceryl trinitrate (nitroglycerin); metHb, methemoglobin; oxyHb, oxyhemoglobin; sGC, soluble guanylate cyclase; SOD, superoxide dismutase; TEA, triethanolamine.
ABSTRACT

To elucidate the mechanism underlying reduction of nitroglycerin (GTN) to nitric oxide (NO) by mitochondrial aldehyde dehydrogenase (ALDH2), we generated mutants of the enzyme lacking the cysteines adjacent to reactive Cys-302 (C301S and C303S), the glutamate that participates as a general base in aldehyde oxidation (E268Q), or combinations of these residues. The mutants were characterized regarding acetaldehyde dehydrogenation, GTN-triggered enzyme inactivation, GTN denitration, NO formation, and soluble guanylate cyclase activation. Lack of the cysteines did not affect dehydrogenase activity, but impeded GTN denitration, aggravated GTN-induced enzyme inactivation, and increased NO formation. A triple mutant lacking the cysteines and Glu-268 catalyzed sustained formation of superstoichiometric amounts of NO and exhibited slower rates of inactivation. These results suggest three alternative pathways for the reaction of ALDH2 with GTN, all involving formation of a thionitrate/sulfenyl nitrite intermediate at Cys-302 as the initial step. In the first pathway, which predominates in the wild-type enzyme and reflects clearance-based GTN denitration, the thionitrate apparently reacts with one of the adjacent cysteine residues to yield nitrite and a protein disulfide. The predominant reaction catalyzed by the single and double cysteine mutants requires Glu-268 and results in irreversible enzyme inactivation. Finally, combined lack of the cysteines and Glu-268 shifts the reaction towards formation of free NO radical, presumably through homolytic cleavage of the sulfenyl nitrite intermediate. Though the latter reaction accounts for less than 10% of total turnover of GTN metabolism catalyzed by wild-type ALDH2, it is most likely essential for vascular GTN bioactivation.
INTRODUCTION

Nitroglycerin (glyceryl trinitrate, GTN) has been used for the treatment of angina pectoris since the late nineteenth century. Bioactivation of GTN in vascular smooth muscle is thought to yield NO that activates soluble guanylate cyclase (sGC), resulting in cGMP-mediated vasorelaxation (Fung, 2004). Although GTN activates sGC non-enzymatically in the presence of cysteine (Gorren et al., 2005) or high concentrations of ascorbate (Kollau et al., 2007) via NO release, there is a large body of evidence (Mayer and Beretta, 2008) that vascular bioactivation of GTN involves an enzymatic reaction catalyzed by mitochondrial aldehyde dehydrogenase (ALDH2). In 2002, Stamler and co-workers showed that ALDH2 denitrates GTN to 1,2-glyceryl dinitrate (1,2-GDN) and nitrite and proposed that reduction of nitrite by components of the mitochondrial electron transfer chain could provide the link between ALDH2-catalyzed GTN metabolism and vasorelaxation (Chen et al., 2002). However, we found that (i) GTN activates sGC in the presence of purified ALDH2 (Kollau et al., 2005), (ii) ALDH2 catalyzes the reduction of GTN to NO, accounting for 5-10% of total GTN turnover (Beretta et al., 2008a; Wenzl et al., 2009), and (iii) mitochondrial biotransformation of nitroglycerin is not affected by respiratory substrates or inhibitors (Kollau et al., 2009), suggesting that ALDH2 catalysis is sufficient for GTN bioactivation in blood vessels.

The mechanism underlying ALDH2-catalyzed GTN reduction is not known, but the reaction appears to occur within the active site pocket with Cys-302 serving as the common essential nucleophile (Larson et al., 2007; Wenzl et al., 2009). It was proposed by Stamler and co-workers (Chen et al., 2002) that GTN denitration resembles ester hydrolysis by ALDH2 (Mann and Weiner, 1999), i.e. nucleophilic attack of a nitro group of GTN by Cys-302, resulting in formation of a thionitrate intermediate and release of the corresponding alcohol, preferentially 1,2-GDN. The thionitrate would then release nitrite either through nucleophilic attack of one of the adjacent cysteine residues (Cys-301 or Cys-303), resulting
in formation of a disulfide in the active site, or through Glu-268-aided hydrolysis, yielding a
sulfenic acid derivate of Cys-302 that could undergo S-thiolation (Biswas et al., 2006) and
would also lead to the formation of a disulfide with one of the adjacent cysteine residues.
Both possibilities would agree well with the fact that NAD\(^+\) is not essential but increases
reaction rates and with the observation that GTN-triggered oxidative inactivation of ALDH2
is partially prevented by thiols like dithiothreitol (DTT) or dihydrolipoic acid (Wenzel et al.,
2007; Beretta et al., 2008b). Interestingly, we found that mutation of Glu-268 resulted in
unaltered GTN denitration by ALDH2 in the absence of NAD\(^+\), whereas the nucleotide
cofactor led to pronounced inhibition of 1,2-GDN formation by E268Q-ALDH2, probably
due to involvement of Glu-268 in the structural organization of the NAD\(^+\) binding pocket.
Another interesting feature of E268Q-ALDH2 was that fractional NO formation from GTN
was increased to up to 50\% of total GTN turnover (Wenzl et al., 2009), pointing to an
independent three electron reduction pathway of GTN denitration in addition to the two
electron reduction pathway yielding nitrite that is predominant in the wild-type enzyme.

In the present study we investigated the role of the cysteine residues adjacent to the
catalytically active Cys-302 in GTN bioactivation as well as mechanism-based oxidative
enzyme inactivation, which represents an attractive explanation for the phenomenon of
nitrate tolerance because long-term exposure of blood vessels to GTN causes inactivation of
ALDH2 (Towell et al., 1985; Daiber et al., 2004; Chen et al., 2007). Besides single and
double mutants of Cys-301 and Cys-303 (C301S-ALDH2, C303S-ALDH2, and
C301S/C303S-ALDH2, hereafter referred to as the ‘double mutant’), we additionally tested
E268Q/C301S/C303S-ALDH2 (hereafter referred to as the ‘triple mutant’), in order to
exclude the possibility that Glu-268 substituted for either of the cysteines in the reaction.
MATERIALS & METHODS

Materials

Bovine lung sGC was purified as described (Russwurm and Koesling, 2005). Human ALDH2 was expressed in Escherichia coli BL21(DE3) and purified as described (Zheng et al., 1993; Beretta et al., 2008a). Concentrations are expressed per monomer assuming a molecular weight of 54 kDa. Sephacryl S-300 HR was obtained from GE Healthcare Europe GmbH (Vienna, Austria). [α-32P]GTP (400 Ci/mmol) was from PerkinElmer Life and Analytical Sciences (Vienna, Austria). [2-14C]GTN (50-60 mCi/mmol) was from American Radiolabeled Compounds, purchased through Humos Diagnostica GmbH (Maria Enzersdorf, Austria). Nitropohl® ampoules (G. Pohl-Boskamp GmbH. & Co, Hohenlockstedt Germany), containing 4.4 mM GTN in 250 mM glucose, were obtained from a local pharmacy; dilutions were made in 50 mM triethanolamine(TEA)/HCl buffer. 2,2-Diethyl-1-nitroso-oxyhydrzone (DEA/NO; Enzo Life Sciences Corporation, Lausen, Switzerland) was dissolved and diluted in 10 mM NaOH. All other chemicals were from Sigma-Aldrich GmbH (Vienna, Austria).

Site-directed Mutagenesis

E268Q, C301S, C303S, C301S/C303S and E268Q/C301S/C303S mutations were inserted using the QuickChange® II site-directed mutagenesis kit (Stratagene). The following mutagenic sense primers were used to introduce the mutation (bold type) and to add the BamHI restriction site (underlined) as a silent mutation for screening of mutants to the cDNA encoding for WT-ALDH2: 5'-GTT CTT CAA CCA GGG CCA GTC CTG CTG TGC CGG aTC CCG GAC CTT CGT G-3’ for C301S, 5’-GTT CTT CAA CCA GGG CCA GTG CTG CTC TGC CGG aTC CCG GAC CTT CGT G-3’ for C303S, and 5’-GTT CTT CAA CCA GGG CCA GTC CTG CTC TGC CGG aTC CCG GAC CTT CGT G-3’ for C301S/C303S.

