

Molecular Characterization of Binding Loop E in the Nematode Cys-Loop GABA Receptor[§]

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Received May 2, 2018; accepted August 30, 2018

ABSTRACT

Nematodes exhibit a vast array of cys-loop ligand-gated ion channels with unique pharmacologic characteristics. However, many of the structural components that govern the binding of various ligands are unknown. The nematode cys-loop GABA receptor uncoordinated 49 (UNC-49) is an important receptor found at neuromuscular junctions that plays an important role in the sinusoidal movement of worms. The unique pharmacologic features of this receptor suggest that there are structural differences in the agonist binding site when compared with mammalian receptors. In this study, we examined each amino acid in one of the main agonist binding loops (loop E) via the substituted cysteine accessibility method (SCAM) and analyzed the interaction of various residues by molecular dynamic

simulations. We found that of the 18 loop E mutants analyzed, H142C, R147C, and S157C had significant changes in GABA EC₅₀ and were accessible to modification by a methanethio-sulfonate reagent (MTSET) resulting in a change in *I*_{GABA}. In addition, the residue H142, which is unique to nematode UNC-49 GABA receptors, appears to play a negative role in GABA sensitivity as its mutation to cysteine increased sensitivity to GABA and caused the UNC-49 receptor partial agonist 5-aminovaleric acid (DAVA) to behave as a full agonist. Overall, this study has revealed potential differences in the agonist binding pocket between nematode UNC-49 and mammalian GABA receptors that could be exploited in the design of novel anthelmintics.

Introduction

Nematodes such as the model free-living nematode *Caenorhabditis elegans* and parasitic nematodes such as *Haemonchus contortus* contain a vast array of cys-loop ligand-gated chloride channels that respond to a variety of neurotransmitters including serotonin, tyramine, dopamine, GABA, and glutamate (Jones and Sattelle, 2008; Bartos et al., 2009; Laing et al., 2013). In addition, nematodes exhibit a unique class of acetylcholine-gated chloride channels (Putrenko et al., 2005) and GABA-gated cation channels (Beg and Jorgensen, 2003). However, like vertebrates, nematodes also exhibit anion selective receptors for GABA.

The nematode cys-loop GABA receptor that is the most characterized pharmacologically and functionally is the uncoordinated 49 (UNC-49) receptor (Bamber et al., 1999; Siddiqui et al., 2010). This receptor is expressed at the neuromuscular junctions of nematodes and contributes to the worm's sinusoidal movement (Bamber et al., 2005). With regard to amino acid sequence, the UNC-49 receptor does not appear to be

analogous to any mammalian GABA receptor, and it appears to exhibit unique pharmacologic features. One significant pharmacologic difference is the low sensitivity of the classic GABA receptor antagonist bicuculline (Bamber et al., 2003), which is a characteristic shared among invertebrate GABA receptors (Hosie et al., 1997). Differences have also been observed between the agonist profile of the UNC-49 receptor from *H. contortus* (Hco-UNC-49) and the mammalian receptors. Most notable is the insensitivity of the nematode receptor to sulphonated compounds such as taurine and P4S, which have been shown to exhibit efficacy at the mammalian GABA_A receptor (Kusama et al., 1993; Woodward et al., 1993; del Olmo et al., 2000; Kaji et al., 2015).

The unique pharmacologic features of nematode cys-loop GABA receptors suggest that there are structural differences in the agonist binding pocket. However, compared with their mammalian counterparts, little is known about the structural elements that govern agonist binding to nematode GABA receptors.

The binding site of cys-loop receptors is made up of six discontinuous loops, designated by the letters A to F. The number of loops and their general position appear conserved across phyla (Brejc et al., 2001; Hibbs and Gouaux, 2011). Loops A to C are found within the primary subunit, and loops D to F are in the adjacent subunit. The role of various binding

Research was funded by the Natural Science and Engineering Council of Canada (NSERC) and the Canadian Foundation for Innovation (CFI) to S.G.F. PDB File: Hco-UNC-49B homodimer.

<https://doi.org/10.1124/mol.118.112821>.

[§] This article has supplemental material available at molpharm.aspetjournals.org.

ABBREVIATIONS: cRNA, copy RNA; DAVA, 5-aminovaleric acid; MD, molecular dynamics; MS-222, 3-aminobenzoic acid ethyl ester methane sulphonate salt; MTSET, 2-(trimethylammonium)ethyl methanethiosulfonate; ND96 buffer, [96 mM NaCl, 2 mM KCl, 1.8 mM CaCl₂, 1 mM MgCl₂, 5 mM HEPES]; PS, pregnenolone sulfate; RDL, resistance to dieldrin; SCAM, substituted cysteine accessibility method; UNC, uncoordinated; WT, wild type.

loop residues in the function of mammalian GABA receptors have been examined through site-directed mutagenesis and the substituted cysteine accessibility method (SCAM).

SCAM analysis involves changing the predicted binding site residues to cysteines via site-directed mutagenesis. If the introduced cysteine is modified by a reducing agent such as 2-(trimethylammonium)ethyl methanethiosulfonate (MTSET) (see Fig. 1), GABA binding should be reduced if that cysteine resides in the binding pocket. Binding pocket residues are confirmed through the use of agonists and antagonists, which slow the rate of modification of the introduced cysteines by reducing agents (Kloda and Czajkowski, 2007). However, it is important to note that modification of cysteines by molecules such as MTSET could have more complex effects on receptor activation, so the results should be interpreted with caution.

The detailed knowledge that we now have about the structure of the GABA binding pocket in mammalian GABA receptors has provided an opportunity for comparative analyses on the GABA receptors of more primitive organisms such as nematodes. Several residues in loops A to D have previously been examined in the Hco-UNC-49 receptor through site-directed mutagenesis, which revealed functional similarities when compared with mammalian GABA receptors (Accardi and Forrester, 2011). Binding loop E, on the other hand, is made up of 18 residues; the residues that play key roles in ligand binding have been mapped in both the GABA_A α 1 and the GABA_C ρ 1 subunits via SCAM analysis (Sedelnikova et al., 2005; Kloda and Czajkowski, 2007). We aligned loop E from these two GABA receptor classes with the Hco-UNC-49 receptor, revealing 12 positions that are not completely conserved (Fig. 2).

