

MOL #034249

**Inhibition of vascular endothelial growth factor cotranslational translocation by the  
cyclopeptolide CAM741**

**Hanna Harant, Barbara Wolff, Erwin P. Schreiner, Berndt Oberhauser, Lotte Hofer,  
Nicole Lettner, Sabine Maier, Jan E. deVries and Ivan J.D. Lindley**

Novartis Institutes for BioMedical Research, Brunner Strasse 59, A-1235 Vienna, Austria

Running title: Inhibition of VEGF cotranslational translocation

Correspondence to:

Hanna Harant  
Novartis Institutes for BioMedical Research  
Brunner Strasse 59  
A-1235 Vienna  
Austria  
Tel: (+43)1-80166-330  
Fax: (+43)1-80166-727  
[Hanna.Harant@novartis.com](mailto:Hanna.Harant@novartis.com)

number of text pages: 19

number of Tables: 3

number of Figures: 6

number of References: 33

number of words in Abstract: 163

number of words on Introduction: 569

number of words in Discussion: 1162

Abbreviations: BMH: bis-maleimido-hexane; ER: endoplasmic reticulum; ICAM1: intercellular adhesion molecule 1; MBS: m-maleimidobenzoyl-N-hydroxysuccinimide ester; NC: nascent chain; SEAP: secreted alkaline phosphatase; SP: signal peptide; TGF- $\alpha$ : transforming growth factor- $\alpha$ ; VCAM1: vascular cell adhesion molecule 1; VEGF: vascular endothelial growth factor;

### **Abstract**

The cyclopeptolide CAM741 inhibits cotranslational translocation of vascular cell adhesion molecule 1 (VCAM1), which is dependent on its signal peptide. We now describe the identification of the signal peptide of vascular endothelial growth factor (VEGF) as the second target of CAM741. The mechanism by which the compound inhibits translocation of VEGF is very similar or identical to that of VCAM1, although the signal peptides do not share any obvious sequence similarities. By mutagenesis of the VEGF signal peptide, two important regions, located in the N-terminal and the hydrophobic segments, were identified as critical for compound sensitivity. CAM741 alters positioning of the VEGF signal peptide at the translocon, and increasing hydrophobicity in the h-region reduces compound sensitivity and causes a different, possibly more efficient, interaction with the translocon. Although CAM741 is effective against translocation of both VEGF and VCAM1, the derivative NFI028 is able to inhibit only VCAM1, suggesting that chemical derivatization can alter not only potency, but also the specificity of the compounds.

## **Introduction**

We have recently reported that the cyclopeptolide CAM741, a derivative of the naturally occurring substance Hun-7293, inhibits cotranslational translocation of vascular cell adhesion molecule 1 (VCAM1), which is dependent on its signal peptide (SP) and occurs at the level of VCAM1 SP insertion in the Sec61 translocon (Besemer *et al.*, 2005, Harant *et al.*, 2006). Very similar observations were reported by Garrison *et al.* using cotransin, a compound with related structure (Garrison *et al.*, 2005). These findings demonstrated for the first time that a compound can interfere with the process of cotranslational translocation in a SP-dependent manner.

Amino-terminal, cleavable SPs, when emerging from the ribosome, are recognized by the signal recognition particle (SRP) which then directs the ribosome-nascent polypeptide chain complex to the heterotrimeric Sec61 (composed of the  $\alpha$ -,  $\beta$ - and  $\gamma$  subunits) complex embedded in the membrane of the endoplasmic reticulum (ER) (reviewed by Rapoport *et al.*, 1996; Hegde and Lingappa, 1997; Matlack *et al.*, 1998; Johnson and van Waes, 1999; Stroud and Walter, 1999; Osborne *et al.*, 2005). SPs, usually of 20 to 30 amino-acid residues in length, have a typical three-domain structure, representing a frequently positively charged n-region, a hydrophobic core region (h-region) and a more polar c-region containing the site for SP cleavage (Nielsen *et al.*, 1997; Nielsen *et al.*, 1999). It has been assumed that all SPs interact with the translocon in a similar manner, but there is now increasing evidence that there are remarkable differences between individual SPs (reviewed by Hegde and Bernstein, 2006). It has been shown that some SPs require interaction with accessory translocon components such as translocating chain-associated membrane protein (TRAM) or translocon-associated protein (TRAP), while others do not (High *et al.*, 1993; Mothes *et al.*, 1994; Voigt *et al.*, 1996; Fons *et al.*, 2003). SPs also

contain information on the timing of SP cleavage and subsequent N-glycosylation of the translocated chains (Rutkowski *et al.*, 2001; Rutkowski *et al.*, 2003). During our own studies on the VCAM1 SP, we observed that mutations of the CAM741-sensitive region of the VCAM1 SP caused different association with the translocon components Sec61 $\alpha$  and  $\beta$ , indicating that these mutants are positioned differently within the translocon channel (Harant *et al.*, 2006).

Based on our findings that the cyclopeptolide CAM741 can inhibit the process of cotranslational translocation in a SP-dependent manner, we performed a search for other sensitive SPs to gain more insight into their functionalities. Here we report the identification of the SP of vascular endothelial growth factor-A (here referred as to VEGF) as the second target of CAM741 action.