The E268Q mutation was introduced to the cDNA encoding for WT-ALDH2 as described
(Wenzl et al., 2009). For the E268Q/C301S/C303S mutation the mutagenic sense primer 5'-GCA GCA ACC TtA AGA GAG TGA CCT TGC AGC TGG GGG GGA AG-3' was used to introduce the E268Q mutation (bold type) and to add the AflII restriction site (underlined) as a silent mutation for screening of mutants to the cDNA encoding for C301S/C303S-ALDH2. In order to confirm the desired mutations, sequencing of cDNAs was performed with an ABI 373A automated DNA sequencer (Applied Biosystems, Life Technologies Corporation, Carlsbad, California, USA). All mutants were expressed in *Escherichia coli* BL21(DE3) and purified as described (Zheng et al, 1993; Beretta et al., 2008a).

*Determination of GTN Denitrification*

The rates of GTN metabolism yielding 1,2- and 1,3-GDN were determined according to a described protocol (Kollau et al., 2005). WT and mutated ALDH2 proteins (4 µg each) were incubated with 14C-labeled GTN (≥ 2 µM as indicated) at 37°C for 10 min in a final volume of 0.2 mL 50 mM potassium phosphate (K-phosphate) buffer (pH 7.4), containing 3 mM MgCl₂ and 1 mM NAD⁺ in the absence and presence of 2 mM dithiothreitol (DTT). GTN concentrations ≥ 10 µM were adjusted by addition of unlabelled GTN. Reaction products were extracted twice with 1 mL diethyl ether, separated by thin layer chromatography, and quantified by liquid scintillation counting. Blank values were determined in the absence of protein under identical conditions and subtracted. The results are mean values ± S.E. determined in 3-5 independent experiments. Since 1,3-GDN yields in the concentration range investigated in the present study are all but negligible (Beretta et al., 2010), only the 1,2-GDN yields are presented.

*Determination of GTN-derived NO*

NO formation was measured with a Clark-type electrode (WPI Inc., Berlin, Germany) that was calibrated daily with acidified nitrite as described previously (Mayer et al., 1995).
The electrode was equilibrated in 50 mM TEA/HCl (pH 7.4), containing 1000 U/mL superoxide dismutase (SOD), 0.1 mM diethylene triamine pentaacetic acid (DTPA), 1 mM NAD+, 3 mM MgCl₂, and purified WT or mutated ALDH2 (250 µg/mL) in the absence or presence of 2 mM DTT at 37°C, followed by addition of GTN (final concentration of 0.1 mM). The incubation volume was 0.5 mL. Output current was recorded in 3-6 separate experiments.

**Oxyhemoglobin Assay**

Conversion of oxyhemoglobin (oxyHb) to methemoglobin (metHb) by NO was measured by monitoring the absorbance difference between 420 and 401 nm, which represent the peak and trough in the oxyHb-minus-metHb Soret absorption difference spectrum. OxyHb was prepared from bovine hemoglobin by reduction with a molar excess of sodium dithionite and purification by gel chromatography (Sephadex G25). ALDH2 (250 µg/mL) was incubated in 0.25 mL K-phosphate (50 mM, pH 7.4) containing 1000 U/mL SOD, 10 U/mL catalase, 0.1 mM DTPA, 1 mM NAD+, 3 mM MgCl₂ and 10 µM oxyHb in the absence or presence of 2 mM DTT at 37°C, followed by addition of GTN (0.1 mM). NO concentrations were calculated with an extinction coefficient (ΔΔε₄₂₀₋₄₀₁ = 91.9 mM⁻¹ cm⁻¹) that was determined with K₃Fe(CN)₆ in 3-5 separate experiments. Control experiments in the absence of ALDH2 and DTT showed direct oxidation of oxyHb by GTN. This artifactual reaction, which did not occur in the presence of DTT, was subtracted from all affected data (essentially the slow phase in the absence of DTT).

**Determination of sGC Activity**

The sGC preparation used for the experiments in the present study exhibited a maximal specific activity of ~20 µmol cGMP x min⁻¹ x mg⁻¹ (tested with 10 µM DEA/NO)
and contained 2 mM DTT to protect the enzyme from oxidation, yielding a final DTT concentration of 10 µM in the assays. Purified sGC (50 ng) was incubated at 37°C for 2 min in 100 µL with GTN in the presence of WT or mutant ALDH2 (2 µg each) as indicated in the text and figure legends. Assay mixtures contained 50 mM TEA/HCl (pH 7.4), 0.5 mM [α-32P]GTP (~250,000 cpm), 3 mM MgCl2, and 1 mM cGMP. NAD+ (1 mM) and SOD (1000 U/mL) were present as indicated. Reactions were terminated by addition of 0.45 mL zinc acetate (120 mM) and 0.45 mL sodium bicarbonate (120 mM), followed by isolation and quantification of 32P-cGMP (Schultz and Böhme, 1984). Blank values were determined in the absence of sGC and subtracted. The results are mean values ± S.E. determined in 3-5 independent experiments.

ALDH2 Inactivation by GTN

GTN-triggered inactivation of ALDH2 was determined as described (Beretta et al., 2008b). Dehydrogenase activity of WT or mutant ALDH2 (33 µg/mL each) was measured by monitoring the formation of NADH spectrophotometrically at 340 nm (ε340 = 6.22 mM–1 cm–1) at 25°C. The initial reaction mixture consisted of 0.4 mM acetaldehyde, 0.4 mM NAD+ and 10 mM MgCl2 in 50 mM K-phosphate (pH 7.4). After 2 min reactions were started by addition of ALDH2. About 4 min later 50 µM GTN was added to inactivate the enzyme. After complete inactivation 0.4 mM DTT was added to reactivate the enzyme. The linear absorbance increases after addition of ALDH2 and DTT yielded initial (v0) and restored (vrestored) activities, respectively. The gradual inactivation after GTN addition was fitted to a single-exponential curve yielding the apparent inactivation rate constant (kinact). The results are mean values ± S.E. determined in 4-7 independent experiments.
RESULTS

GTN Denitration

As shown in Fig. 1A, GTN was denitrated within 10 min to ~1 mol of 1,2-GDN per mol of ALDH2 monomer in the absence of DTT by WT-ALDH2 as well as by C301S-ALDH2, C303S-ALDH2, and the double mutant, indicating that these enzyme species are inactivated by GTN after one turnover, as reported earlier for WT-ALDH2 (Chen and Stamler, 2006; Beretta et al., 2008b). We previously found that E268Q-ALDH2 exhibits strongly reduced activities, particularly in the presence of NAD+ (Wenzl et al., 2009). In line with those observations E268Q-ALDH2 was virtually inactive, while the triple mutant formed 4-fold less 1,2-GDN than the double mutant (Fig. 1A).

Figure 1B shows that WT-ALDH2 catalyzed maximally ~14 turnovers in the presence of DTT, while mutation of Cys-301 or Cys-303, alone or in combination, decreased the yield to ~3 turnovers. Because NAD+ was present in the assay, GTN denitration by E268Q-ALDH2 was not detectable at any GTN concentration tested (Wenzl et al., 2009). Surprisingly, GTN denitration by the triple mutant was more pronounced, with a maximum of ~2 turnovers at ≥ 30 µM GTN. The concentration-response curves suggest that the triple mutant exhibits low affinity for GTN (EC50~20 µM) as compared to WT-ALDH (EC50~3 µM). The lack of effect of increasing GTN concentrations on denitration catalyzed by the three cysteine mutants most likely reflects complete inactivation of the enzyme within the 10 min incubation period (see below).