Interestingly, residues such as H142 and F145 appear to be conserved among nematode UNC-49-like receptors. We wondered whether the unique residues present in loop E of nematode GABA receptors have any functional significance when compared with published reports on mammalian receptors and if they could possibly explain some of the pharmacologic characteristics observed. We conducted an examination of each residue in loop E from the *H. contortus* UNC-49 receptor using site-directed mutagenesis and SCAM, two methods that have been used to characterize the mammalian GABA binding site, to gain some insight into differences between mammalian and nematode GABA receptors. Of the residues we examined experimentally and via molecular dynamic simulations, some appeared to have similarity in function with mammalian receptors, but others could have some functional differences.

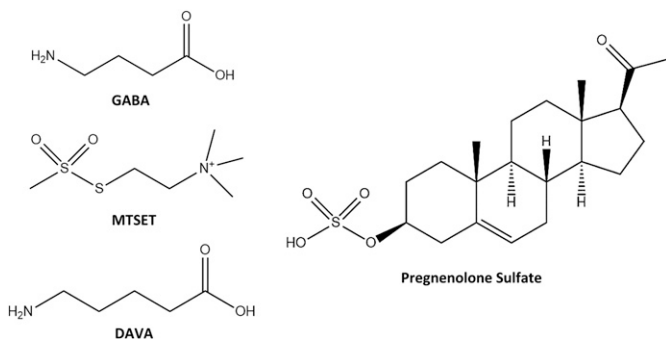


Fig. 1. Chemical structures of GABA, MTSET, DAVA, and PS.

Materials and Methods

Site-Directed Mutagenesis—Primer Design. Amino acid alignments of Hco-UNC-49B and other GABA receptor subunits in the ligand-gated chloride channel (LGCC) family, were used to determine the location of loop E within the *H. contortus* sequence (Fig. 2). The primers were designed using Stratagene's web-based QuikChange Primer Design program (www.stratagene.com/sdm/designer/default.aspx) to create 18 *Hco-unc-49b* (GenBank Accession EU939734.1) mutants: H142C, N143C, S144C, F145C, L146C, R147C, I148C, D149C, S150C, D151C, G152C, T153C, V154C, Y155C, T156C, S157C, Q158C, and R159C. Each pair of mutagenic primers was designed to change each amino acid residue within loop E into cysteine residues by way of nucleotide introduction, resulting in singly mutated cysteine mutants.

Site-Directed Mutagenesis of *Hco-unc-49b*. Previous work in our laboratory found that the binding site for GABA in the Hco-UNC-49BC channel was at the interface of two adjacent UNC-49B subunits. However, the exact stoichiometry of the heteromeric channel is unknown (Accardi and Forrester, 2011). The template used in the site-directed mutagenesis of *Hco-unc-49b* was previously subcloned into a pT7T transcription vector, in which *Xenopus laevis* β -globin untranslated DNA was incorporated into the 5' and 3' ends of the Hco-UNC-49B subunit cDNA (Siddiqui et al., 2010).

A naturally occurring cysteine residue located at position 224 of the template amino acid sequence was removed through site-directed mutagenesis to prevent interference with the results obtained from the introduced cysteine residues. With no alteration in function (Fig. 3;

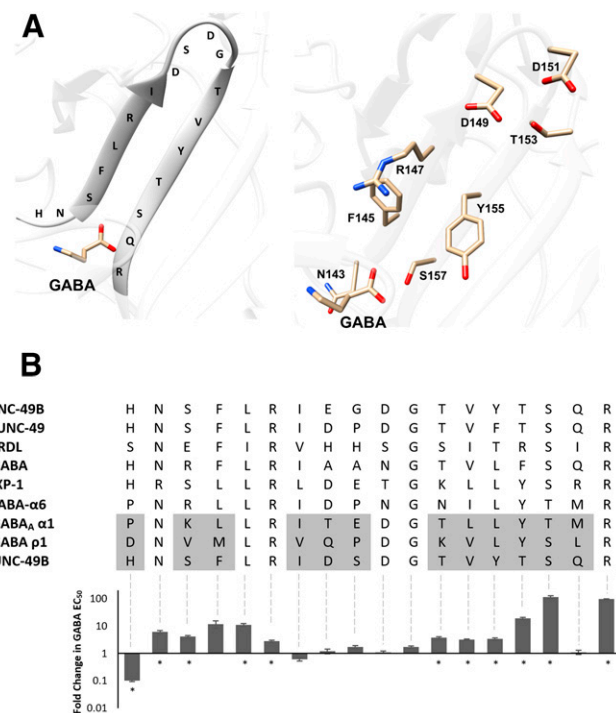


Fig. 2. (A) Model of Hco-UNC-49B homodimer with a docked GABA molecule, highlighting loop E residue positions (left) and the residues that face the binding pocket (right) based on the GluCl crystal structure (PDB 3R1F). (B) Sequence alignment of Hco-UNC-49B loop E residues with the loop E residues from other Cys-loop receptors (*Caenorhabditis elegans* UNC-49, *Brugia malayi* UNC-49, *Drosophila melanogaster* RDL, *Aplysia californica* GABA receptor α -like subunit, *Caenorhabditis elegans* EXP-1, *Ciona intestinalis* GABA α 6-like subunit, human GABA_A, and human GABA_C). Shaded area indicates positions not conserved between Hco-UNC-49B, Hsa-GABA_A α 1, and Hsa-GABA ρ 1. Bar graph below the alignment demonstrates the fold change in GABA EC₅₀ of Hco-UNC-49BC observed when each Hco-UNC-49B loop E amino acid was replaced with a cysteine via site-directed mutagenesis. Each bar represents an average fold change ($n \geq 6$) \pm S.E. The associated EC₅₀ values that were statistically different ($P > 0.001$) from C224A are indicated by an asterisk (*). PBD File: Hco-UNC-49.