VEGF, also termed vascular permeability factor (VPF), is one of the key factors in angiogenesis, and induces endothelial cell proliferation, migration, and tube formation (Leung *et al.*, 1989; Keck *et al.*, 1989). Apart from its essential role in neovascularization, VEGF is also involved in several pathological conditions, such as tumor angiogenesis, diabetic retinopathy, age-related macular degeneration, and psoriasis, making it an attractive target for therapeutic intervention (reviewed by Roy *et al.*, 2006; Cardones and Banez, 2006; Eichler *et al.*, 2006). There exist six isoforms of VEGF, differing in their lengths (121, 145, 165, 183, 189 and 206 amino-acid residues). These isoforms are generated by alternative mRNA splicing, differ in sequences encoded by exons 6 and 7, and are differentially expressed between different cells types (reviewed by Robinson and Stringer, 2001). However, all six VEGF isoforms are controlled by the same 26 amino acid-residue signal peptide (SP).

## **Materials and Methods**

*Materials.* CAM741 and NFI028 were dissolved in DMSO and stored at -20°C. Dulbecco's Modified Eagle Medium (DMEM) was purchased from Invitrogen (Carlsbad, CA), and fetal calf serum (FCS) from Bio-Whittaker (Brussels, Belgium). VEGF<sub>165</sub> ELISA was purchased from R&D Systems (Minneapolis, MN). Superfect was purchased from Qiagen (Hilden, Germany). AttoPhos reagent, rabbit reticulocyte lysate, canine pancreatic microsomal membranes, and RiboMAX<sup>TM</sup> Large Scale RNA production System-T7 were purchased from Promega (Madison, WI). Polyclonal Sec61 $\alpha$  or Sec61 $\beta$  antisera were purchased from Upstate (Millipore; Billerica, MA). Excel Gel<sup>TM</sup> SDS 8-18%, Excel Gel<sup>TM</sup> SDS 12-14% gels and [<sup>35</sup>S]-methionine Redivue were purchased from Amersham (Buckinghamshire, UK). TGF- $\alpha$  was purchased from Becton Dickinson (Bedford, MA).

### *Methods.*

*ELISA.* HaCaT keratinocytes were cultured in DMEM supplemented with 10% heat-inactivated FCS. Cells were seeded into 96-well-plates at a density of 1.5 - 2x10<sup>4</sup> cells/well, grown to confluency, and then incubated with increasing concentrations of CAM741 for 16 hrs. VEGF production was stimulated by addition of 50 ng/ml TGF- $\alpha$  for 24 hours. Supernatants were collected and analysed for VEGF<sub>165</sub> by ELISA.

*Plasmid constructions.* The SP-secreted alkaline phosphatase (SEAP) fusion constructs, VEGF SP mutants fused to the SEAP mature domain, the construct encoding a fusion of the N-terminal tag to the VEGF SP, and the truncated VEGF cDNAs encoding either 81 or 131 amino-acid residues were generated by PCR and subcloned into pcDNA3.1 (Invitrogen, Carlsbad, CA) . All

constructs were confirmed by sequencing. The numbering of amino-acid residues refers to VEGF plus SP.

Truncated cDNAs lacking a stop codon were generated by restriction digestion of the respective plasmid DNAs. In case of the SEAP fusion constructs, plasmids were linearized by digestion with either BamH I (encoding 54 amino-acid residues SEAP mature domain) or BstE II (encoding 146 amino-acid residues SEAP mature domain). Linearized plasmids were used as templates for creation of RNAs by the RiboMAX™ Large Scale RNA production System-T7.

*Transient transfections of HEK293 cells.* HEK293 cells were cultivated in DMEM supplemented with 10% FCS and passaged twice a week. For transfection,  $1.5 \times 10^4$  cells in a volume of 100  $\mu$ l were seeded per well of a 96-well plate, transfected with 0.2  $\mu$ g/well plasmid DNA and 0.5  $\mu$ l/well Superfect and treated with increasing concentrations of CAM741 or NFI028. Supernatants were harvested after 24 hrs, and analysed for SEAP secretion using the AttoPhos reagent. Fluorescence was recorded using the SPEKTRamax GEMINI XS (Molecular Devices).

*In vitro translocation experiments.* In vitro translation, targeting and translocation assays, and chemical crosslinking were performed with truncated RNAs using rabbit reticulocyte lysate, canine pancreatic microsomal membranes (Promega) and [ $^{35}$ S]-methionine (Amersham), in the presence of CAM741 or DMSO (vehicle control) as described earlier (Besemer *et al.*, 2005; Harant *et al.*, 2006). Immunoprecipitations were performed with a polyclonal Sec61 $\alpha$  or Sec61 $\beta$  antiserum. Proteins were separated on Excel Gel™ SDS 8-18% or, where stated, on high resolution Excel Gel™ SDS 12-14% gels. Fixed and dried gels were exposed to X-ray films.