NO Formation

There was a striking discrepancy between GTN denitration and formation of NO catalyzed by the different ALDH2 variants. In the absence of DTT, all cysteine mutants (C301S and C303S as well as double and triple mutants) yielded NO peak concentrations that
were between 2.5- and 4.4-fold higher than observed with the wild-type enzyme (Fig. 2 & Table 1). Initial rates of NO formation were between 2.8- and 7.2-fold higher as compared to wild-type. Thus, mutation of these residues apparently caused a shift of GTN turnover towards formation of NO. A similar though less pronounced shift was observed in the presence of DTT, which increased the peak concentration of NO formed by WT-ALDH2 8-fold but caused only between 2- and 4-fold increases of NO peak heights for the mutants. As a result, in the presence of DTT, incubation of GTN with C301S-ALDH2 and wild-type enzyme yielded the same NO peak heights, while the other cysteine mutants showed between 1.3- and 1.5-fold higher NO peaks. All four mutants catalyzed NO formation with between 1.4- and 2.9-fold higher initial rates than WT-ALDH2 (Table 1 & Suppl. Fig. S1).

Considering that the wild-type enzyme denitrated approx. 7 times more GTN than the mutants (Fig. 1B), these data indicate that the fraction of GTN reduced to NO was increased upon mutation of Cys-301, Cys-303 and/or Glu-268.

The mutant enzymes also differed from WT-ALDH2 with respect to the decay rates of the NO signal, particularly in the presence of DTT (Fig. 2 & Table 1). C303S-ALDH2 and the double mutant exhibited approx. 2-fold higher decay rates than the wild-type enzyme, whereas the decay of the signal observed in the presence of the triple mutant was slower. Since the observed rate of NO decay correlates with the rate of ALDH2 inactivation (Beretta et al., 2010), these data suggest that ALDH2 inactivation is accelerated upon mutation of one or both cysteines, whereas mutating Glu-268 protects against inactivation. The latter observation agrees with previous results obtained with the single E268Q mutant (Wenzl et al., 2009).

All experiments described above were performed in the presence of SOD. We have previously shown that the presence of SOD is essential for the electrochemical detection of NO with WT-ALDH2, but not with E268Q-ALDH2 (Wenzl et al., 2009). Similarly, when we omitted SOD from the reaction mixture in the present study, NO was not detectable with
WT-ALDH2 and the three Cys-mutants, whereas NO formation was clearly observed in the absence of SOD with the triple mutant (Suppl. Fig. S2). We previously proposed that this phenomenon might be due to significant superoxide production by the WT enzyme, but not by E268Q-ALDH. For an alternative interpretation, see Scheme 2 in the Discussion.

We repeated some of the electrode experiments with 2 µM instead of 0.1 mM GTN (Suppl. Fig. S3). It has been shown previously that NO formation is close to the electrochemical detection limit with 1 µM GTN for the WT enzyme (Beretta et al., 2010). In the present study we attained a peak concentration of ~0.13 µM with 2 µM GTN. Considerably higher peaks were obtained with the double and triple mutants (0.67 and 0.60 µM, respectively), whereas a lower peak was observed for E268Q-ALDH2 (~0.07 µM).

Oxyhemoglobin Oxidation

The NO electrode experiments provided unequivocal evidence for the formation of NO, but because the curves are complex functions of simultaneous NO formation and disappearance, this method does not allow reliable quantification of the amount of NO released, precluding use of the data for calculation of turnover numbers. Therefore we also measured NO formation with the oxyhemoglobin assay. Though more susceptible to possible interferences by redox-active agents than the electrochemical method, this assay has the advantage of measuring product accumulation in addition to reaction rates. As illustrated by Fig. 3 and Suppl. Fig. S4, metHb formation by ALDH/GTN-derived NO was biphasic, with a rapid phase occurring within the first minute of incubation, and a slow phase that continued beyond 20 min. The relative amounts of NO produced in the fast phase by the various enzymes (Table 2) correlated well with the corresponding peak concentrations observed with the NO electrode (Table 1), suggesting that the results of the oxyhemoglobin assay do indeed represent NO formation.
GTN alone caused some oxyHb autoxidation ($\Delta \Delta A_{420-401} = 0.12 \pm 0.01$) that, unlike enzyme-mediated NO formation, did not show a fast initial phase and was virtually abolished by DTT (Fig. S5). After correction for that artifact, [NO]$_{\text{slow}}$ turned out to be negligible in the absence of DTT, with the possible exception of the mutants lacking Glu-268 (Table 2). In the absence of DTT, none of the enzymes catalyzed stoichiometric amounts of NO from GTN within 20 min (~0.3-2 µM NO catalyzed by ~5 µM ALDH2), although elimination of Cys-301 and Cys-303, as well as additional mutation of Glu-268, led to significant increases in NO formation. In the presence of DTT, all enzyme species produced ~2-3 µM NO in 20 min, with the notable exception of the triple mutant, which generated ~10 µM NO, corresponding to 2.2±0.3 turnovers (Table 2).

*Soluble Guanylate Cyclase Activation*

Activation of purified sGC was measured in the presence of wild-type and mutated ALDH2 to test for GTN bioactivation at therapeutically relevant submicromolar concentrations of the nitrate (Fig. 4A). In the absence of both NAD$^+$ and SOD none of the ALDH2 variants stimulated sGC beyond basal activity determined with 0.1 µM GTN in the absence of ALDH2 (~0.2 µmol cGMP x min$^{-1}$ x mg$^{-1}$). NAD$^+$ did not affect basal cGMP formation (not shown) but increased sGC activity in the presence of WT-ALDH2 about 10-fold (1.8±0.5 µmol cGMP x min$^{-1}$ x mg$^{-1}$). A similar effect of NAD$^+$ was observed with C301S-ALDH2, C303S-ALDH2, and the double mutant. In line with a previous report (Wenzl et al., 2009) NAD$^+$ did not affect sGC activation mediated by E268Q-ALDH2. Interestingly, however, the nucleotide cofactor caused a ~7-fold increase in sGC activity in the presence of the triple mutant.

SOD caused an increase in basal sGC activity (~1.4 µmol cGMP x min$^{-1}$ x mg$^{-1}$, not shown), probably due to stabilization of airborne NO (Friebe et al., 1998). Addition of wild-type or mutant ALDH2 (2 µg each) caused only slightly higher rates of cGMP formation in
the presence of SOD (between 2.2±0.3 and 2.7±0.4 µmol x min⁻¹ x mg⁻¹), but pronounced effects were observed in the combined presence of SOD and NAD⁺. Under these conditions WT-ALDH2 stimulated sGC activity to 8.9±0.6 µmol cGMP x min⁻¹ x mg⁻¹. In the presence of the cysteine mutants (C301S, C303S, and the double mutant) the rates of cGMP formation were even higher, ranging from 12.4±0.5 to 14.7±0.7 µmol x min⁻¹ x mg⁻¹. As previously reported (Wenzl et al., 2009), hardly any activation was observed in the presence of E268Q-ALDH2 under these conditions. The effect of the triple mutant was modest, most likely reflecting low apparent GTN affinity (see below).

Fig. 4B illustrates the effect of GTN concentration on GTN-triggered sGC activation mediated by the various ALDH2 species in the presence of SOD and NAD⁺. In the presence of WT-ALDH2 GTN exhibited an EC₅₀ of 52±15 nM, and mutation of Glu-268 led to an approximately 50-fold rightward shift of the curve. The double mutant was the most effective ALDH2 variant, yielding an EC₅₀ as low as 17±8 nM and significantly higher maximal activity as compared to wild-type (18.7±1.0 vs. 11.7±0.4 cGMP x min⁻¹ x mg⁻¹). Again, mutation of Glu-268 to Gln caused a large (~28-fold) rightward shift of the curve.