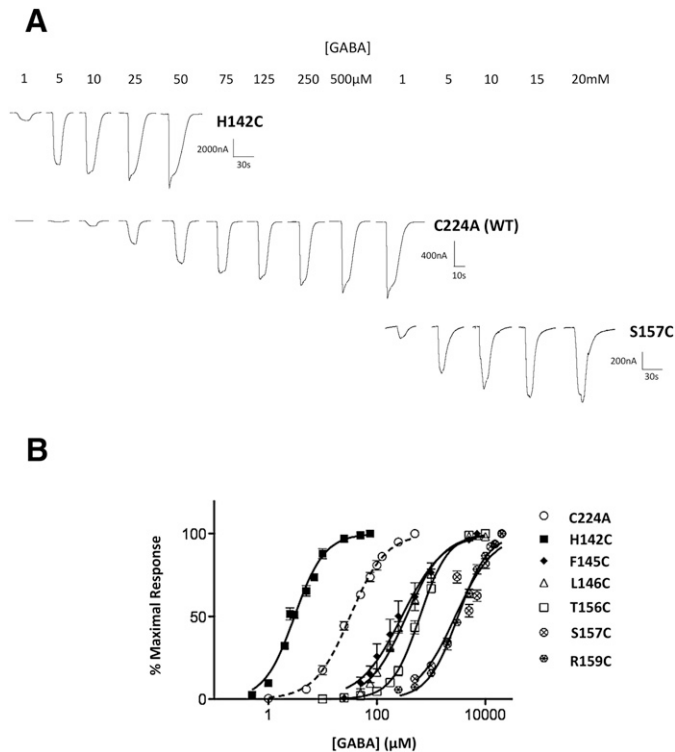


Fig. 3. (A) Representative electrophysiological tracings of Hco-UNC-49BC with mutated Hco-UNC-49B subunits H142C, C224A (cysteine-less mutant used as template for creation of other mutants), and S157C. (B) Dose-response curve of Hco-UNC-49B mutants showing differences in GABA sensitivity, with normalized currents. Each point represents the mean with bars of S.E. ($n > 6$).

Table 1), the cysteine-less mutant (C224A) was used as template and a baseline from which all data obtained could be compared throughout.

All site-directed mutagenesis was conducted using the QuikChange Site-Directed Mutagenesis Kit (Stratagene, La Jolla, CA) according to the manufacturer's instructions. Confirmation that the correct mutation was present was verified using DNA sequencing (Génome Québec, Montreal, Canada).

cRNA Preparation—In Vitro Transcription. Plasmid constructs containing either the mutated *Hco-unc-49b* and wild-type (WT) *Hco-unc-49c* (GenBank Accession EU049602.1) were linearized and used as a template (0.4–1.0 μg) to create capped *Hco-unc-49* copy RNA (cRNA). The cRNA was made using the mMessage mMachine in vitro transcription reaction and the T7 RNA polymerase provided within the transcription kit (Ambion, Austin, TX). The cRNA was precipitated using lithium chloride and was brought to a concentration of 0.5 ng/1 upon resuspension in nuclease free water. Each in vitro transcription reaction yielded approximately 10–25 μg of cRNA.

Expression of Hco-UNC-49BC in *Xenopus laevis* Oocytes. Oocytes were obtained surgically from female *Xenopus laevis* (Nasco, Fort Atkinson, WI), which were housed in temperature-controlled rooms at the University of Ontario Institute of Technology (UOIT) according to the methodology outlined in Abdelmassih et al. (2017) and adhering to the guidelines of the UOIT Animal Care Committee and the Canadian Council of Animal Care. The frogs were anesthetized with 0.15% 3-aminobenzoic acid ethyl ester methane sulphonate salt (MS-222; Sigma-Aldrich, Oakville, ON, Canada). Lobes of the ovary were extracted, split into smaller pieces of 10–20 oocytes, and defolliculated in a treatment of 2 mg/ml collagenase-II (Sigma-Aldrich) and OR2 solution (82 mM NaCl, 2 mM KCl, 1 mM MgCl₂, 5 mM HEPES, pH 7.5) for 2 hours with gentle rocking at room temperature. After defolliculation, the oocytes were stored at 20°C in ND96 buffer (96 mM NaCl, 2 mM KCl, 1.8 mM CaCl₂, 1 mM MgCl₂,

TABLE 1

Comparison of GABA EC₅₀ and Hill slope values of Hco-UNC-49B mutants assembled as heteromeric Hco-UNC-49BC receptors

Fold change values indicate change in GABA EC₅₀ of each mutant normalized to C224A mutant (cysteine-less mutant with wild-type responses, used as template for creation of other loop E mutants).

Mutant	EC ₅₀ ± S.E. (μM) (Hill Coefficient ± S.E.)	<i>n</i>	Fold Change
C224A	32.56 ± 2.29 (1.37 ± 0.11)	7	1
H142C	2.70 ± 0.22 (1.44 ± 0.07) ^a	7	0.1
N143C	199.69 ± 22.86 (1.33 ± 0.11) ^a	7	6.1
S144C	133.76 ± 11.96 (1.31 ± 0.07) ^a	7	4.1
F145C	383.53 ± 114.30 (1.40 ± 0.04)	7	11.8
L146C	361.39 ± 42.59 (1.30 ± 0.04) ^a	7	11.1
R147C	90.32 ± 6.54 (1.82 ± 0.16) ^a	8	2.8
I148C	19.37 ± 2.62 (1.53 ± 0.13)	8	0.6
D149C	39.79 ± 7.11 (2.15 ± 0.27)	7	1.2
S150C	54.39 ± 6.95 (1.78 ± 0.13)	7	1.7
D151C	36.11 ± 3.25 (1.43 ± 0.08)	7	1.1
G152C	54.91 ± 5.01 (1.48 ± 0.06)	7	1.7
T153C	122.21 ± 10.01 (1.71 ± 0.08) ^a	6	3.8
V154C	104.05 ± 5.10 (1.51 ± 0.14) ^a	7	3.2
Y155C	111.49 ± 10.68 (1.64 ± 0.09) ^a	7	3.4
T156C	628.48 ± 50.61 (1.71 ± 0.06) ^a	6	19.3
S157C	3683.50 ± 413.66 (1.43 ± 0.09) ^a	8	113.1
Q158C	35.44 ± 6.63 (1.44 ± 0.10)	7	1.1
R159C	3102.33 ± 130.43 (1.53 ± 0.03) ^a	6	95.3

^a Values statistically significantly different from C224A ($P < 0.001$).

5 mM HEPES, pH 7.5) supplemented with 100 μg/ml gentamycin and 0.275 μg/ml pyruvic acid (Sigma-Aldrich). Supplemented ND96 solution was replaced twice every 24-hour period.

Cytoplasmic injections of cRNA were performed on stage V and VI oocytes using a Drummond Nanoject II (Drummond Scientific Company, Broomhall, PA) assisted by micromanipulators (World Precision Instruments, Sarasota, FL). Each oocyte was injected with 50 nl of a mixture of mutated *Hco-unc-49b* and WT *Hco-unc-49c* (equal amounts of 0.5 ng/nl cRNA of each). Electrophysiological recordings were taken after receptor expression, approximately 2 to 5 days after the cRNA injection.

Compounds Tested and Their Preparation. MTSET was obtained from Toronto Research Chemicals (Toronto, ON, Canada). GABA, 5-aminovaleic acid (DAVA), and pregnenolone sulfate (PS) were obtained from Sigma-Aldrich. All compounds were initially dissolved in ND96 except for PS, which was dissolved in dimethylsulfoxide for the preparation of a stock solution. Compounds were diluted in ND96 as working solutions. The structures of the compounds are found in Fig. 1.

