**N-Phtharyl-L-Tryptophan (RG108), like Clozapine (CLO), Induces Chromatin Remodeling in Brains of Prenatally Stressed Mice**

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Received June 26, 2018; accepted October 19, 2018

**ABSTRACT**

Schizophrenia (SZ), schizoaffective (SZA), and bipolar (BP) disorder are neurodevelopmental psychopathological conditions related, in part, to genetic load and, in part, to environmentally induced epigenetic dysregulation of chromatin structure and function in neocortical GABAergic, glutamatergic, and monoaminergic neurons. To test the above hypothesis, we targeted our scientific efforts on identifying whether the molecular epigenetic signature of postmortem brains of patients with SZ, SZA, and BP disorder are also present in the brains of adult mice born from dams prenatally restraint stressed (PRS) during gestation. The brains of PRS mice, which are similar to the brains of patients with SZ and BP disorder, show an ~2-fold increased binding of DNMT1 to psychiatric candidate promoters (glutamic acid decarboxylase 67, Reelin, and brain-derived neurotrophic factor), leading to their hypermethylation, reduced expression, as well as the behavioral endophenotypes reminiscent of those observed in the above psychiatric disorders. To establish whether clozapine (CLO) produces its behavioral and molecular action through a causal involvement of DNA methylation/demethylation processes, we compared the epigenetic action of CLO with that of the DNMT1 competitive inhibitor N-phthaly-L-tryptophan (RG108). The intracerebroventricular injection of RG108 (20 nmol/day per 5 days), similar to the systemic administration of CLO, corrects the altered behavioral and molecular endophenotypes that are typical of PRS mice. These results are consistent with an epigenetic etiology underlying the behavioral endophenotypic profile in PRS mice. Further, it suggests that PRS mice may be useful in the preclinical screening of antipsychotic drugs acting to correct altered epigenetic mechanisms.

**Introduction**

The development of new and effective drugs that also show reduced toxicity to treat the complex symptomatology of schizophrenia (SZ), schizoaffective (SZA), and bipolar (BP) disorder is impeded by the lack of pertinent diagnostic tools to assess disease prodromes, progression severity, and beneficial drug responses. Additional fundamental hurdles to the identification of new drugs to effectively treat SZ, SZA, and BP disorder include an insufficient appreciation of the causal and developmental mechanisms underlying the symptoms characteristic of these diseases. Moreover, the inability to reproduce the complex nature of these disorders in laboratory animals is an additional barrier to drug discovery. SZ and related psychiatric disorders are neurodevelopmental disorders with a significant epigenetic pathophysiology brought about by exposure to prenatal or perinatal environmental insults, including stress, drugs, maternal infection, and trauma (Bock et al., 2015; Khashan et al., 2008; van Os et al., 2010; Markham and Koenig, 2011; McGowan et al., 2011; Zhang et al., 2013; Cao-Lei et al., 2015, 2016; Labouesse et al., 2015) and a variable genetic load (Weinberger 1987; Fatemi and Folsom, 2009; Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014; Sekar et al., 2016). Epigenetic studies indicate that aberrant DNA methylation profiles likely play an important neurodevelopmental role in psychiatric disease pathogenesis and as a target for drug discovery (Mill et al., 2008; Ptak and Petronis, 2010; Grayson and Guidotti, 2013). We have previously established that prenatally restraint-stressed (PRS) mice exhibit SZ-like endophenotypic characteristics that can be used to identify robust and efficacious antipsychotic drugs that normalize epigenetic mechanisms (Dong et al., 2016).

The frontal cortices of PRS mice, similar to those of SZ, SZA, and BP postmortem brains, exhibit an epigenetic profile that includes an increase in Dnmt1 binding to the glutamic acid decarboxylase 67 (Gad1), Reelin (Reln), and brain-derived neurotrophic factor (Bdnf) promoters (Matrisiano et al., 2013; Dong et al., 2015b, 2018) and an enrichment of 5-methylcytosine (5mC) and 5-hydroxymethylcytosine at...
these same promoter regions (Dong et al., 2015a, 2016). These increases in promoter methylation are associated with reduced levels of synapse-associated proteins, such as Bdnf and Reln (Dong et al., 2015a, 2016) and behavioral deficits that resemble aspects of the positive and negative symptoms (Jones et al., 2011), and the cognitive impairment, detected in patients with SZ, SZA and BP disorder (Murmu et al., 2006; Lisman et al., 2008; Fine et al., 2014; Ruzicka et al., 2015).