**Acetaldehyde Dehydrogenation**

Since Cys-301 and Cys-303 may be involved in oxidative enzyme inactivation through formation of a disulfide bridge with Cys-302 after one turnover of GTN denitration, we also investigated inhibition by GTN and restoration by DTT of the aldehyde dehydrogenase activity of the three cysteine mutants in comparison to WT-ALDH2 (Fig. 5). These experiments were not performed with E268Q-ALDH2 and the triple mutant, because in the absence of Glu-268 dehydrogenase activity is hardly detectable under the applied conditions. Fig. 5 shows that oxidation of acetaldehyde (v₀) was not significantly affected by mutation of Cys-301 and/or Cys-303. While single mutation of these residues did not affect the rate of ALDH2 inactivation, the double mutant was inactivated ~2.5x faster than WT-
ALDH ($k_{\text{inact}} (15.2\pm1.3)\times10^{-3}$ s$^{-1}$ vs. $(6.3\pm0.7)\times10^{-3}$ s$^{-1}$). The activity of GTN-exposed WT-ALDH2 was restored to about 45% of $v_0$ by DTT ($v_{\text{restored}}$ in Fig. 5). The cysteine mutants exhibited significantly lower restored activities (~20-30% of $v_0$), indicating that mutation of these residues shifts the effect of GTN towards DTT-irreversible enzyme inactivation.
DISCUSSION

In their initial report on the role of ALDH2 in GTN bioactivation, Stamler and coworkers showed that the enzyme reduces GTN to 1,2-GDN and nitrite, and proposed a mechanism involving formation of a disulfide bridge between the catalytic nucleophile Cys-302 and one of the adjacent cysteines (Chen et al., 2002). The inactivation of ALDH2 after each turnover was shown to be DTT-reversible, suggesting that it contributes to the development of nitrate tolerance. Though this mechanism may account for the main route of ALDH2-catalyzed GTN denitration, our recent work demonstrating an additional pathway yielding NO instead of nitrite (Beretta et al., 2008a) and partially DTT-irreversible inactivation of the enzyme by GTN (Beretta et al., 2008b) indicates that the reaction between ALDH2 and GTN may be more complex than originally assumed. In the present study we mutated the cysteine residues adjacent to the catalytic nucleophile Cys-302 and the putative general base Glu-268 in human ALDH2 to clarify their function in GTN metabolism and GTN-induced enzyme inactivation. The results not only confirmed that GTN denitration can be dissociated from NO formation and ALDH2 inactivation, but also demonstrate that NO formation and ALDH2 inactivation represent different processes. For the sake of brevity and clarity, we will limit the discussion to those aspects of the results that suggest three distinct pathways for the reaction of ALDH2 with GTN. A detailed discussion of other aspects is provided as Supplementary Material.

NO formation and ALDH2 inactivation can be distinguished from clearance-based GTN denitration

WT-ALDH2 produced \( \leq 1 \) equiv. of 1,2-GDN/ALDH2 from GTN in the absence of DTT, while DTT increased the yield approx. 20-fold. In line with the hypothesis that GTN denitration results in DTT-reversible disulfide formation between Cys-302 and one of the
adjacent Cys residues, we found that mutating one or both of these residues impeded multiple turnover, without affecting single turnover.

Mutating the adjacent Cys residues did not affect NO formation to the same extent. Consequently, the mutants showed a considerable increase in the relative yield of NO compared to GTN denitration (from 5% for WT-ALDH2 to 22% for the double mutant in the presence of DTT). These results suggest two distinct pathways for GTN denitration with the minor one yielding NO. They also demonstrate that, at least in vitro, Cys-301 or Cys-303 do not participate in NO formation, and suggest competition between disulfide formation and NO generation.

Mutation of Cys-301 and Cys-303 also affected inactivation of the enzyme. The double mutant was inactivated 2- to 3-fold faster than WT-ALDH2, and all three Cys-mutants showed more pronounced irreversible inactivation, indicating that this process, as well, can be separated from GTN denitration.

**NO formation can be distinguished from ALDH2 inactivation**

The results with the Cys-mutants (C301S, C303S, and the double mutant) demonstrate that ALDH2-catalyzed NO formation can be distinguished from the main GTN denitration pathway. The results obtained with the Glu-268 mutants (E268Q-ALDH2 and the triple mutant) show that NO formation can be dissociated from ALDH2 inactivation as well. As reported and discussed by us previously (Wenzl et al., 2009), E268Q-ALDH2 exhibits generally low activity accompanied by a higher relative contribution of NO formation. The triple mutant combined some of the characteristics of E268Q and the Cys-mutants. Like these mutants, it exhibited a large contribution of NO production. Like E268Q-ALDH2, it displayed rather low activity, though not to the same extent. It shared with the three Cys-mutants a higher production of NO than WT-ALDH2 in single turnover (i.e. in the absence of DTT). However, unlike the other three Cys-mutants, the triple mutant did not experience
rapid inactivation. Consequently, it sustained catalysis over at least 20 min, resulting in several-fold higher NO yields despite a low GTN denitration rate. Indeed, the triple mutant converts GTN almost exclusively to 1,2-GDN and NO.

Most importantly, the triple mutant produced superstoichiometric NO (in 20 min > 2 NO/ALDH active site), which demonstrates that NO formation is a catalytic process that is distinguishable from enzyme inactivation. Three main conclusions can be drawn from the present data. Clearance-based denitration of GTN, NO formation, and irreversible ALDH2 inactivation are distinct processes, Cys-301 and Cys-303 are critically involved in the main denitration pathway, and Glu-268 plays a central role in irreversible enzyme inactivation.

**Proposed Reaction Mechanism**

As illustrated in Scheme 1, the results suggest that ALDH2-catalyzed GTN biotransformation involves three distinct pathways. In a first step common to all three, GTN reacts with Cys-302 of the enzyme (E<sub>red</sub>) to form an intermediate (E<sub>int</sub>). The main reaction catalyzed by the wild-type enzyme results in formation of nitrite and DTT-reversible inactivation (path a). This pathway requires Cys-301 and Cys-303. A smaller fraction of the enzyme (path b) is inactivated irreversibly in a reaction that is strongly promoted by Glu-268. The third pathway yields NO in a reaction that results in formation of a DTT-reducible oxidized enzyme species (path c). This reaction cycle predominates for the triple mutant.

A more concrete, but inevitably more speculative version of the proposed model is presented in Scheme 2. GTN denitration (A) requires Cys-302 and appears to yield 1,2-GDN almost exclusively. Accordingly, and in line with previous proposals (Chen et al., 2002), the intermediate (E<sub>int</sub> in Scheme 1) may be a thionitrate. As previously suggested (Beretta et al, 2008a), the thionitrate may reversibly rearrange to a sulfenyl nitrite as an alternative intermediate (B). The main pathway catalyzed by the wild-type enzyme most likely involves formation of nitrite and a disulfide between Cys-302 and one of the vicinal cysteines (C).
DTT can regenerate the reduced cysteine residues, thereby enabling catalytic conversion of GTN to 1,2-GDN and nitrite (clearance-based GTN metabolism).

Accepting the putative role of Glu-268 as a general base catalyst, this residue is expected to affect the reaction by promoting electrophilic attack of the intermediate on a nascent hydroxide ion. How this might affect the outcome of the reaction is unclear. One attractive possibility, illustrated in Scheme 2, is that the intervention of Glu-268 results in heterolysis of the sulfenyl nitrite, which might yield a protein-sulfinate and nitroxyl (D). This would explain both irreversible inactivation (by formation of a sulfinate that is not reducible by DTT) and SOD-dependent NO formation (because SOD oxidizes nitroxyl to NO, Zeller et al., 2009).

For the third pathway, we suggest homolytic cleavage of the sulfenyl nitrite (E). DTT may react with the sulfinyl radical or a subsequently formed species to regenerate Cys-302, thereby enabling multiple turnovers of NO formation. Notwithstanding the lack of information on the structure of reaction intermediates and the oxidized species formed upon exposure of the protein to GTN, the present study demonstrates that ALDH2 can produce NO directly from GTN in a catalytic process and, thus, supports our proposal that ALDH2-catalyzed reduction of GTN to NO essentially contributes to vascular bioactivation of GTN in vivo.

**Physiological Relevance**

The surprising discovery that bioactivation of GTN involves catalysis by ALDH2 (Chen et al., 2002), raised the question how nitrite, the main product of GTN bioconversion by ALDH2, is linked to vascular relaxation. Originally it was suggested that reduction of nitrite by components of the mitochondrial respiratory chain might provide that link. However, known affinities of nitrite for such components appear too low, and interfering
with respiration did not affect mitochondrial biotransformation of GTN (Kollau et al., 2009), suggesting that ALDH2 catalysis alone is sufficient.