Two-Electrode Voltage Clamp Electrophysiology. Using an Axoclamp900A voltage clamp (Molecular Devices, Sunnyvale, CA), two-electrode voltage clamp electrophysiology was used to observe and record the channel activity of the Hco-UNC-49B/C channel. Glass electrodes were pulled from borosilicate capillaries using a P-97 Flaming/Brown micropipette puller (Sutter Instruments, Novato, CA), and filled with 3 M KCl (1–5 MΩ resistance). Each of the two electrodes were connected to Axon Instrument Headstages (Molecular Devices) using Ag|AgCl wires.

Oocytes were held at a membrane voltage at –60 mV throughout the experiment. Compounds were perfused over the oocytes (at 8 ml/min) through a gravitational flow system into an RC-1Z perfusion chamber (Warner Instruments, Holliston, MA). ND96 buffer was used to dilute the various compounds, in addition to being used as a wash solution in between drug applications. Recordings and associated data were obtained using Axon Instruments Digidata 1440, Clampex 10.2, and were analyzed via Clampfit 10.2 (Molecular Devices).

The EC₅₀ was determined for each mutant by perfusing increasing concentrations of GABA (or other agonists) and recording the current observed which included the concentration of agonist that produced maximal current.

For experiments using the negative allosteric modulator of GABA receptors, PS, oocytes expressing mutant or WT Hco-UNC-49BC

receptors were first exposed to EC₅₀ concentrations of GABA to ensure channel functionality. Oocytes were then exposed to a coapplication of an increasing range (5–500 μM) of PS concentrations mixed with the corresponding EC₅₀ GABA concentration. Changes in current were recorded and inhibitory dose–response curves were generated to determine the IC₅₀ value for the mutant being tested.

Substituted Cysteine Accessibility Method. SCAM was used to characterize all Hco-UNC-49B mutants as well as the cysteine-less mutant (C224A) using the reducing agent MTSET. During testing, an oocyte was washed and hit with EC_{20–50} concentrations of GABA in 5-minute intervals until the response stabilized within 10%. Once stabilized, MTSET (1 mM for 1 minute) was perfused over the oocyte, followed by 5 minutes of washing with ND96 solution. The EC_{20–50} concentrations of GABA for the mutant being tested were reapplied, and currents were recorded (Sedelnikova et al., 2005; Kloda and Czajkowski, 2007).

Statistical Analysis. Dose–response curves were generated by Prism 5.0 (GraphPad Software, San Diego, CA) using the following equation in the set-up log (agonist) versus normalized response-variable slope:

$$I_{\max} = \frac{1}{1 + \left(\frac{EC_{50}}{[D]}\right)^h}$$

where I_{\max} is the maximal response, $[D]$ is the concentration of agonist, EC_{50} is the concentration of agonist that is required to produce half-maximal current, and h is the Hill coefficient. Responses used to produce dose–response curves were normalized as a percentage of the maximal current produced by the oocytes individual maximal response to GABA.

The effect of MTSET modification was calculated using the following formula:

$$\left(\frac{I_{\text{GABA-post}}}{I_{\text{GABA-pre}}}\right) - 1$$

where $I_{\text{GABA-post}}$ is the current produced from GABA activation of the receptor after the application of MTSET, and $I_{\text{GABA-pre}}$ is the current produced from GABA activation before MTSET was applied (Kloda and Czajkowski, 2007).

The EC₅₀ values, h values, and S.E. were determined using Prism 5.0 from a minimum of five oocytes from two different *X. laevis*. Statistical significance was derived from Student's t test using the Bonferroni correction or a one-way analysis of variance with Dunnett's multiple comparison test. $P < 0.001$ was considered statistically significant.

Homology Modeling. The *C. elegans* glutamate-gated chloride channel crystal structure (PDB 3RIF; Hibbs and Gouaux, 2011) was used as a template in MODELER 9.14 (Sali and Blundell, 1993) for the generation of a Hco-UNC-49B extracellular domain homodimer (Data Supplement PDB File). The most energetically favorable models were determined based on their DOPE score and PROCHECK Ramachandran plot analysis, as described in Kaji et al. (2015).

Computational Agonist Docking. The energetically reduced zwitterion form of GABA was obtained from the Zinc database (<http://zinc.docking.org>; Irwin et al., 2012). The GABA molecule was prepared for docking using AutoDock Tools (Morris et al., 2009) and was docked using AutoDock Vina (Trott and Olson, 2010). The center of the 30 × 30 × 30 Å search box located in the aromatic box of the agonist binding site was used for agonist docking. A maximum of 50 binding models, all within a range of 5 kcal mol⁻¹ from the best scoring pose, were generated (Kaji et al., 2015).

Molecular Dynamics Simulations. The homology model of the protein we generated, containing the docked GABA molecule in the binding area, was used as the initial structure for the molecular dynamics (MD) simulations. The GROMACS 2016.4 software package (www.gromacs.org), employing the CHARMM36 force field (Best et al., 2012), was used to run the simulations. The CHARMM General Force

Field (CGenFF) program, which is an automated method to provide parameters and charges by analogy (Vanommeslaeghe et al., 2010, 2012; Vanommeslaeghe and MacKerell, 2012; Yu et al., 2012; CGenFF interface at <https://cgenff.paramchem.org>), was used to acquire the CHARMM force field parameters for the GABA molecule. The simulation box was solvated with ~21,000 tip3p water molecules, along with Cl⁻ counter ions to neutralize the total charge of the system.

To carry out the MD simulation, after a brief energy minimization, a 2 nanosecond equilibration simulation was performed under the NVT (number of particle, volume of the system, and temperature are constant) conditions at 300 K and a time step of 2 femtoseconds. The positions of the protein and the ligand were restrained to prevent any structural changes during the equilibration process.

In the production run, under the NPT (number of particle, pressure of the system, and temperature are constant) conditions, the system was kept at 300 K and 1 atm with a time step of 2 femtoseconds for 500 nanoseconds. The Parrinello-Rahman isotropic pressure coupling (with $\tau_p = 5$ picoseconds and compressibility 4.5×10^{-5} bar⁻¹) and the Nose-Hoover thermostat (with a time constant of 0.1 picoseconds) were employed for these production runs. Long-range electrostatics were calculated with the particle-mesh Ewald method. The trajectory was printed every 10 picoseconds.