Interestingly, the enrichment of 5mC at the Bdnf, Gad1, and Reln promoters in the medial prefrontal cortex (PFC) and hippocampus of PRS mice correlates with the behavioral endophenotypes (Dong et al., 2015a, 2016). Although there is general agreement that altered epigenetic mechanisms play an important role in the etiopathogenesis of SZ and BP disorder (Guidotti et al., 2011; Grayson and Guidotti, 2013), little is known regarding the effects of drugs, including typical and atypical antipsychotic drugs, on the epigenetic modifications of GABAergic, glutamatergic, or monoaminergic neurons in the neocortices of patients with SZ and BP disorder.

In the current study, we first replicated previous results on the hypermethylation of Bdnf, Reln1, and Gad1 promoters in PRS mice. To validate the PRS mouse as a reasonable epigenetic animal model for studying the actions of specific groups of neuroleptic drugs acting on altered DNA methylation, we compared the effects of a typical antipsychotic drug, haloperidol (HAL) [acting at dopamine receptor D2 (DRD2)], and two atypical antipsychotics: clozapine (CLO) [active at various receptors including 5-hydroxytryptamine receptor (HTR) 1A, HTR2A, HTR2C, and adenosine receptor A1 (ADORA1)] and risperidone (RIS) (active at DRD2, HTR2A, HTR2C, and ADORA1 receptors) (Table 1) (Jarskog et al., 2007; Meltzer, 2012) with the effects of serotonin type 2A (5HT2A), 5HT2C, and adenosine receptor A1 (ADORA1) with various receptors including 5-hydroxytryptamine receptor (HTR) 1A, HTR2A, HTR2C, and adenosine receptor A1 (ADORA1) and risperidone (RIS) (active at DRD2, HTR2A, HTR2C, and ADORA1 receptors) (Table 1) (Jarskog et al., 2007) with the effects of N-phthalyl-L-tryptophan (RG108), a recently identified compound that selectively targets the catalytic domain of DNMT1 (for structure, see the study by Asgatay et al., 2014).

### Materials and Methods

**Animals and PRS Procedure.** All procedures were performed according to guidelines of the National Institutes of Health for animal research and were approved by the Institutional Animal Care Committee of the University of Illinois at Chicago. Timed pregnant mice (Swiss albino ND4) were obtained from Envigo (Indianapolis, IN). Pregnant dams were individually housed with a 12-hour light/dark cycle and food and water available ad libitum. The pregnant mice were left to habituate to the new housing conditions for 1 to 2 days and were then randomly assigned to nonstressed (NS) or stressed groups. NS dams were left undisturbed throughout gestation, whereas stressed dams were subjected to repeated episodes of restraint stress, as described previously (Dong et al., 2016). Experiments were performed on mice between PNDs 60 and 85. Male mice were chosen for the experiments because they exhibit more robust and consistent stereotyped behavior, reduced social interactions, prepulse inhibition, and fear-conditioning deficits than female mice. However, in a recent study (Dong et al., 2018) we also established that female PRS mice show more anxiety than male mice.

**Drug Treatment.** CLO (Sandoz Pharmaceuticals, Princeton, NJ), HAL (Millipore-Sigma, St. Louis, MO), and RIS (Tocris Bioscience, Minneapolis, MN) were dissolved as previously described (Dong et al., 2008). RG108 was dissolved in 10% dimethylsulfoxide (DMSO).

Drugs (CLO 5 mg/kg, HAL 1 mg/kg, and RIS 0.5 and 5 mg/kg) were administered to PRS and NS mice at PND 60–75 subcutaneously twice daily for 5 consecutive days. It is noteworthy that drugs used in this study elicited catalepsy (HAL), and sedation (CLO, RIS) which are maximal after approximately 1 hour but disappear after 18 hours of washout when the animals were resubmitted to the behavioral tests.