As a possible solution we found that ALDH2 catalyzed direct NO formation from GTN in a minor pathway, accounting for 5-10% of the total yield (Beretta et al., 2008a). We also demonstrated that ALDH2 is irreversibly inactivated by GTN during catalysis, which might offer a potential mechanism for the development of nitrate tolerance (Beretta et al., 2008b). However, these issues remain controversial, partly because of the lack of a detailed mechanism for the proposed reactions. In addition, the apparent linkage between NO formation and ALDH2 inactivation seemed to limit the production of NO to one turnover, which greatly diminished the potential physiological significance of the observed reactions. The present study goes some way in answering the remaining questions. In summary:

(i) ALDH2 catalyzes three distinct pathways of GTN biotransformation that involve different active site residues.

(ii) The assignment of specific functions of active site residues to the different catalytic pathways allows the first data-based attempt to describe the complex mechanism of ALDH2-catalyzed GTN bioconversion.

(iii) Clearance-based metabolism yielding nitrite and bioactivation yielding NO involve two separate pathways, implying that 1,2-glyceryl dinitrate formation measured in blood vessels may not be a valid measure for vascular ALDH2-catalyzed GTN bioactivation.

(iv) The demonstration that NO formation is catalytic and occurs through a pathway distinct from that resulting in irreversible enzyme inactivation greatly increases the potential in vivo relevance of direct NO formation.

(v) Finally, the observation that direct NO formation can be dissociated from irreversible inactivation and clearance-based denitrification may pave the way for the design of
ALDH2 substrates that are more efficient vasodilators and less prone to develop vascular
tolerance than GTN.

**AUTHORSHIP CONTRIBUTIONS**

*Participated in research design:* Wenzl, Beretta, Russwurm, Koesling, Schmidt, Mayer, and
Gorren

*Conducted experiments:* Wenzl, Beretta, and Griesberger

*Contributed new reagents or analytic tools:* Russwurm and Koesling

*Performed data analysis:* Wenzl, Beretta, Griesberger, Schmidt, Mayer, and Gorren

*Wrote or contributed to the writing of the manuscript:* Wenzl, Mayer, and Gorren
REFERENCES


FOOTNOTES

This work was supported by the Fonds zur Förderung der Wissenschaftlichen Forschung in Austria [Grants P20669, P21693, W901 DK Molecular Enzymology]; and the Deutsche Forschungsgemeinschaft [Grant KO1157/4-1].

Reprint requests:
Dr. Ton Gorren
Dept. of Pharmacology & Toxicology
Karl-Franzens-Universität Graz
Universitätsplatz 2, A-8010, Austria
Tel.: 43-316-380 5569
Fax: 43-316-380 9890
e-mail: antonius.gorren@uni-graz.at

1 Although GTN denitration is a necessary first step in the formation of NO, the thionitrate is metabolized along three different pathways, only one of which yields NO. Accordingly, denitration is not inseparably linked to NO formation, and the NO-to-1,2-GDN-ratio may vary widely.
FIGURE LEGENDS

Scheme 1. Illustration of the proposed pathways involved in ALDH2-catalyzed GTN biotransformation. Three pathways for ALDH2-catalyzed GTN denitration are inferred. Since all reactions strictly depend on Cys-302, we propose an initial step, common to all three pathways, in which the enzyme (E_red) reacts with GTN to form an intermediate (E_int). From there, the main pathway results in DTT-reversible inactivation and formation of nitrite (path $a$). This pathway requires the presence of Cys-301 and Cys-303. A smaller fraction of the enzyme (path $b$) is inactivated irreversibly in a reaction that is strongly promoted by Glu-268. The present results identify a third pathway that yields NO and results in DTT-reversible inhibition (path $c$). This reaction cycle, which will produce NO in superstoichiometric amounts, is the dominant pathway for the triple mutant.

Scheme 2. Hypothetical reaction mechanism for ALDH2-catalyzed GTN denitration. A: GTN denitration requires Cys-302 and yields 1,2-GDN almost exclusively. Consequently, the intermediate formed in the reaction between GTN and ALDH2 is most likely a Cys-302-centered thionitrate. B: Reversible rearrangement of the thionitrate may yield a sulfenyl nitrite as an alternative intermediate. C: The main pathway for the wild-type enzyme probably involves formation of nitrite and a disulfide bridge between Cys-302 and one of the vicinal cysteines. Certain thiols like DTT enable multiple turnovers of this pathway by reduction of the disulfide. D: Glu-268 may stimulate hydrolysis of the sulfenyl nitrite, yielding nitroxyl and an irreversibly inactivated sulfinate at Cys-302. E: Homolysis of the sulfenyl nitrite may yields NO and a sulfanyl radical. Superstoichiometric NO formation with the triple mutant suggests that DTT can regenerate the reduced Cys-302 residue from this or a subsequently formed enzyme species.
Figure 1. GTN denitration by wild-type and mutant ALDH2. WT or mutant (C301S, C303S, C301S/C303S, E268Q, or E268Q/C301S/C303S) ALDH2, 0.36 µM final) was incubated with 2 µM or the indicated concentration of 14C-labeled GTN in the absence (A) or presence (B) of 2 mM DTT at 37°C for 10 min in a final volume of 200 µL K-phosphate (50 mM, pH 7.4), containing 3 mM MgCl₂ and 1 mM NAD⁺. Reaction products were extracted and quantified by radio thin layer chromatography as described in Materials & Methods. Data are the mean values ± S.E. of three to five independent experiments and expressed as mol 1,2-GDN catalyzed by 1 mol of ALDH2 monomer.

Figure 2. Formation of GTN-derived NO in the presence of WT and mutant ALDH2. ALDH2 (WT, C301S/C303S or E268Q/C301S/C303S, 4.5 µM final) was incubated in 0.5 mL of 50 mM TEA/HCl (pH 7.4), containing 1000 U/mL SOD, 3 mM MgCl₂, 0.1 mM DTPA, 1 mM NAD⁺, and 0.1 mM GTN at 37°C in the absence and presence of 2 mM DTT. NO formation was monitored with a Clark-type electrode. The traces shown are representative for three to six similar experiments.

Figure 3. Formation of metHb from oxyHb by GTN-derived NO in the presence of WT and mutant ALDH2. WT or mutant ALDH2 (WT, C301S/C303S or E268Q/C301S/C303S, 4.5 µM final) was incubated in a final volume of 250 µL K-phosphate (50 mM, pH 7.4) containing 1000 U/mL SOD, 10 U/mL catalase, 0.1 mM DTPA, 1 mM NAD⁺, 3 mM MgCl₂ and 10 µM oxyHb in the absence (A) or presence (B) of 2 mM DTT at 37°C, followed by addition of GTN (0.1 mM). The traces plot the changes in the absorption difference between 420 and 401 nm (peak-minus-trough in the oxyHb/metHb difference spectrum) over time (ΔΔA420-401), and are representative for three to five similar experiments.
Figure 4. Effects of NAD+ and SOD on sGC activation by GTN in the presence of WT and mutant ALDH2. A. Purified bovine lung sGC (50 ng) was incubated at 37°C for 2 min in a final volume of 100 µL with 0.1 µM GTN and 0.36 µM of WT or mutant (C301S, C303S, C301S/C303S, E268Q, or E268Q/C301S/C303S) ALDH2. NAD+ (1 mM) and SOD (1000 U/mL) were present as indicated. Assay mixtures contained 50 mM TEA/HCl (pH 7.4), 0.5 mM [α-32P]GTP (~250 000 cpn), 3 mM MgCl2, and 1 mM cGMP. 32P-cGMP was isolated and quantified as described in Materials & Methods. Data are mean values ± S.E. of three to five independent experiments. B. GTN concentration dependence. Purified bovine lung sGC (50 ng) was incubated at 37°C for 2 min as described above with 1 mM NAD+ and 1000 U/mL SOD and the indicated concentrations of GTN in the presence of WT or mutant (E268Q, C301S/C303S, or E268Q/C301S/C303S) ALDH2 (0.36 µM each). Data are mean values ± S.E. of three to five independent experiments.