The gromacs utilities, such as `gmx-mpi mindist`, `gmx-mpi gyrate`, and `gmx-mpi rms`, as well as the H-bond calculator plugin of VMD (Visual Molecular Dynamics) (Humphrey et al., 1996; <http://www.ks.uiuc.edu/Research/vmd/>), were used to calculate the results. The default values for distance (d) and angle (θ) ($d < 3.0$ Å and $\theta < 20^\circ$) are used to determine the H-bonds. Ionic bonds between oppositely charged residues were determined at $d < 3.0$ Å.

Results

Characterization of Loop E Cysteine Mutants in Hco-UNC-49BC. The importance of loop E amino acid residues in the Hco-UNC49 receptor was first assessed by determining the impact of each cysteine mutation on the GABA activation of the channel. Each loop E mutant was exposed to increasing concentrations of GABA to determine the EC₅₀ values. Upon recording, good expressions of all receptors with mutated subunits were observed, all receptors were functional, and clean tracings were obtained (Fig. 3A). The cysteine-less mutant C224A was an appropriate control for this study as the GABA EC₅₀ (33 μM) was similar to WT (40 μM; Siddiqui et al., 2010), and this mutant was not sensitive to MTSET treatment.

Of the 18 mutants tested, 12 showed a substantial shift in GABA EC₅₀ compared with the C224A cysteine-less mutant from which they were derived (representative dose–response curves are seen in Fig. 3B). Of the mutants that displayed a different EC₅₀ from C224A (33 μM ± 2), H142C showed an increase in GABA sensitivity (2.7 μM ± 0.2). The remaining 11 mutants (N143C, S144C, F145C, L146C, R147C, T153C, V154C, Y155C, T156C, S157C, and R159C) had a decrease in GABA sensitivity, with S157C and R159C showing the greatest decrease (3683.50 μM ± 413.66 and 3102.33 μM ± 130.43, respectively). Finally, six of the cysteine mutants, I148C, D149C, S150C, D151C, G152C, and Q158C, showed minimal change in GABA EC₅₀. The EC₅₀ values that were statistically significantly different from C224A ($P > 0.001$) are shown in Table 1.

Determination of Cysteine Accessibility. SCAM was performed on all loop E Hco-UNC-49B mutants to determine which were accessible to MTSET modification and the impact of modification on channel function. Overall, we found that

most of the cysteine mutants were either not influenced or positively influenced by MTSET treatment. Specifically, out of the 18 mutants tested, only H142C, R147C, and S157C were significantly ($P < 0.001$) affected by MTSET modification (Fig. 4). Furthermore, only S157C had a decrease in the percentage change in I_{GABA} ($-49.8\% \pm 7.8\%$). Conversely, H142C and R147C all showed an increase in the percentage change in I_{GABA} ($88\% \pm 4\%$ and $52.5\% \pm 17\%$, respectively).

Pharmacologic Characterization of H142C Mutant.

Upon discovering significant hypersensitivity to GABA in H142C, we investigated two other compounds: DAVA and PS. Dose–response curves were created for both compounds (Fig. 5, A and C) with respect to the H142C mutant as well as the cysteine-less C224A baseline mutant. In addition, DAVA behaved as a partial agonist at the C224A receptor (I_{max} 21.2% \pm 2.7%), but as a full agonist at the H142C receptor (I_{max} 86.5% \pm 2.8%; Table 2). On the other hand, no significant difference was observed in the IC_{50} of PS between the C224A and H142C receptors (Fig. 5; Table 2). For the C224A receptor, the IC_{50} for PS was 47 μM compared with 70 μM for H142C.

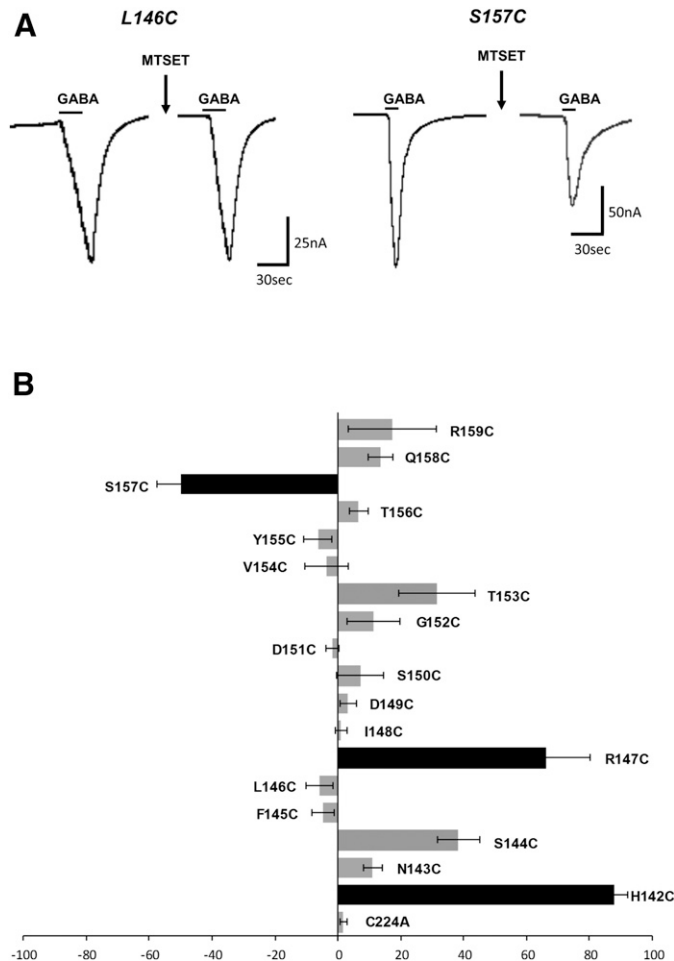


Fig. 4. Effect of MTSET on current evoked from GABA activation of Hco-UNC-49BC receptors with mutated Hco-UNC49-B subunits. (A) Representative tracings of current elicited from GABA (EC_{20-50}) before and after MTSET application (1 mM, 1 minute). (B) Percentage change in current from treatment of mutant Hco-UNC-49BC receptors with MTSET. Each bar indicates the mean \pm S.E. ($n > 3$). Black bars denote statistically significant difference from C224A ($P < 0.001$).