<table>
<thead>
<tr>
<th>Drug</th>
<th>Typical Antipsychotic Class</th>
<th>Affinity for Neurotransmitter Receptors&lt;sup&gt;a&lt;/sup&gt;</th>
<th>Treatment Efficacy in PRS Mice (Correction of SZ-Like Behavior)</th>
<th>Promoter Hypomethylation&lt;sup&gt;b&lt;/sup&gt;</th>
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<tr>
<td>CLO 5 mg/kg</td>
<td>Atypical</td>
<td>++++</td>
<td>+</td>
<td>Decreased&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>HAL 1 mg/kg</td>
<td>Typical</td>
<td>++++</td>
<td>++</td>
<td>No effect&lt;sup&gt;d&lt;/sup&gt;</td>
</tr>
<tr>
<td>RIS 0.5 mg/kg</td>
<td>Atypical</td>
<td>++++</td>
<td>+++</td>
<td>No effect&lt;sup&gt;d&lt;/sup&gt;</td>
</tr>
<tr>
<td>RIS 5.0 mg/kg</td>
<td>Atypical</td>
<td>++++</td>
<td>++++</td>
<td>No effect&lt;sup&gt;d&lt;/sup&gt;</td>
</tr>
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</table>

**Affinity for neurotransmitter receptors:** (−) minimal to none; (+) low; (++) moderate; (+++) high; (++++) marked. Data are from Jarskog et al. (2007).

**Treatment Efficacy in PRS Mice:** (−) minimal to none; (+) low; (++) moderate; (+++) high; (++++) marked. Data are from Dong et al. (2018).

<sup>a</sup> Data are from Dong et al. (2018).

<sup>b</sup> Data are from this article.

<sup>c</sup> Data are from Dong et al. (2016).

<sup>d</sup> Data are from Jarskog et al. (2007).
It should be noted that drug doses and duration of treatment were chosen on the basis of preliminary studies showing that CLO (5 mg/kg) requires multiple days of treatment to improve behavioral deficits and elicits demethylation of hypermethylated Reln promoters in mice treated protractedly with large doses of methionine (Dong et al., 2008; Guidotti et al., 2009, 2011; Alachkar et al., 2018). RG108 is a noncompetitive antagonist of DNM1 activity (Asagaya et al., 2014). Because RG108 does not enter the brain at sufficient concentrations due to the blood-brain barrier, it was administered by intracerebroventricular infusion using an osmotic mini pump connected with a brain infusion kit (Alzet; Durect Corporation, Cupertino, CA). The pump delivered a 0.5 µl/h solution containing 100 nmol of RG108 in 10% DMSO in 72 µl for 5 days. Vehicle (VEH; 10% DMSO alone) was administered to corresponding control groups using a similar pump delivery regimen. Behavioral tests were conducted 18 hours after the last administration of RG108, antipsychotic drug, or VEH.

Behavioral Tests. Stereotyped rearing activity and social interactions were examined in succession on the same day between 10:00 AM and 3:00 PM. We selected PNDs 60–75 for behavioral testing because at this postnatal time, the performance of the offspring was more reproducible and stable than the performance measured at earlier developmental time points (Matrisciano et al., 2013). The total number of interruptions of the vertical sensors was taken as a measure of stereotyped behavior. Activities were recorded for 15 minutes.

Social Interaction. Social interaction was measured using the “Thee-Chambers Apparatus” described in detail previously (Dong et al., 2016). Social approaches were defined as the ratio of the sniffing time for the wire cup enclosing the stranger mouse versus that for the empty cup. The reliability of the measurements was assessed by correlating the scores of different raters.

Experimental Design. For these experiments, we used three independent cohorts of mice. In the first cohort, we compared CLO to HAL in six groups of mice: Group 1, NS mice receiving VEH; Group 2, NS mice receiving CLO; Group 3, NS mice receiving HAL; Group 4, PRS mice receiving VEH; Group 5, PRS mice receiving CLO; and Group 6, PRS mice receiving HAL. In the second cohort, we have: Group 1, NS mice receiving VEH; Group 2, NS mice receiving RIS (0.5 mg/kg); Group 3, NS mice receiving RIS (5 mg/kg); Group 4, PRS mice receiving VEH; Group 5, PRS mice receiving RIS (0.5 mg/kg); and Group 6, PRS mice receiving RIS (5 mg/kg). In the third cohort we have: Group 1, NS mice receiving VEH; Group 2, NS mice receiving RG108; Group 3, PRS mice receiving VEH; and Group 4, PRS mice receiving RG108 (100 nmol/day per 5 days i.c.v.). Because these experiments are labor-intensive and time-consuming, we used a different cohort for each drug tested.

For the statistical analyses, to reduce the effect of variability among cohorts, results of CLO, HAL, RIS, and RG108 treatments were compared with the values of the respective controls and were expressed as the percentage of VEH-treated control mice.