Figure 5. Inactivation of WT and mutant ALDH2 by GTN. GTN-induced inactivation of WT and mutant ALDH2 (C301S, C303S, and C301S/C303S) was assayed in the presence of 0.4 mM acetaldehyde, 0.4 mM NAD+ and 10 mM MgCl2 in 250 µL K-phosphate (50 mM, pH 7.4). After 2 min the reaction was started by addition of ALDH2 (0.59 µM) and monitored to obtain initial rates of acetaldehyde oxidation (v0). About 4 min later 50 µM GTN was added for determination of inactivation rate constants (k_{inact}) by fitting to a single-exponential curve. After complete inactivation 0.4 mM DTT was added to determine restored activities (v_{restored}). Data are mean values ± S.E. of four to seven independent experiments.
Table 1. GTN-derived NO formation in the presence of WT and mutant ALDH2.

WT or mutant ALDH2 (4.5 µM) was incubated in 0.5 mL TEA/HCl (50 mM, pH 7.4), containing 3 mM MgCl2, 1 mM NAD+, 0.1 mM DTPA, 1000 U/mL SOD, and 0.1 mM GTN at 37°C in the absence and presence of 2 mM DTT. NO formation was monitored with a Clark-type electrode. Data are mean values ± S.E. of 3-6 independent experiments.

<table>
<thead>
<tr>
<th>ALDH2 variant</th>
<th>[NO]_{peak}</th>
<th>NO formation rate</th>
<th>NO half-life</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>µM</td>
<td>nmol x min^{-1} x mg^{-1}</td>
<td>s</td>
</tr>
<tr>
<td>WT</td>
<td>0.15±0.02</td>
<td>1.21±0.03</td>
<td>3.2±0.5</td>
</tr>
<tr>
<td>E268Q</td>
<td>0.14±0.01</td>
<td>0.17±0.02</td>
<td>3.9±0.2</td>
</tr>
<tr>
<td>C301S</td>
<td>0.57±0.07</td>
<td>1.20±0.07</td>
<td>12.2±1.5</td>
</tr>
<tr>
<td>C303S</td>
<td>0.37±0.04</td>
<td>1.52±0.04</td>
<td>8.9±1.6</td>
</tr>
<tr>
<td>C301S/C303S</td>
<td>0.66±0.02</td>
<td>1.83±0.02</td>
<td>23.4±2.0</td>
</tr>
<tr>
<td>E268Q/C301S/C303S</td>
<td>0.50±0.05</td>
<td>1.73±0.05</td>
<td>11.6±0.4</td>
</tr>
</tbody>
</table>
Table 2. Formation of metHb from oxyHb by GTN-derived NO.

WT or mutant ALDH2 (4.5 µM) was incubated for 20 min in 0.25 mL K-phosphate (50 mM, pH 7.4) containing 1000 U/mL SOD, 10 U/mL catalase, 0.1 mM DTPA, 1 mM NAD+, 3 mM MgCl₂, and 10 µM oxyHb in the absence or presence of 2 mM DTT, followed by addition of 0.1 mM GTN at 37°C. Blank values were determined in the absence of protein under otherwise identical conditions and subtracted. Curves were fitted as explained in Materials & Methods to estimate fast and slow contributions to NO formation. Please note that in many instances, especially in the presence of DTT, NO was still being formed at the end of the incubation, so that [NO]_{slow} and the NO/ALDH stoichiometry merely represent the values observed after 20 min. Results are mean values ± S.E. determined in 3-5 independent experiments.

<table>
<thead>
<tr>
<th>ALDH2 species</th>
<th>[NO]_{fast}</th>
<th>[NO]_{slow}</th>
<th>[NO]_{20min}/[ALDH]</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>μM</td>
<td>μM</td>
<td></td>
</tr>
<tr>
<td>WT</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>–DTT</td>
<td>0.36±0.01</td>
<td>1.97±0.07</td>
<td>0.94±0.13</td>
</tr>
<tr>
<td>+DTT</td>
<td>1.97±0.07</td>
<td>0.94±0.13</td>
<td>0.08±0.01</td>
</tr>
<tr>
<td>–DTT</td>
<td>0.94±0.13</td>
<td>0.08±0.01</td>
<td>0.65±0.03</td>
</tr>
<tr>
<td>E268Q</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>–DTT</td>
<td>0.35±0.03</td>
<td>0.4±0.2</td>
<td>1.95±0.01</td>
</tr>
<tr>
<td>+DTT</td>
<td>0.50±0.03</td>
<td>1.95±0.01</td>
<td>0.17±0.04</td>
</tr>
<tr>
<td>–DTT</td>
<td>1.95±0.01</td>
<td>0.17±0.04</td>
<td>0.54±0.01</td>
</tr>
<tr>
<td>C301S</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>–DTT</td>
<td>1.06±0.02</td>
<td>-0.2±0.2</td>
<td>0.87±0.15</td>
</tr>
<tr>
<td>+DTT</td>
<td>1.5±0.2</td>
<td>0.87±0.15</td>
<td>0.24±0.01</td>
</tr>
<tr>
<td>–DTT</td>
<td>0.87±0.15</td>
<td>0.24±0.01</td>
<td>0.53±0.06</td>
</tr>
<tr>
<td>C303S</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>–DTT</td>
<td>0.52±0.02</td>
<td>-0.2±0.2</td>
<td>0.1±0.6</td>
</tr>
<tr>
<td>+DTT</td>
<td>2.15±0.09</td>
<td>0.1±0.6</td>
<td>0.12±0.01</td>
</tr>
<tr>
<td>–DTT</td>
<td>0.1±0.6</td>
<td>0.12±0.01</td>
<td>0.50±0.13</td>
</tr>
<tr>
<td>C301S/C303S</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>–DTT</td>
<td>1.2±0.3</td>
<td>0.2±0.3</td>
<td>0.20±0.04</td>
</tr>
<tr>
<td>+DTT</td>
<td>2.9±0.4</td>
<td>0.20±0.04</td>
<td>0.27±0.09</td>
</tr>
<tr>
<td>–DTT</td>
<td>0.20±0.04</td>
<td>0.27±0.09</td>
<td>0.69±0.09</td>
</tr>
<tr>
<td>E268Q/C301S/C303S</td>
<td>1.18±0.06</td>
<td>3.1±0.3</td>
<td>0.9±0.4</td>
</tr>
</tbody>
</table>

\( a \) The slow phase in the absence of DTT did not significantly differ from zero except for the mutants lacking Glu-268. \( b \) The slow phase was ignored for the estimation of the NO/ALDH stoichiometry in the absence of DTT for WT-, C301S-, and C303S-ALDH2.
Scheme 1

**Chemical Reactions:**

1. **DTT**
   - $E_{red} \rightarrow E_{int}$
   - $E_{int} \rightarrow E_{ox}$

2. **GTN**
   - $E_{red} \rightarrow E_{inact}$

3. **Cys-302**
   - $E_{red} \rightarrow E_{int}$
   - $E_{int} \rightarrow E_{ox}$

4. **Cys-301/3**
   - $E_{int} \rightarrow E_{ox}$

5. **E_{ox}'**
   - $E_{ox}' \rightarrow E_{int}$

6. **NO**
   - $E_{int} \rightarrow E_{ox}$
   - $E_{int} \rightarrow E_{ox}'$

7. **Glu-268**
   - $E_{int} \rightarrow E_{inact}$

8. **NO_2^-**
   - $E_{ox} \rightarrow E_{ox}$
Figure 2

Graph showing the concentration of NO (µM) over time (s) for different samples.