MD Simulations. As a further analysis of loop E residues we conducted molecular simulations. To ensure that the simulations provided a meaningful analysis of the role of loop E residues and their possible affect on GABA binding, we first examined the interaction of GABA with well-studied residues within the binding pocket. During the course of the simulation, the GABA molecule remained inside the initial binding pocket and interacted with key side chains previously implicated as essential for GABA binding in both mammalian (Newell et al., 2004) and the UNC-49 receptor (Accardi and Forrester, 2011; Kaji et al., 2015) (Fig. 6A). For example the positive amino group of GABA forms ionic bonds with carboxylate of E185 (loop B) 94% of the time. Likewise the carboxylate group of GABA forms ionic bonds with R87 (loop D) 99% of the time, and S236 (loop C) forms hydrogen bonds with GABA 35% of the time. These values were similar to MD simulations of the *Drosophila* resistance to dieldrin (RDL) receptor (Ashby et al., 2012).

With respect to loop E residues, the molecular simulations revealed only two residues forming hydrogen bonding with GABA. GABA most frequently formed H-bonds with S157 (74.25% of the time) and TYR155 (53.36% of the time) (Fig. 6, A and D).

ARG147 did not interact with the GABA molecule, except for during the first few nanoseconds of the simulation (Fig. 6B; Fig. 7A). Alternately, the R147 side chain mainly associated with the D149 side chain (Fig. 7A) via an ionic bond (35%

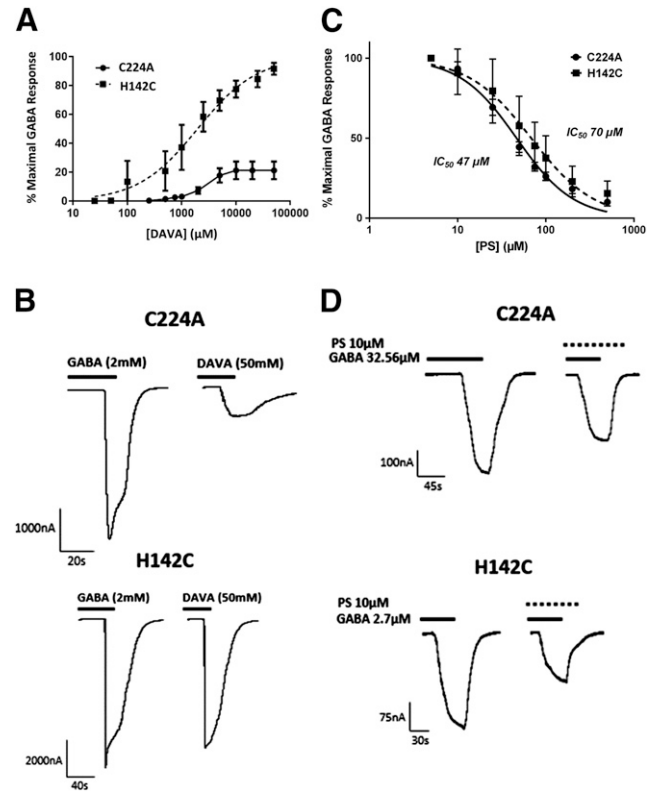


Fig. 5. Effect of loop E H142C mutation in Hco-UNC-49B on Hco-UNC-49BC response to DAVA and PS. (A) DAVA dose–response curve comparing C224A and H142C mutant receptors. (B) Comparison of maximal current produced by GABA and DAVA in C224A and H142C mutant receptors. (C) PS dose–response curve comparing C224A and H142C mutant receptors. (D) Comparison of current inhibited by PS in the presence of GABA (EC_{50}) in C224A and H142C mutant receptors.

TABLE 2

EC₅₀, Hill coefficient, and maximal responses of DAVA and GABA produced by mutant Hco-UNC-49BC receptors H142C and C224A

Compound	H142C			C224A		
	EC ₅₀ ± S.E. (μM) (Hill Coefficient ± S.E.)	% Maximal GABA ± S.E.	<i>n</i>	EC ₅₀ ± S.E. (μM) (Hill Coefficient ± S.E.)	% Maximal GABA ± S.E.	<i>n</i>
DAVA	1353.2 ± 246.2 (1.15 ± 0.16)	86.5 ± 2.8 ^a	7	2464 ± 107.2 (2.18 ± 0.1)	21.2 ± 2.7	5
GABA	2.7 ± 0.22 (1.44 ± 0.07) ^a	100	7	32.56 ± 2.29 (1.37 ± 0.11)	100	7

^a Values statistically significantly different from C224A ($P < 0.001$).

during the course of the simulation). However, the most recurrent interaction between the two residues was a H-bond between the backbone of R147 and Y155 as part of the antiparallel β -sheet secondary structure of loop E (~81% as an acceptor and ~19% as a donor) (Fig. 6B; Fig. 7B).

Electrostatic interactions between H142 and S144 were also observed. However, the interaction was variable, and only 0.39% of the time was the interaction hydrogen bonding. The distance between these two groups fluctuated between ~1 nm (not interacting) and ~0.4 nm (interacting) (Fig. 6C; Fig. 8). For the last 200 nanoseconds of the simulation, the separation distance became more dynamic. Further investigation indicated that there was a small structural change in the structure of loop E at $t \sim 300$ nanoseconds, where the S144 and F145 (at the beginning of the loop) lose their β -structure (Supplemental Fig. 1). However, this change did not have any major impact on the results calculated in the system (Supplemental Fig. 2; Supplemental Table 1).

Discussion

A comparative analysis of residues between mammalian and invertebrate GABA receptors can provide some valuable information on ligand-binding sites and receptor function. Indeed, a comparative mutagenesis approach was key to identifying the neurosteroid binding site in cys-loop GABA receptors (Hosie et al., 2006). Our study describes the characterization of the 18 residues found in binding loop E of the Hco-UNC-49 receptor.

In this investigation we determined which residues, when mutated to cysteine, affected the sensitivity of the Hco-UNC-49 receptor to GABA and other agonists. Moreover, exposure of the introduced cysteines to the reducing agent MTSET revealed which residues were accessible to MTSET modification (e.g., in an aqueous environment) and affected GABA sensitivity. However, the SCAM results should be interpreted with caution because molecules such as MTSET have been shown to have more complex effects, such as acting as tethered partial agonists (Wang et al., 2010). Alternatively the enhancement in I_{GABA} might be caused by MTSET acting as a positive allosteric modulator.

This study revealed some loop E residues within Hco-UNC-49 that may exhibit similar properties as those found in human receptors, but it also revealed several with possible distinct functions in the nematode receptor. Overall, it appears that the loop E of the Hco-UNC-49 receptor is more tolerant to cysteine mutagenesis than the GABA_A receptor (Kloda and Czajkowski, 2007).