PPC Dissection. A Jacobowitz mouse brain slicer matrix was used for the dissection of the cortex. Starting from the frontal lobe (anteroposterior, 4.28 mm), thre slices of 1 mm were mounted on a cold coverslip and disks of 1.5 mm in diameter were punched from the frontoparietal motor and sensory cortices. Using this procedure, we obtained ~20 mg of tissue for DNA/RNA studies (Impagnatiello et al., 1996).

Real-Time Polymerase Chain Reaction. The polymerase chain reaction (PCR) was carried out using an Applied Biosystems Real-Time PCR System (Thermo Fisher Scientific, Foster City, CA) using SYBR Green Master Mix (Fermentas, Glen Burnie, MD). For additional details, see the study by Dong et al. (2016).
Results

CLO, but neither HAL nor RIS, Corrects the Behavioral Endophenotype in PRS Mice. Table 1 reports the high-affinity neurotransmitter receptor profile of three antipsychotic drugs, CLO, HAL, and RIS, each of which was tested for the ability to correct the behavioral endophenotypes and the molecular abnormalities in PRS mice. We previously established that PRS mice exhibit several behavioral alterations, including increased stereotypy, deficits in social interaction time, hyperlocomotor activity in an open field, and deficits in prepulse inhibition of acoustic startle (Matrisciano et al., 2013). The behavioral studies with PRS mice include increased vertical activity (stereotyped behavior) and reduced social interaction. These tests were selected because they produce limited distress for the animals that are then used for subsequent biochemical studies (Dong et al., 2016). For the behavioral experiments, young adult (PND 60–75), NS, and PRS male mice were treated for 5 days with either VEH or the drug under study and then withdrawn for 18 hours to eliminate any acute effects of antipsychotic drugs on behavior. As shown in Fig. 1, we replicated previous reports (Dong et al., 2016) that CLO corrects the behavioral endophenotypes associated with PRS mice, whereas HAL fails to do so. In addition, we observed that RIS (0.5 or 5 mg/kg) was ineffective at reducing the increased stereotyped behavior (Fig. 1A) or relieving the social interaction deficit of PRS mice (Fig. 1B). To establish whether CLO produces its behavioral effects via a causal control of DNMT-mediated promoter methylation, we compared the epigenetic action of CLO with that of the DNMT1 competitive inhibitor RG108 in PRS mice. The intracerebroventricular infusion of RG108 (100 nmol/day for 5 days), similar to CLO (5 mg/kg twice a day for 5 days), corrects the behavioral endophenotypes (Fig. 1).

CLO, but neither HAL nor RIS, Corrects the Increased DNMT1 Binding and 5mC Promoter Hypemethylation of SZ-Associated Genes in PRS Mice. To establish the concept that Bdnf promoter methylation modulates the expression of its mRNA, we tested whether HAL or RIS could correct the local DNA methylation enrichment at cytosines proximal to the transcriptional start site of the Bdnf-ix promoter in the dorsal hippocampus of PRS mice. We observed that the enrichment of cytosine methylation at the Bdnf-ix promoter (Fig. 2) was not reduced in either PRS or NS mice treated with HAL or RIS. In contrast, in a replication of our previous study (Dong et al., 2016) with new cohorts of PRS and NS mice, we showed that CLO corrects both the increased Dnmt1 binding (Fig. 3A) and 5mC enrichment (Fig. 3B) to selected target promoters (Bdnf-ix, Gad1, and Reln). Collectively, these data suggest that CLO, unlike HAL or RIS, may limit DNA methylation, either by reducing the expression of Dnmts, or more likely by interfering with Dnmt1-DNA binding capability (Grayson and Guidotti, 2013, 2016) or Dnmt1 enzymatic activity.

RG108, Like CLO, Corrects the Behavioral Phenotype and Alterations in Target Promoters in the PFC of PRS Mice. We also established that RG108, administered intracerebroventricularly, corrects the methylation enrichment as well as the increased binding of DNMT1 to the Gad1, Reln, and Bdnf-ix promoters in the cortex of PRS mice (compare Figs. 3 and 4). An important observation of this study is that CLO, like valproic acid (VPA) (Dong et al., 2016), has no effect on either the behavior or promoter methylation or the expression of mRNA and protein that we studied when administered to control mice or when injected only one time, 18 hours before testing. A possible explanation for the failure of RIS or HAL to correct these abnormalities is that these drugs are not effective at decreasing DNA methylation.
of CLO to demethylate target promoters in NS mice is that the dynamic state of DNA methylation may be accelerated in PRS mice when compared with the NS mice. Hence, the steady-state methylation process may be very sensitive to short-term inhibition in PRS mice.