## Results

*CAM741 inhibits cotranslational translocation of VEGF which is dependent on its SP.*

To search for SPs which, apart from the known VCAM1 SP (Besemer *et al.*, 2005), could be sensitive to inhibition of translocation by CAM741, a panel of ten different SPs fused to the mature region of secreted alkaline phosphatase (SEAP) was tested as described (Table 1). The SPs were selected as representatives of different classes of secreted or membrane proteins involved in inflammation, immune regulation and angiogenesis. The chemokines, CCL22 (macrophage-derived chemokine; MDC), CCL2 (monocyte chemoattractant protein; MCP-1) and CXCL8 (interleukin-8) were included. The inflammatory cytokines interferon- $\gamma$  (IFN- $\gamma$ ), interleukin-12p40 subunit (IL-12p40) and interleukin-13 (IL-13) were also used, as was VEGF. From the membrane proteins, the SP of chemokine receptor CCR7 (chemokine (C-C motif) receptor 7), E-selectin and intercellular adhesion molecule-1 (ICAM-1) were chosen. While seven of the SP-SEAP constructs showed only partial inhibition of SEAP release at the highest concentration of CAM741 (10  $\mu$ M), SEAP fusion constructs of CXCL8 and the ICAM1 SPs showed some sensitivity to inhibition by CAM741 (Table 1). However, from all SPs tested, that of VEGF was identified as most sensitive to inhibition by CAM741, being only 4-fold less sensitive than the VCAM1 SP (Harant *et al.*, 2006) (Table 1).

To determine that inhibition of SEAP release of the transfected VEGF SP fusion construct by CAM741 also occurs at the level of cotranslational translocation, truncated RNAs encoding the VEGF SP fused to 146 amino-acid residues of the SEAP mature domain were used for *in vitro* translocation experiments. The SEAP mature domain contains a glycosylation site at position 122, and glycosylation and protection from exogenous proteases therefore indicate translocation



to the ER lumen after release of the nascent chains (NCs) from the ribosome by high salt/puromycin. In vitro translocation of truncated VEGF SP-SEAP NCs produced two glycosylated fragments, which were protected from protease digestion (Figure 1A, left panel). Glycosylation was confirmed by treatment with endoglycosidase F (Figure 1A, right panel). The slower migrating glycosylated fragment represents the VEGF SP-SEAP construct with the SP still attached to it, while the faster migrating glycosylated fragment represents the SEAP mature region after SP cleavage. The fastest migrating band, representing unprocessed NCs was almost completely degraded by proteinase K treatment (Figure 1A, left panel). However, in the presence of 1  $\mu$ M CAM741, formation of both glycosylated fragments was inhibited, and again the remaining, unprocessed fragment was degraded by added protease (Figure 1A, left panel).

To exclude any effect of the SEAP fusion partner on the VEGF SP, we also generated 131 amino-acid residues translocation intermediates from wild-type (wt) VEGF<sub>165</sub> which contains an N-glycosylation site at position 101 (position 75 of mature VEGF) (Figure 1B, middle panel). In the control reaction, a slower migrating, glycosylated fragment was formed only in the presence of microsomal membranes, which was resistant to degradation by proteinase K. This fragment could be degraded only after permeabilisation of the membranes with Triton X-100, while the residual unglycosylated product was almost completely degraded also in the absence of detergent. However, in the presence of CAM741, only the unglycosylated fragment formed which was largely degraded by proteinase K (Figure 1B, right panel).

Taken together, while CAM741 does not prevent formation of the tight, salt-resistant binding of VEGF NCs to the translocon, their translocation to the ER is prevented by the compound, witnessed by lack of signal peptide cleavage and glycosylation and their sensitivity to degradation by exogenous protease.

Finally, the effect of CAM741 on release of endogenously expressed VEGF was analysed. HaCaT keratinocytes upregulate the splice variant VEGF<sub>165</sub> in response to treatment with transforming growth factor- $\alpha$  (TGF- $\alpha$ ) (Gille *et al.*, 1998). However, preincubation of HaCaT cells with increasing concentrations of CAM741 dose-dependently inhibited TGF- $\alpha$ -induced VEGF<sub>165</sub> release, as determined by ELISA (Figure 2).

#### *Mapping of the CAM741-sensitive region of the VEGF SP.*

Although the VEGF SP is sensitive to translocation inhibition by CAM741, it shares no similarities with the highly sensitive VCAM1 SP within the primary amino-acid sequence apart from an identical cleavage site for the signal peptidase complex (Table 3). However, the cleavage site of the VCAM1 SP could be replaced without loss in sensitivity, indicating that it does not contain key residues required for the compound effect (Harant *et al.*, 2006). To identify the region critical for inhibition by CAM741, mutagenesis of the VEGF SP was performed. Mutations at different positions of the VEGF SP caused changes in compound sensitivity, with two regions, the n-region and parts of the h-region, recognized as being essential for compound sensitivity (Table 2). The contribution of the n-region to compound sensitivity was identified by removal of the N-terminal amino-acid residues 2-5 or by changing leucines at positions 4 and 5 into glutamine residues, as both mutants required higher concentrations of CAM741 for inhibition (Table 2). However, although these results strongly indicate an involvement of the n-region in compound sensitivity, it is dispensable for successful translocation, as SEAP secretion was not affected in these less sensitive mutants.