Another distinction in the overall results of this study was that only one of the mutations (S157C) resulted in a decrease in I_{GABA} when modified by MTSET whereas others exhibited either no change or an increase in current. Studies on the

human GABA_C (which also used MTSET) receptor appeared to show the opposite trend (Sedelnikova et al., 2005). The results presented here may indicate an overall difference in the structure of loop E and the role it may play in nematode cys-loop GABA receptors when compared with their mammalian counterparts.

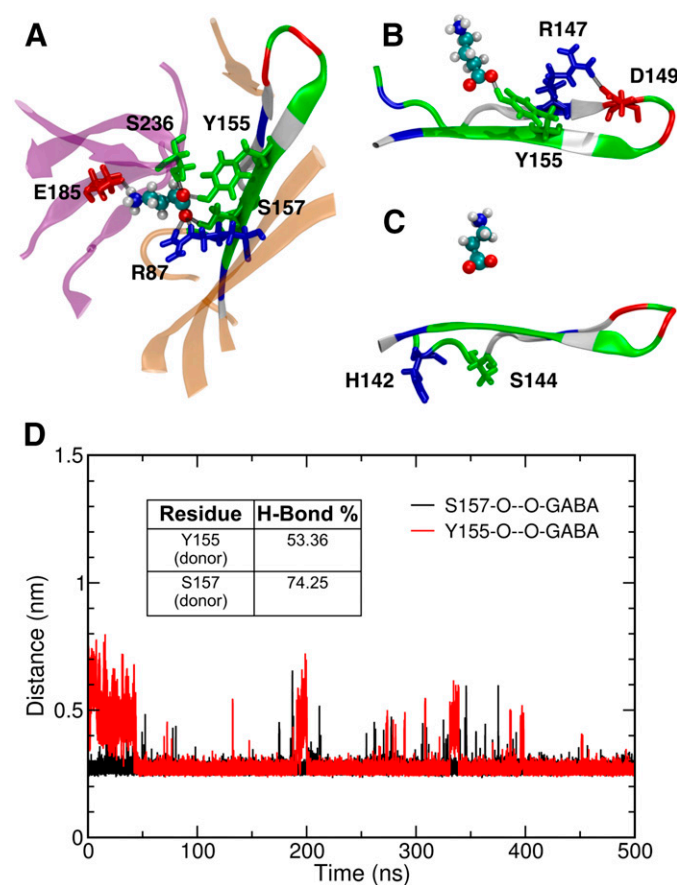


Fig. 6. Molecular simulation snapshots of the Hco-UNC-49B homodimer with GABA docked. E185 is in loop B, S236 is in loop C, R87 is in loop D, and H142, S144, R147, D149, Y155, and S157 are in loop E. (A) GABA interacting with nearby residues within the binding pocket. Gray dashed lines represent the H-bonds. The orange-colored backbone and displayed loop E are part of the same subunit; the purple-colored regions belong to the other subunit of the dimer. (B) Interaction between GABA, Y155, R147, and D149 and (C) H142 interacting with S144. Loop E is presented in residue type colors. The positively charged residues are in blue, the negatively charged residues are in red, the polar residues are in green, and nonpolar residues are in white. (D) Distance analysis between GABA and Y155 and S157 over the course of the simulation. The distances are calculated between the oxygens of the OH group on the side chains of Y155 and S157 and oxygens of GABA. *Inset:* Percentage of hydrogen bonding between residues and GABA. Residue numbering for Hco-UNC-49B corresponds to the first methionine of the amino acid sequence. PDB File: Hco-UNC-49.

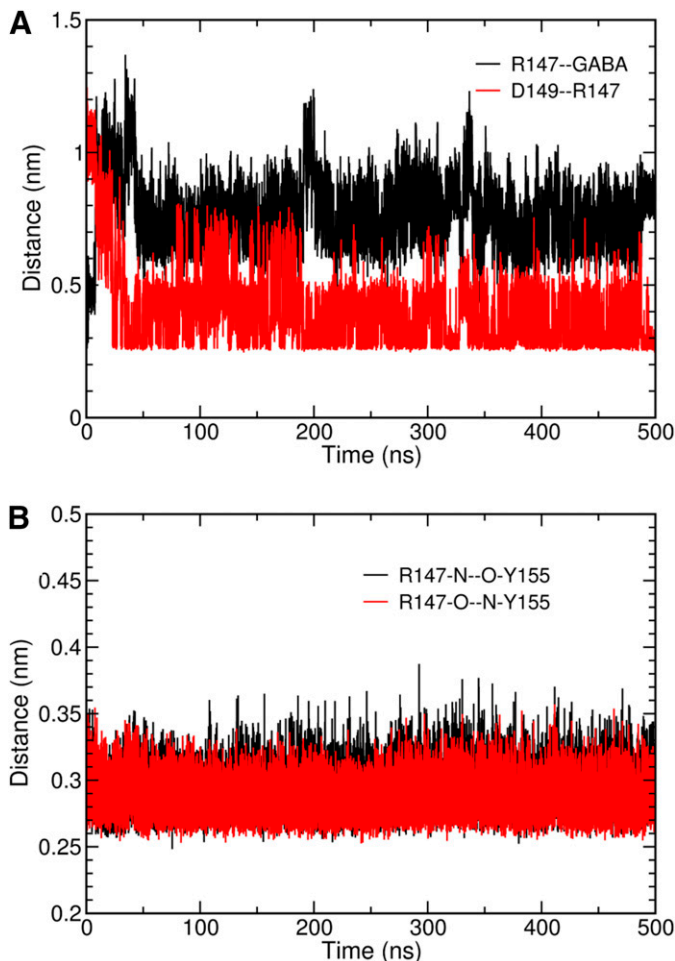


Fig. 7. MD analysis of the interactions between residues and GABA (A) The minimum distance between the three nitrogen atoms of R147 sidechain and the two oxygen atoms of the GABA (black), and the oxygen atom of the D149 side chain and the three nitrogen atoms of the R147 side chain (red). The black line indicates an initial interaction between the R147 and the GABA molecule and separation afterward; the red line indicates an initial separation between the R147 and the D149 and interaction for the rest of the simulation. (B) Minimum distances between the oxygen and nitrogen atoms in the backbone of the R147 and the Y155.

In this study six mutants (I148C, D149C, S150C, D151C, G152C, and Q158C) did not affect the functionality of the receptor, exhibiting EC_{50} values close to those obtained from C224A (the cysteine-less mutant displaying WT function). According to our model (Fig. 2), I148 to G152 are at the top of loop E and therefore positioned $\sim 13\text{--}17$ Å away from GABA. Interestingly, the UNC-49 G152 residue, which is conserved among all GABA receptors, appears to be much more tolerant to a change to cysteine (1.7-fold decrease), compared with mammalian receptors where this change resulted in either a 42-fold reduction in GABA sensitivity or nonfunctional receptors (Sedelnikova et al., 2005; Kloda and Czajkowski, 2007).