The Restorative Action of CLO on Altered Epigenetic Mechanisms Is not Mediated by Histone-Post-Translational Covalent Modifications. The restorative effects of CLO on the molecular and behavioral endophenotypes of PRS mice can be duplicated by the histone deacetylase inhibitor VPA (Tremolizzo et al., 2005; Simonini et al., 2006) directly acting on histone acetylation levels. Of note, VPA increases histone acetylation levels in both NS and PRS mice (Tremolizzo et al., 2005). In addition, there is one report (Huang et al., 2007) that demonstrates an increase in the levels of H3K27me3 in the cortex of mice treated chronically with CLO (5 mg/kg, daily for 21 days). Therefore, we tested whether single (1 hour before) or repeated injections of CLO (5 mg/kg twice a day for 5 days and followed by an 18-hour withdrawal period) alter the levels of H3Kac, H3K4me3, or H3K27me3 in NS mice. In Fig. 5, we show representative images of the Western blot for H3K27me3. The values were normalized to the levels of nonmethylated or nonacetylated H3. No significant differences in the levels of these histone variants were detected in the medial PFC of mice receiving CLO. For example, the levels of H3Kac in the PFC are 110% of control 6 days after the last day treatment. The data are expressed as the mean ± S.D. of five to six mice per group. Two-way ANOVA, group effect for 5mC: Bdnf-IX (F1,17= 318, *P < 0.001). Average value of 5mC levels in three groups of NS mice is about 1% of the input.

Discussion

In this study, we describe a neurodevelopmental, epigenetic mouse model for psychiatric disease (the in utero stressed mouse or PRS mouse) that in the cerebral cortex expresses an increased DNMT1 binding and 5-cytosine hypermethylated of promoters (Reln, Gad1, and Bdnf) that represent the core epigenetic pathophysiological signature underlying SZ, SZA, and BP disorders. This model responds to the administration of CLO with striking behavioral and neuroepigenetic improvements, whereas HAL and RIS fail to do so. The behavioral and molecular effects of CLO are mimicked by the intracerebroventricular injection of the DNMT inhibitor RG108 (Asgatay et al., 2014), suggesting that PRS mice may help to identify novel families of neuroleptic drugs capable of reducing DNA hypermethylation and correcting the behavioral endophenotypes associated with these disorders.

Mice born from mothers stressed during gestation show that, as in humans, environmental insults occurring at critical periods of central nervous system development can trigger epigenetic modifications at psychiatric disorder candidate genes, including Reln, Gad1, and Bdnf, leading to their downregulation (Khashan et al., 2008; van Os et al., 2010; Markham and Koenig 2011; McGowan et al., 2011; Zhang et al., 2013; Cao-Lei et al., 2015, 2016; Scheinost et al., 2017). These epigenetic changes are paralleled by behavioral deficits that are phenotypically reminiscent of those observed in psychiatric diseases (Dong et al., 2016). Hence, the PRS mouse model has value as an experimental epigenetic model for SZ,
Tukey of five animals per group and analyzed with two-way ANOVA followed by NS mice receiving RG108. The data are expressed as the mean reversed by treatment with RG108. Note that there is no change of 5mC in Reln, and Gad1 promoter regions in the frontal cortex of PRS mice are Fig. 4.

PRS mice. We confirmed previous results that doses of HAL molecular (chromatin remodeling) alterations observed in one typical (HAL) antipsychotic drug on the behavioral and molecular neuroepigenetic alterations induced by typical and atypical neuroleptic drugs (Jarskog et al., 2007), it should be possible to identify or exclude the contribution of each receptor or combination of receptors and their corresponding signal transduction mechanisms to the regulation of altered chromatin remodeling events in PRS mice.