Two residues within the central hydrophobic h-region were identified as critical for CAM741 sensitivity, leucine 12 and alanine 13. Substitutions of these residues by different aliphatic residues generated VEGF SP mutants with either increased or decreased compound sensitivities. Replacing leucine 12 by valine resulted in a mild decrease in sensitivity while a more pronounced decrease was observed with isoleucine at this position. When alanine 13 was changed to valine, leucine or isoleucine, again a slight decrease was observed for valine, but a clear reduction in sensitivity for leucine or isoleucine at this position. However, changing both Leu12 and Ala13 by valine or isoleucine, sensitivity to CAM741 was further decreased (Table 2).

Conversely, reducing hydrophobicity at these positions, by replacement of leucine 12 by glycine, caused enhancement in sensitivity, as did the conversion into alanine. However, substitution of both Leu12 and Ala13 by glycine residues resulted in a hypersensitive VEGF SP variant, which was inhibited by CAM741 at low nanomolar concentrations (Table 2). Taken together, the sensitivity to CAM741 can be modulated by either increasing or decreasing hydrophobicity and/or size of the aliphatic residues at positions 12 and 13.

Changing of other residues in the h-region, such as leucines 14 and 15 by either alanine or glycine did not affect sensitivity to CAM741. However, conversion of leucines at position 16 and 18 into valines also caused reduction in sensitivity (Table 2).

Apart from hydrophobicity, a specific presentation of residues of the h-region may be required for the inhibitory effect of CAM741. Proline, a residue with known helix-breaking potential, was

introduced at two different positions within the h-region. When alanine 13 was converted to proline, sensitivity to CAM741 markedly increased. Also changing tyrosine at position 17, a polar residue between leucines 16 and 18, into proline (Y17P) clearly increased sensitivity to CAM741. The results from both mutants would suggest that a specific optimal conformational presentation of residues of the h-region mediates sensitivity to CAM741, which additionally involves the n-region. We therefore tested the effect of removal of the N-terminal residues 2-5 in the highly sensitive VEGF SP mutant Y17P and show that in the absence of these residues sensitivity was clearly reduced (Table 2). These data show that both the n- and h-regions mediate compound sensitivity, and maximal inhibition by CAM741 requires an interplay between these two segments.

The response of selected VEGF SP mutants to inhibition by CAM741 at the level of cotranslational translocation was analysed by in vitro translocation assays of truncated VEGF SP-SEAP NCs (all fused to 146 amino-acid residues SEAP mature domain) in the presence of increasing concentrations of CAM741. The results from the transient transfections were also reflected by these experiments (Figure 3).

*Altered orientation of the VEGF NCs relative to the translocon component Sec61 $\beta$  by CAM741.*

Targeting of the VCAM1 NCs to the translocon is not prevented by CAM741, but translocation inhibition occurs at the step of VCAM1 SP insertion into the translocon (Besemer *et al.*, 2005; Harant *et al.*, 2006). As the VEGF NCs could also be sedimented with the microsomal membranes after high salt/puromycin treatment in the presence of CAM741, we analysed targeted VEGF NCs at the translocon by chemical crosslinking experiments.

When using the heterobifunctional crosslinker m-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS), targeted VCAM1 NCs can be crosslinked to Sec61 $\alpha$  both in the absence or presence of CAM741. However, chemical crosslinking with the homobifunctional cysteine-reactive crosslinker bis-maleimido-hexane (BMH), only in the presence of compound a clearly enhanced crosslink to Sec61 $\beta$  was observed, suggesting an altered orientation of the VCAM1 NCs towards this translocon component. This observation was also shared with Garrison et al. using the structurally related compound cotransin (Besemer *et al.*, 2005; Garrison *et al.*, 2005). We now employed the same chemical crosslinkers to study the translocon environment of targeted 81 amino-acid residues VEGF NCs. When subjected to crosslinking with MBS followed by immunoprecipitation with a Sec61 $\alpha$  antiserum, crosslinks to Sec61 $\alpha$  were observed both in the absence or presence of CAM741 (Figure 4A, left panels, labeled MBS). Chemical crosslinking was then performed with BMH followed by immunoprecipitation with a Sec61 $\beta$  antiserum. When using this crosslinker, visible crosslinks can only form between the cysteine residues present in the mature region of VEGF (positions 52 and 77) and the single cysteine residue in the cytosolic tail of Sec61 $\beta$ . However, only cysteine at position 52 is likely to be accessible to crosslinker, as cysteine at position 77 is too close to the peptidyl transferase site. Although cysteine 52 may also still be located within the ribosome, its distance from the peptidyl transferase site is sufficient for chemical crosslinking to the cytoplasmic tail of Sec61 $\beta$ , as reported for opsin NCs (Laird and High, 1997). In the presence of CAM741, an enhanced crosslink between the VEGF NCs and Sec61 $\beta$  was observed, although there were already basal crosslinks to Sec61 $\beta$  seen in the absence of compound, and their enhancement by CAM741 was less pronounced (Figure 4A, right panels, labeled BMH) compared to targeted VCAM1 NCs

(Besemer *et al.*, 2005). The identity of other crosslinked products which formed differently between control and CAM741-treated reactions is currently unknown (Figure 4A, indicated by open arrowheads), with similar sized products also formed with targeted VCAM1 NCs (Besemer *et al.*, 2005).