- WT
- C301S/C301S
- C301S/C303S + DTT
- E268Q/C301S/C303S + DTT

The graph illustrates the NO production kinetics for each condition.
Figure 3
Figure 4

A

\[ \mu \text{mol cGMP x min}^{-1} \times \text{mg}^{-1} \]

- WT
- E268Q
- C301S
- C303S
- C301S/C303S
- E268Q/C301S/C303S

B

\[ \mu \text{mol cGMP x min}^{-1} \times \text{mg}^{-1} \]

\[ \text{GTN (\mu M)} \]

- WT
- E268Q
- C301S/C303S
- E268Q/C301S/C303S
Figure 5
SITE-DIRECTED MUTAGENESIS OF ALDEHYDE DEHYDROGENASE-2 SUGGESTS THREE DISTINCT PATHWAYS OF NITROGLYCERIN BIOTRANSFORMATION*

M. Verena Wenzl, Matteo Beretta, Martina Griesberger, Michael Russwurm, Doris Koesling, Kurt Schmidt, Bernd Mayer, and Antonius C.F. Gorren

Department of Pharmacology and Toxicology, Karl-Franzens-Universität Graz, A-8010 Graz, Austria (M.V.W., M.B., M.G., K.S., B.M., A.C.F.G.)

Department of Pharmacology and Toxicology, Ruhr-Universität Bochum, D-44780 Bochum, Germany (M.R., D.K.)
SUPPLEMENTARY FIGURES

Figure S1. ALDH2-catalyzed NO release by GTN as measured with the NO-sensitive electrode.

Panels A and B show the first 50 s of representative traces in the absence (A) or presence (B) of DTT to illustrate the initial rates of NO formation. The initial rates in Table 1 were estimated from the steepest rise between consecutive data points (1.5 s apart, indicated by circles). Alternatively, we determined initial rates from fits to the initial parts of the curve (indicated by filled symbols) to an appropriate function (monoexponential or linear). Values obtained in this way were somewhat (1.2 to 1.6-fold) higher. Panels C, D, and E, illustrate the effect of DTT on the rates of NO formation for WT-ALDH2, the double, and the triple mutant respectively. Panels F, G, and H show traces for WT-ALDH2, the double, and the triple mutant respectively, on extended time scales (up to 30 min in the case of the triple mutant), to illustrate the effect of DTT on the decay of the NO signal. Decay rates were estimated from monoexponential fits to the latter parts of the curves (indicated by circles). Experimental conditions were as for Fig. 1. CC and ECC stand for the C301S/C303S double mutant and the E268Q/C301S/C303S triple mutant, respectively.

Figure S2. ALDH2-catalyzed NO release by GTN in the absence of SOD.

Representative traces are shown for NO detection from 0.1 mM GTN in the presence of NAD⁺ (1 mM) and DTT (2 mM) for the double mutant (red), and the triple mutant (blue). Experimental conditions as in Fig. 1, except for the absence of SOD.

Figure S3. ALDH2-catalyzed NO release from 2 µM GTN as measured with the NO-sensitive electrode.

Shown are representative plots for the detection of NO from 2 µM GTN in the presence of NAD⁺ (1 mM), DTT (2 mM), and SOD (1000 U/mL) for WT-ALDH2 (black), E268Q-
ALDH2 (green), the double mutant (red), and the triple mutant (blue). Experimental conditions as in Fig. 1, except for the lower concentration of GTN. Estimated initial NO formation rates: 12 nM/s for WT, 6.6 nM/s for E268Q, 58 nM/s for the double mutant, 39 nM/s for the triple mutant.

**Figure S4. ALDH2-catalyzed NO release by GTN as measured with the oxyhemoglobin assay.**

Representative traces for different enzyme species in the absence (A) and presence (B) of DTT. Note the similar slow phases in the absence of DTT for all enzymes, including the inactive C302S-mutant, in panel A. Experimental conditions were as for Fig. 3. CC and ECC stand for the C301S/C303S double mutant and the E268Q/C301S/C303S triple mutant, respectively. C302S-ALDH is virtually inactive with respect to GTN denitration (~2% of WT, Wenzl al., 2009), and yields no signal at the NO electrode (results not shown).

**Figure S5. Illustration of the artifactual nature of the slow phase in methemoglobin formation in the absence of DTT.**

Representative traces for GTN-induced methemoglobin formation in the absence (black) and presence (blue) of WT-ALDH2, as well as the difference between both curves (red). Also shown is a trace for methemoglobin formation by GTN without ALDH2 in the presence of DTT (green). Note that the slow phase in the absence of DTT without enzyme fully accounts for the slow phase observed in the presence of WT-ALDH2, and is abolished in the presence of DTT.
Supplementary Figure S3
SUPPLEMENTARY DISCUSSION

GTN Denitration

As expected, mutation of one or both cysteines did not affect GTN denitration in the absence of DTT, i.e. under single turnover conditions (see Fig. 1A), with yields corresponding to 0.6-1.0 turnovers. In agreement with the proposed formation of a disulfide after each turnover, rates of denitration catalyzed by WT-ALDH2 were markedly increased by DTT and this effect was virtually abolished upon mutation of the two cysteine residues adjacent to Cys-302. Surprisingly, however, a similar effect was observed when only one of these cysteines was eliminated (see Fig. 1B), suggesting that mutation of one cysteine impedes the reactivity of the remaining one, perhaps by increasing the apparent pK_a value. It is likely that the thiolate anion is the (more) reactive state of the cysteines for disulfide formation. Since the proximity of the thiols to each other is expected to lower the pK_a values, elimination of either one might reduce the reactivity of the other. The results also suggest that both thiols are more or less equivalent. Although the inhibitory effect of mutating one or both cysteines was extensive, all mutants catalyzed more than one turnover in the presence of DTT. A possible explanation is slow formation of a disulfide bridge between DTT and Cys-302 and regeneration of the enzyme by the second DTT thiolate group or by another DTT molecule.

The effect of mutation of Glu-268 is dominated by the previously observed inhibition of all activities in the presence of NAD^+, which has been ascribed to unfavorable binding of the nucleotide cofactor (Wenzl et al., 2009). The activity of the triple mutant, although still low, was considerably higher, suggesting that removal of both cysteines changes the binding mode of NAD^+ to a more favorable position. Although Glu-268 is not essential for GTN denitration (Wenzl et al., 2009), it is conceivable that it substitutes for Cys-301 and/or Cys-303 in GTN denitration by the double mutant. The low activity of the triple mutant compared
to the double mutant would seem to corroborate that, but more likely reflects the overall lower activity of the glutamate mutants.

*Acetaldehyde Dehydrogenation*

Mutation of Cys-301 and/or Cys-303 did not affect acetaldehyde oxidation (see Fig. 5). The rate of inactivation by GTN was not affected by elimination of one cysteine, but increased when both cysteines adjacent to Cys-302 were eliminated. Since the rate of inactivation reflects the reaction rate with GTN (Beretta et al., 2008b), this observation suggests that the double mutant reacts faster with GTN than the wild-type enzyme. The activities of the GTN-exposed cysteine mutants restored by DTT were significantly lower as compared to wild-type, suggesting a shift towards irreversible inactivation of the enzyme. This observation supports the assumption that the DTT-reversible fraction of inactivated enzyme reflects disulfide formation.

*NO Formation*

The results on NO formation (Figs. 2 and 3, Tables 1 and 2) demonstrate that Cys-301 and Cys-303 do not participate in 3-electron reduction of GTN. This corroborates our previous proposal (Wenzl et al., 2009, Beretta et al., 2010) that ALDH2-catalyzed clearance-based GTN denitration and bioactivation (yielding NO) represent distinct reactions. In the absence of DTT, elimination of one or both cysteines increased NO formation ~3-4-fold, as revealed by the peak concentrations in the electrode experiments as well as by the fast phase in the oxyhemoglobin assay. These observations suggest that in WT-ALDH2 the NO generating reaction competes with Cys-301/Cys-303-supported clearance-based denitration.

In the presence of DTT, regeneration of Cys-302 not only enables multiple turnovers of clearance-based denitration but also allows for additional NO formation. Consequently, DTT is expected to increase the yield of NO, particularly for WT-ALDH2, which exhibited
the greatest capacity for DTT-supported multiple turnovers (see Fig. 1). Indeed, DTT increased NO peak height in the electrode studies 8-fold for WT-ALDH2, while smaller increases were observed with the cysteine mutants. The latter effect suggests that the NO producing pathway is also stimulated by DTT to some extent.