For example, we compared two atypical (CLO and RIS) and atypical neuroleptic drugs (Jarskog et al., 2007), it should be possible to identify or exclude the contribution of each receptor or combination of receptors and their corresponding signal transduction mechanisms to the regulation of altered chromatin remodeling events in PRS mice. We confirmed previous results that doses of HAL that are capable of blocking DRD2 function and inducing extrapyramidal side effects but do not have serotonin receptor affinity, fail to normalize the altered behavioral endophenotypes or to reduce promoter hypermethylation in PRS mice (Table 1). In contrast, CLO, which binds with high affinity to serotonin receptors but with low affinity to DRD2 (Table 1), normalizes the altered endophenotypes (Fig. 1) and aberrant chromatin remodeling (Figs. 2 and 3). These data suggest that the effects of CLO are not mediated solely by the blockade of DRD2 function.

To further clarify this issue, we studied RIS (an antipsychotic with high affinity for both DRD2 and HTR1A, HTR2A, HTR2C, and ADORA1; see Table 1). This atypical antipsychotic drug failed to induce the restorative behavioral and molecular effects reported with CLO (Figs. 1 and 2). The lack of effects of HAL or RIS on the behavioral and molecular neuroepigenetic endophenotypes of PRS mice support the concept that DRD2 as well HTR1A, HTR2A, HTR2C, and ADORA1 are not necessary to explain the corrective action of CLO in our PRS mouse model. However, we cannot rule out a role for CHM1 receptors at this point. The question of whether CLO modulates DNA methylation dynamics bypassing specific neurotransmitter receptors by directly interfering with the binding of DNMT1 to candidate SZ gene promoters or by activating DNA demethylation processes also remains unresolved.

To establish whether the restorative effects of CLO on altered Dnmt1 binding and DNA methylation processes are independent from their action on neurotransmitter receptors and instead dependent on a direct action on DNA methylating and demethylating enzymes and cofactors, we treated mice with RG108, a competitive Dnmt1 antagonist with no known neurotransmitter receptor agonist or antagonist affinity (Asgatay et al., 2014). This drug reduces the increased DNMT1 binding to the Bdnf, Reln, and Gad1 promoters, and also reduces the enrichment of 5mC to these promoters and restores the behavioral deficits in PRS mice. To explain the behavioral and biochemical action of CLO, we studied whether CLO, like VPA, could alter histone post-transcriptional modifications. In addition, there has been at least one report

Fig. 4. The increased DNMT1 binding (A) and levels of 5mC (B) at Bdnf-ix, Reln, and Gad1 promoter regions in the frontal cortex of PRS mice are reversed by treatment with RG108. Note that there is no change of 5mC in the NS mice receiving RG108. The data are expressed as the mean ± S.D. of five animals per group and analyzed with two-way ANOVA followed by Tukey’s multiple comparisons. (A) DNMT1 binding at Bdnf-ix (F1,16 = 11.39 P = 0.004), Reln (F1,16 = 6.15, P = 0.02), and Gad1 (F1,16 = 9.26, P = 0.008). *P < 0.05 when PRS mice are compared with the respective NS VEH-treated mice. #P < 0.001 when treated PRS mice are compared with PRS mice receiving VEH. (B) 5mC enrichment at Bdnf-ix (F1,16 = 9.75 P = 0.007), Reln (F1,16 = 6.75, P = 0.02), and Gad1 (F1,16 = 15.94, P = 0.001). *P < 0.05 when PRS mice are compared with the respective NS VEH-treated mice. #P < 0.05 when treated PRS mice are compared with PRS mice receiving VEH.

SZA, and BP disorder and can be useful for screening potential new anti-SZ/SZA/BDI disorder drugs for preclinical efficacy and perhaps prophylactic intervention.

Despite the large numbers of preclinical and clinical efforts aimed at delineating the role of major neurotransmitter receptors in the anti-SZ profile of different classes of antipsychotic drugs, little is known regarding the role that these receptors play in the regulation of the neurodevelopmental and psychiatric disorders. Here, we show that by investigating the behavioral and molecular neuroepigenetic alterations induced by typical and atypical neuroleptic drugs (Jarskog et al., 2007), it should be possible to identify or exclude the contribution of each receptor or combination of receptors and their corresponding signal transduction mechanisms to the regulation of altered chromatin remodeling events in PRS mice.


Dong, E., Ruzicka, W.B., Grayson, D.R., and Guidotti, A. (2015b) DNA-methyltransferase1 (DNMT1) binding to CpG-rich GABAergic and BDNF promoters is increased in the brain of schizophrenia and bipolar disorder patients. Schizophr Res 167:35–41.


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