*CAM741 differentially alters positioning of VEGF SP mutants at the translocon.*

In the presence of CAM741, not only the VCAM1 mature region but also the VCAM1 SP has an altered orientation relative to Sec61 $\beta$  (Harant *et al.*, 2006). We asked whether this is also true for VEGF. Valine 30 (fourth amino-acid residue of the SEAP mature domain) was replaced by a cysteine residue in the wt VEGF SP-SEAP construct, the highly sensitive mutant VEGF (L12G, A13G) SP-SEAP, and in the constructs VEGF (L12I, A13I) SP-SEAP and VEGF ( $\Delta$ 2-5) SP-SEAP which have greatly reduced sensitivities to inhibition by CAM741. We chose position 30 for this substitution to exclude any effect of potential SP cleavage on crosslinking of targeted NCs, as it is located downstream of the cleavage site but in close vicinity to the SP. The constructs were then evaluated by transient transfections of HEK293 cells. As shown in Table 2, the constructs displayed a pattern of CAM741 sensitivity comparable to those containing valine at position 30, although overall sensitivity of the mutants was lower with cysteine at this position.

Short translocation intermediates (VEGF SP + 54 amino-acid residues SEAP mature domain) were generated and targeted nascent chains subjected to chemical crosslinking with BMH and immunoprecipitation with a Sec61 $\beta$  antiserum. At this length of the NCs, the only cysteine residue available for crosslinking is provided at position 30 (Figure 4B). Similar to the VCAM1

SP (Harant *et al.*, 2006), a dose-dependent increase in Sec61 $\beta$  crosslinks of targeted VEGF (V30C) SP-SEAP NCs by CAM741 was observed. However, as also observed for wt VEGF NCs of 81 amino- acid residues (Figure 4A), there were already some basal crosslinks to Sec61 $\beta$  detected in the absence of compound (Figure 4B). The highly sensitive construct VEGF (L12G, A13G, V30C) SP-SEAP showed only weak crosslinks to Sec61 $\beta$  in the vehicle-treated reaction, but also formation of Sec61 $\beta$  crosslinks with increasing concentrations of CAM741. In contrast, the construct VEGF (L12I, A13I, V30C) SP-SEAP with greatly reduced sensitivity to CAM741, formed basal crosslinks to Sec61 $\beta$  in the absence of compound which were not enhanced by the presence of CAM741 (Figure 4B). However, the VEGF SP mutant lacking the amino-acid residues 2-5 (VEGF ( $\Delta$ 2-5, V30C) SP-SEAP), although having strongly reduced sensitivity to CAM741, showed only weak basal crosslinks with Sec61 $\beta$  and some enhanced formation of the Sec61 $\beta$  crosslink only at 1  $\mu$ M CAM741 (Figure 4B). These data indicate that substitution of Leu12 and Ala13 by isoleucines affects SP association with the translocon, and could indicate a different and more efficient interaction with the translocon. In contrast, the presence of the glycines at positions 12 and 13 could cause inefficient association of the SP with the translocon, suggesting that the amino-acid residues within the h-region may control the strength of translocon binding. The construct VEGF ( $\Delta$ 2-5) SP-SEAP, which lacks the N-terminal amino-acid residues 2-5, but contains no mutations within the h-region, shows only weak basal crosslinks to Sec61 $\beta$ , which further supports the idea that basal crosslinking to Sec61 $\beta$  is mediated through the h-region. However, the individual sensitivities of the VEGF SP mutants to CAM741 is mirrored by the different concentrations of compound required to induce Sec61 $\beta$  crosslinks.

*CAM741 inhibits N-terminal translocation of a tag fused to the VEGF SP.*

Translocation of a 17 amino-acid residues tag (N-tag) fused to the VCAM1 SP was inhibited by CAM741, which additionally indicated incorrect SP insertion into the translocon channel caused by the presence of compound (Harant *et al.*, 2006). We therefore performed targeting of 81 amino-acid residues VEGF NCs fused to the 17 residues N-tag in the presence or absence of CAM741. This tag contains a diagnostic glycosylation site according to the constructs used by Heinrich *et al.* (Heinrich *et al.*, 2000), and N-terminal translocation of the tag is visualized by glycosylation (Figure 5). Glycosylation can only occur at the N-tag, as at a chain length of 81 amino-acid residues, the wt VEGF does not contain any glycosylation sites (Figure 5, left upper panel). Glycosylation of the N-terminal tag was further confirmed by treatment with endoglycosidase F (Figure 5, right upper panel). However, in the presence of the N-tag, efficient glycosylation was seen in the control reaction, but was clearly reduced when CAM741 was present (Figure 5, middle upper panel). In addition, inhibition of N-terminal translocation of the tag by CAM741 was dose-dependent (Figure 5, lower panel). Together, these data indicate altered insertion of the VEGF SP in the presence of compound.