To understand the observations with the NO electrode at low [GTN] (Fig. S3), it is important to realize that the impact of multiple turnovers will be diminished at low GTN concentrations. Consequently, the striking difference in shape of the traces of the double mutant on the one hand, and the triple mutant and WT-ALDH2 on the other (see Fig. 2) disappears, and all NO peaks decay with similar apparent rates (Fig. S3). Indeed, there is a striking resemblance between the traces observed at low GTN concentrations in the presence of DTT (Fig. S3) and the traces observed at 100 µM GTN in the absence of DTT, i.e. under single turnover conditions (Fig. 2). Another important factor is the affinity for GTN of the various enzymes, which comes into play more prominently at lower GTN concentrations. A low affinity for GTN is the cause of the small peak height with E268Q-ALDH2 (~0.07 µM). Finally, NO peak heights will tend to be reduced at higher GTN concentrations, since NO autoxidation, which is second-order in [NO], will become relatively faster.

According to real-time monitoring in the oxyhemoglobin assay, NO formation was biphasic, although the slower phase was absent with some ALDH2 variants (see Fig. 3). In the absence of DTT, a slow phase was also observed when GTN was added to oxyhemoglobin in the absence of ALDH2, exposing this phase as an artifact caused by a reaction between oxyhemoglobin and GTN. After subtraction of this phase from all curves, no slow phase remained for the WT-enzyme and the cysteine mutants (see Table 2). In the presence of DTT, no slow artifacts were observed, indicating that DTT can prevent or reverse the reaction between oxyhemoglobin and GTN. Roughly speaking, the fast phase corresponds to rapid NO formation in the first turnover, whereas subsequent turnovers give rise to the slow phase. However, because of their low affinity for GTN, the two mutants lacking Glu-268 react with
GTN so slowly that part of the NO generated in the first turnover spills over into the slow phase. The pronounced biphasic character of the results obtained in the presence of DTT demonstrates that the rate-limiting step in catalysis occurs after NO generation, suggesting that it involves regeneration of active ALDH2.

Generally, the results obtained with the oxyhemoglobin assay confirm the electrode data, showing that DTT caused a 10-fold increase in the amount of NO formed after 20 min in the presence of WT-ALDH2, and smaller increases (2- to 4-fold) for the cysteine mutants. The results also show that, although the cysteine mutants tend to produce more NO in single-turnover (from 7% for the wild-type enzyme to 26 % for the double mutant), the total amount of NO produced after 20 min in the presence of DTT was similar for all variants, with values ranging from 2.3±0.6 to 3.1±0.4 μM. This may be due to increased susceptibility of the cysteine mutants to irreversible inactivation, in line with the results on acetaldehyde dehydrogenation (see Fig. 5) and the observed decay of the NO electrode signals (see Fig. 2 & Table 1).

The results discussed so far suggest a model involving two pathways of ALDH2-catalyzed GTN biotransformation, one leading to formation of nitrite and a (DTT-reversible) protein disulfide, and the other resulting in 3-electron reduction of GTN to yield NO and an inactivated enzyme species that is not reduced by DTT. However, the results obtained with E268Q-ALDH2 and the triple mutant compel us to revise this model. As observed previously (Wenzl et al., 2009), mutation of Glu-268 led to a pronounced shift from clearance-based GTN decomposition towards the NO generating pathway. In the absence of DTT, the NO electrode response resembled that of the wild-type enzyme, and a similar fast phase was also apparent in the oxyhemoglobin assay. However, unlike the enzyme species discussed thus far, the E268Q mutant additionally showed a considerable slow phase. Consequently, E268Q-ALDH2 catalyzed formation of significantly more NO than WT-ALDH2. In the presence of DTT, NO formation was slower but more sustained with E268Q-ALDH2 (Table 2), resulting
in a yield of NO in the same range as that of WT-ALDH2 and the cysteine mutants. The results suggest that mutation of Glu-268 lowers the reactivity of the enzyme towards GTN, but increases its resistance against irreversible inactivation.

Even more telling were the observations with the triple mutant. While sharing the low reactivity of E268Q-ALDH2 regarding aldehyde oxidation and GTN denitration, it resembled the cysteine mutants in producing considerably more NO under single-turnover conditions (without DTT). Results were most striking, however, in the presence of DTT. Under these conditions the triple mutant produced far more NO than the other enzymes in a sustained slow reaction (Fig. 3B). In fact, the triple mutant produced superstoichiometric concentrations of NO after 20 min (>2 NO/ALDH active site), indicating that NO formation by the triple mutant is a catalytic process that can be dissociated from irreversible inactivation. The results suggest that eliminating the adjacent cysteines increases NO production by driving the reaction away from clearance-based GTN denitration, whereas mutation of Glu-268 increases NO production through greater resistance of the enzyme to irreversible inactivation.

**sGC activation**

The main purpose of determination of sGC activation by ALDH2/GTN in the present study was to investigate if the processes observed in the other experiments at micromolar concentrations of GTN are still operative at physiologically relevant submicromolar concentrations. The general conclusion from the studies is that this is indeed the case. For several reasons, however, sGC activity is not directly comparable with the observations with the NO electrode and the oxyhemoglobin assay, presented in Tables 1 and 2 respectively. Most importantly, due to the very low (subnanomolar) dissociation constant of sGC for NO, the rates of cGMP formation are strongly affected by the apparent GTN affinities of the various mutants, and saturation at low GTN may result in loss of information on the rates of
NO formation. In addition, alternate reaction products and/or intermediates (e.g. reactive oxygen species, nitrosothiols) are likely to affect sGC activity.

An important consequence of the application of the sGC assay to measure NO formation is an immense increase in sensitivity. For instance, GTN denitration by WT-ALDH2 exhibited a $K_m$ of approx. 3 µM GTN. However, because of the extremely high affinity of sGC for NO, sGC activation by WT-ALDH2/GTN occurred with an EC$_{50}$ of 50 nM. The observed EC$_{50}$ for sGC activation by GTN will depend on the amount of NO produced, which, in turn, will be determined by the ALDH2 concentration, the GTN concentration, the apparent affinity of ALDH2 for GTN, the maximal activity of ALDH2, and the incubation time.

Introduction of NAD$^+$ or SOD into a ALDH2/GTN assay mixture that hardly activated sGC caused a considerable increase in cGMP formation, while addition of both caused a greater than additive effect. This can be explained by the different modes of action of the two compounds. Whereas NAD$^+$ causes a general activity increase, resulting in higher formation rates of NO and the presumed co-product O$_2^-$, SOD acts by scavenging superoxide (alternatively, it may transform HNO into NO, see main text); the combination can be shown to have greater than additive effects. However, whereas the effects of SOD and NAD$^+$ alone were similar for WT-ALDH2 and the Cys-mutants, the combination had a greater effect on the Cys-mutants. This suggests that the difference between WT-ALDH2 and the mutants cannot be ascribed to a simple change in NO availability. This is also suggested by the higher maximal activities observed for the cysteine mutants. The origin of the additional activation by the cysteine mutants is presently unclear.

E268Q-ALDH2 did not perform well under the conditions of the sGC assay when NAD$^+$ was also present. As was noted previously (Wenzl et al., 2009), NAD$^+$, which stimulates the GTN denitration by the wild-type enzyme, greatly suppresses GTN denitration
by E268Q-ALDH2, as well as direct NO formation, although not to the same extent. The cause of these inhibitory effects is not known yet, but may involve transformation to an unproductive conformation of the enzyme (Wenzl et al., 2009). In addition, the present results suggest a marked decrease of the apparent affinity of the enzyme for GTN. Consequently, the conditions applied in the sGC assay (low GTN concentration, high NAD$^+$ concentration, short incubation time) were very unfavorable for the E268Q mutant. The same is true for the triple mutant as well, although in this case the ability of the enzyme to continue producing NO over longer periods partly compensates for the inhibition by NAD$^+$. It is to be expected that under more suitable conditions (lower enzyme concentration, longer incubation times, and less NAD$^+$) E268Q-ALDH2 and, particularly, the triple mutant will outperform the other ALDH species. More work is needed to resolve these issues.