R147 appears to be conserved across many phyla (Fig. 2). We found a small 2.7-fold change in EC_{50} when this arginine was mutated to a cysteine; when MTSET was applied, there was a significant increase in GABA current. These results are very different from what was found for the GABA $_A$ $\alpha 1$ and the GABA $_C$ $\rho 1$ receptors. When this arginine residue was mutated

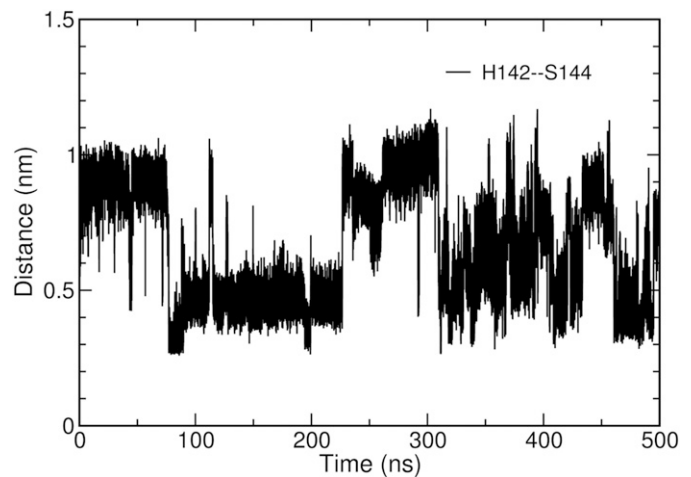


Fig. 8. The minimum distance between the two nitrogen atoms of H142 sidechain and the oxygen atom of the S144 side chain.

to a cysteine in the GABA $_A$ $\alpha 1$ subunit, there was a 260-fold decrease in EC_{50} compared with that of the WT (Kloda and Czajkowski, 2007), and the same mutation in GABA $_C$ $\rho 1$ resulted in a receptor that was expressed too low for further analysis (Sedelnikova et al., 2005). However, in both receptors it was determined that this arginine does not directly interact with GABA.

Homology modeling of the Hco-UNC-49 receptor suggests that R147 faces the binding pocket but does not interact with GABA, based on our MD simulations. However, the positive charge of MTSET (when bound to the introduced cysteine) may stabilize the negative carboxyl group of GABA, which may explain the increase in GABA current we observed after application. In the insect RDL receptor, mutations at this position (i.e., R166) with either an A or G produced non-functional channels. However, it has been suggested that this residue was not likely to be essential for GABA binding (Ashby et al., 2012).

It is possible that R147 has a different role in nematode UNC-49 receptors compared with mammalian and possibly even *Drosophila* receptors. We found evidence for a molecular interaction between the side chains of R147 and D149 (Fig. 7A). In the GABA $_A$ $\alpha 1$ and the GABA $_C$ $\rho 1$ receptors the residues in the analogous positions as D149 are nonionic polar residues (Fig. 2). Therefore, the potential interactions between R147 and other residues may provide some unique features of the nematode GABA receptor.

H142C produced a hypersensitive channel. Treatment with MTSET revealed that the introduced cysteine was accessible for modification, and the resulting modification increased the GABA current. Our MD simulations suggests that H142 does not face the binding pocket, so it may affect the binding site in other ways. Indeed, the mutation H142C not only had a positive effect on GABA, but also affected the Hco-UNC-49 receptor partial agonist DAVA, converting it to a full agonist. This mutation had minimal effect on the sensitivity of the neurosteroid PS, a negative allosteric modulator shown to bind at a site away from the agonist binding site (Wardell et al., 2006). Because histidine is an amino acid with a positively charged bulky side group, it is possible that its removal results in structural changes that may positively

affect both GABA and DAVA binding. One possibility is that H142 has some electrostatic interaction with S144 (Fig. 6C; Fig. 8) and the H142C change alters this interaction, which in turn alters the binding site. It is interesting that H142 appears to be conserved within the nematode phyla and is also present in the unique nematode cation GABA receptor EXP-1 (Beg and Jorgensen, 2003). The exact role of this residue in nematode receptors is unknown, but it could be involved in the unique pharmacologic properties of nematode UNC-49-like GABA receptors.

One residue that may have similar functions across phyla is S157. Mutation of this position to cysteine produces a receptor that was 113-fold less sensitive to GABA; when S157C is modified by MTSET, the GABA current is reduced. The analogous residues in human GABA receptors are T129 in the GABA_A receptor and S168 in the GABA_C receptor. In both cases these residues were confirmed to lie within the binding pocket (Sedelnikova et al., 2005; Kloda and Czajkowski, 2007) where they form an H-bond with GABA. The molecular simulations also revealed high occurrences of H-bonds between S157 and GABA. In the insect RDL receptor it appears that a serine at this position is also essential, where it makes a H-bond with the carboxyl group of GABA (Ashby et al., 2012). However, in the Hco-UNC-49 receptor, MD simulations also detected less frequent H-bonding between Y155 and GABA. The equivalent residue in mammalian receptors and several other GABA receptors is a nonpolar leucine. We anticipate, that in the Hco-UNC-49 receptor, Y155 may play a minor role in GABA binding. Whether it contributes to some unique functions of the nematode UNC-49 receptor will be focus of further study.

Conclusion

This was the first study to conduct a comprehensive analysis of a single binding loop from an invertebrate GABA receptor. We found that, of the loop E mutants we analyzed, H142C, R147C, and S157C had significant changes in EC₅₀ and were accessible to modification by MTSET, which affected GABA sensitivity. Furthermore, as a result of this study, H142 was identified as a residue that may have an important role in the sensitivity and selectivity of the UNC-49 receptor. More work is required to fully characterize the structure of the Hco-UNC-49 binding pocket, but the present study suggests that there are structural differences when compared with mammalian receptors. This further highlights the potential of Hco-UNC-49 as a future drug target.

Acknowledgments

We thank Micah Callanan for assistance with the molecular modeling.

Authorship Contributions

Participated in research design: Kwaka, de Haan, Forrester.
Conducted experiments: Kwaka, Hassan Khatami, Foster, Cochrane, Habibi.
Contributed new reagents or analytic tools: de Haan, Forrester.
Performed data analysis: Kwaka, Hassan Khatami.
Wrote or contributed to the writing of the manuscript: Kwaka, Hassan Khatami, Forrester.

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