*Differential sensitivity of the VEGF and VCAM1 SPs to the cyclopeptolide derivative NFI028.*

In addition to side chain modification of HUN-7293 providing compounds such as CAM741, variation of the peptidic backbone was investigated. These efforts disclosed cyclic hexapeptides such as NFI028 as novel structural type of potent inhibitor of VCAM-1 expression (Schreiner *et al.*, 2003; Figure 6). To determine if this fundamental structural difference has an impact on VCAM1 and VEGF SP-dependent translocation, HEK293 cells were transiently transfected with VCAM1 ( $\Delta$ 2-10) SP-SEAP or VEGF SP-SEAP, and treated with increasing concentrations of



NFI028. While this compound was able to inhibit SEAP release from cells transfected with VCAM1 ( $\Delta$ 2-10) SP-SEAP at low concentrations, it did not inhibit release of SEAP by cells transfected with VEGF SP-SEAP, demonstrating that although both SPs respond to CAM741, the VEGF SP shows no sensitivity to this derivative (Table 3). Based on the observations above, that the sensitivity of the VEGF SP to CAM741 could be enhanced by mutations within the h-region, we tested whether three of the highly sensitive mutants, VEGF (A13P) SP-SEAP, VEGF (Y17P) SP-SEAP and VEGF (L12G, A13G) SP-SEAP could respond to inhibition by NFI028. However, there was only a partial response to NFI028 observed with the mutants VEGF (A13P) SP-SEAP or VEGF (Y17P) SP-SEAP. The highly CAM741-sensitive mutant VEGF (L12G, A13G) SP-SEAP showed some increased sensitivity to NFI028, although it was markedly reduced compared to CAM741 (Table 3). These data indicate that, apart from general similarities between the VEGF and VCAM1 SPs which make them susceptible to inhibition by CAM741, differences in the composition of the SP may account for NFI028 selectivity.

## Discussion

Inhibition of cotranslational translocation through the SP was identified by us as a novel approach to interfere with the expression of proteins undergoing the secretory pathway. The proof of concept was provided by the discovery of the cyclopeptolide CAM741, which potently inhibits cotranslational translocation of VCAM1 (Besemer *et al.*, 2005). At the same time, cotransin, a compound of similar structure and activity has been reported by Garrison *et al.* (Garrison *et al.*, 2005). The mechanism by which this process is inhibited has been shown to be dependent on the VCAM1 SP at the level of its attachment to the Sec61 translocon (Besemer *et al.*, 2005; Garrison *et al.*, 2005; Harant *et al.*, 2006). We were interested whether other SPs would also be able to respond to CAM741 to learn more about this mechanism. Garrison *et al.* reported some SPs with partial sensitivity to cotransin, which however lack any obvious consensus motif in their sequences (Garrison *et al.*, 2005).

From a panel of SPs analysed, the most sensitive SP identified was that of VEGF; inhibition of VEGF SP-SEAP release required only 4-fold higher concentrations of CAM741 than inhibition of VCAM1 SP-SEAP (Besemer *et al.*, 2005; Harant *et al.*, 2006). By *in vitro* translocation experiments we provide evidence that this inhibition also occurs at the level of cotranslational translocation. Moreover, the mechanism appears to be very similar or identical to that observed for VCAM1 translocation inhibition, although the two SPs do not share any obvious similarities within their primary sequences.

These findings prompted us to analyse the features of the VEGF SP responsible for inhibition. In contrast to the VCAM1 SP, where the critical residues are located in the h-region and the polar c-

region upstream of the cleavage site (Harant *et al.*, 2006), residues of the VEGF SP required for translocation inhibition are located in the n-region and the h-region, where positions Leu12 and Ala13 were identified as most critical for sensitivity. Aliphatic residues with increased hydrophobicity and/or size at these positions resulted in a decrease in sensitivity, while conversely reducing hydrophobicity or tendency for  $\alpha$ -helix formation resulted in enhanced sensitivity to the compound. This indicates, that these residues are located in or near a part of the h-region essential for translocon interaction. The h-region has been reported to be required for SP interaction with the translocon (Mothes *et al.*, 1998), and CAM741 could interfere at this level. Amino-acid changes within the h-region of the prolactin SP have been reported to alter its association with the translocon relative to Sec61 $\alpha$  and Sec61 $\beta$ , consequently affecting further processing such as SP cleavage and N-glycosylation of the mature domain (Rutkowski *et al.*, 2003). We have observed that mutations in the VCAM1 SP, where hydrophobicity was increased in the CAM741-sensitive region, caused a different association with the translocon, witnessed by enhanced crosslinks to Sec61 $\alpha$  and Sec61 $\beta$  compared to the wt VCAM1 SP (Harant *et al.*, 2006). Also with VEGF SP variants, a different association with the translocon was seen in chemical crosslinking experiments. While wt VEGF showed some basal crosslinks to Sec61 $\beta$ , the less sensitive mutant VEGF (L12I, A13I, V30C) SP-SEAP already formed clearly visible basal crosslinks with Sec61 $\beta$  in the absence of compound, possibly due to more efficient translocon interaction. This demonstrates individual association of the VEGF SP variants with the Sec61 translocon, and could even involve different interaction sites.

The strength or site of translocon binding may be one explanation for the individual compound sensitivities of the VEGF SP mutants. However, the less sensitive mutant VEGF ( $\Delta$ 2-5, V30C)

SP-SEAP which lacks the N-terminal residues 2-5 but has no mutations in the h-region, gave only low basal crosslinks to Sec61 $\beta$  and these only increased at the highest concentration of compound tested. This demonstrates that compound sensitivity can be reduced despite the presence of an intact h-region. Thus, this segment appears to mediate translocon interaction and determines proximity to Sec61 $\beta$ , but compound sensitivity of the VEGF SP requires an interplay between the n- and h-region, which could argue for a specific conformational requirement for optimal inhibition.

During the translocation process, transmembrane domains can acquire a limited degree of protein folding, such as formation of an  $\alpha$ -helix, and, depending on the features of the transmembrane segment, already inside the ribosome (Mingarro *et al.*, 2000; Woolhead *et al.*, 2004). Certain mutations in the h-region of the VEGF SP could support formation of a stabilized helix which may enhance efficiency in translocon binding. In contrast, introduction of residues with helix-breaking potential, such as glycine or proline, could decrease helix formation propensities. Model SPs have been shown to initially insert with the N-terminus facing towards the luminal side, followed by a reorientation with growing chain lengths. Such a dynamic reorientation has been suggested to occur more easily with unstable or kinked helices rather than stabilized helices (Rosch *et al.*, 2000; Goder and Spiess, 2003). Alterations in helix formation propensity and thus enhanced flexibility of the VEGF SP within the translocon could be another reason for increased sensitivity to translocation inhibition by CAM741.

Although some of the VEGF SP mutants showed higher sensitivity to inhibition by CAM741, they showed very little response to the derivative NFI028, which however is fully active against

VCAM1. Only the mutant VEGF (L12G, A13G) SP-SEAP showed some sensitivity to NFI028, although much higher concentration of the compound was required compared to VCAM1, indicating that specific features of the VEGF SP account for the poor response to NFI028. Although currently a direct binding of compound to the SP cannot be fully excluded, from our data it seems more likely that there exists a competition between the compound and SP for binding to a specific site in the translocon required to initiate the translocation process. However, as previously shown, the compound does not prevent SP binding to the translocon, but may rather force the SP into a position where luminal translocation cannot occur, resulting in synthesis of the growing polypeptide chains towards the cytosolic side (Besemer *et al.*, 2005; Harant *et al.*, 2006). According to this hypothesis, both CAM741 and NFI028 could interact with the translocon, although binding of NFI028 would be weaker. The compounds could compete with the VCAM1 SP which interacts with the translocon inefficiently, while the VEGF SP binds more efficiently and cannot be competed by NFI028. This assumption is supported by the observation that mutants of the VCAM1 SP with only slightly reduced sensitivities to CAM741 had much lower sensitivities to NFI028 (*unpublished observations*). However, the similarity in CAM741-sensitivity but the large difference in NFI028-sensitivity of the VCAM1 and VEGF (L12G, A13G) SP would argue against this model and suggest that not only competition but also additional SP-dependent features contribute to this selectivity.

The logical next step therefore will be the analysis of several SPs which show different degrees of sensitivity to CAM741 and study of their association with the translocon. In addition, it would be interesting to evaluate whether insensitive SPs can be converted into sensitive ones by introduction of mutations based on our findings.

*Acknowledgements.* We wish to thank Roland Reuschel, Waltraud Mayer-Granitzer and Eva-Marie Haupt for sequencing, Christiane Dascher-Nadel for generation of the VEGF constructs and SP-SEAP fusion constructs. We also wish to thank Siegfried Höfinger, Piroška Devay and Markus Jaritz for helpful discussions.

## References

Besemer J, Harant H, Wang S, Oberhauser B, Marquardt K, Foster C A, Schreiner E P, de Vries J E, Dascher-Nadel C and Lindley I J (2005) Selective Inhibition of Cotranslational Translocation of Vascular Cell Adhesion Molecule 1. *Nature* 436:290-293.

Cardones AR and Banez L L (2006) VEGF Inhibitors in Cancer Therapy. *Curr Pharm Des* 12:387-394.

Eichler W, Yafai Y, Wiedemann P and Fengler D (2006) Antineovascular Agents in the Treatment of Eye Diseases. *Curr Pharm Des* 12:2645-2660.

Fons RD, Bogert B A and Hegde R S (2003) Substrate-Specific Function of the Translocon-Associated Protein Complex During Translocation Across the ER Membrane. *J Cell Biol* 160:529-539.

Garrison JL, Kunkel E J, Hegde R S and Taunton J (2005) A Substrate-Specific Inhibitor of Protein Translocation into the Endoplasmic Reticulum. *Nature* 436:285-289.

Gille J, Khalik M, Konig V and Kaufmann R (1998) Hepatocyte Growth Factor/Scatter Factor (HGF/SF) Induces Vascular Permeability Factor (VPF/VEGF) Expression by Cultured Keratinocytes. *J Invest Dermatol* 111:1160-1165.

Goder V and Spiess M (2003) Molecular Mechanism of Signal Sequence Orientation in the Endoplasmic Reticulum. *EMBO J* 22:3645-3653.

Harant H, Lettner N, Hofer L, Oberhauser B, de Vries J E and Lindley I J (2006) The Translocation Inhibitor CAM741 Interferes With Vascular Cell Adhesion Molecule 1 Signal Peptide Insertion at the Translocon. *J Biol Chem* 281:30492-30502.

Hegde RS and Bernstein H D (2006) The Surprising Complexity of Signal Sequences. *Trends Biochem Sci* 31:563-571.

Hegde RS and Lingappa V R (1997) Membrane Protein Biogenesis: Regulated Complexity at the Endoplasmic Reticulum. *Cell* 91:575-582.



Heinrich SU, Mothes W, Brunner J and Rapoport T A (2000) The Sec61p Complex Mediates the Integration of a Membrane Protein by Allowing Lipid Partitioning of the Transmembrane Domain. *Cell* 102:233-244.

High S, Martoglio B, Gorlich D, Andersen S S, Ashford A J, Giner A, Hartmann E, Prehn S, Rapoport T A and Dobberstein B (1993) Site-Specific Photocross-Linking Reveals That Sec61p and TRAM Contact Different Regions of a Membrane-Inserted Signal Sequence. *J Biol Chem* 268:26745-26751.

Johnson AE and van Waes M A (1999) The Translocon: a Dynamic Gateway at the ER Membrane. *Annu Rev Cell Dev Biol* 15:799-842.

Keck PJ, Hauser S D, Krivi G, Sanzo K, Warren T, Feder J and Connolly D T (1989) Vascular Permeability Factor, an Endothelial Cell Mitogen Related to PDGF. *Science* 246:1309-1312.

Laird V and High S (1997) Discrete Cross-Linking Products Identified During Membrane Protein Biosynthesis. *J Biol Chem* 272:1983-1989.

Leung DW, Cachianes G, Kuang W J, Goeddel D V and Ferrara N (1989) Vascular Endothelial Growth Factor Is a Secreted Angiogenic Mitogen. *Science* 246:1306-1309.

Matlack KE, Mothes W and Rapoport T A (1998) Protein Translocation: Tunnel Vision. *Cell* 92:381-390.

Mingarro I, Nilsson I, Whitley P and von Heijne G (2000) Different Conformations of Nascent Polypeptides During Translocation Across the ER Membrane. *BMC Cell Biol* 1:3.

Mothes W, Jungnickel B, Brunner J and Rapoport T A (1998) Signal Sequence Recognition in Cotranslational Translocation by Protein Components of the Endoplasmic Reticulum Membrane. *J Cell Biol* 142:355-364.

Mothes W, Prehn S and Rapoport T A (1994) Systematic Probing of the Environment of a Translocating Secretory Protein During Translocation Through the ER Membrane. *EMBO J* 13:3973-3982.

Nielsen H, Brunak S and von Heijne G (1999) Machine Learning Approaches for the Prediction of Signal Peptides and Other Protein Sorting Signals. *Protein Eng* 12:3-9.

Nielsen H, Engelbrecht J, Brunak S and von Heijne G (1997) Identification of Prokaryotic and Eukaryotic Signal Peptides and Prediction of Their Cleavage Sites. *Protein Eng* 10:1-6.

Osborne AR, Rapoport T A and van den Berg, B (2005) Protein Translocation by the Sec61/SecY Channel. *Annu Rev Cell Dev Biol.* 21:529-550

Rapoport TA, Jungnickel B and Kutay U (1996) Protein Transport Across the Eukaryotic Endoplasmic Reticulum and Bacterial Inner Membranes. *Annu Rev Biochem* 65:271-303.

Robinson CJ and Stringer S E (2001) The Splice Variants of Vascular Endothelial Growth Factor (VEGF) and Their Receptors. *J Cell Sci* 114:853-865.

Rosch K, Naeher D, Laird V, Goder V and Spiess M (2000) The Topogenic Contribution of Uncharged Amino Acids on Signal Sequence Orientation in the Endoplasmic Reticulum. *J Biol Chem* 275:14916-14922.

Roy H, Bhardwaj S and Yla-Herttuala S (2006) Biology of Vascular Endothelial Growth Factors. *FEBS Lett* 580:2879-2887.

Rutkowski DT, Lingappa V R and Hegde R S (2001) Substrate-Specific Regulation of the Ribosome- Translocon Junction by N-Terminal Signal Sequences. *Proc Natl Acad Sci U S A* 98:7823-7828.

Rutkowski DT, Ott C M, Polansky J R and Lingappa V R (2003) Signal Sequences Initiate the Pathway of Maturation in the Endoplasmic Reticulum Lumen. *J Biol Chem* 278:30365-30372.

Schreiner EP, Oberhauser B and Foster C A (2003) Inhibitors of Vascular Cell Adhesion Molecule-1 Expression. *Expert Opinion on Therapeutic Patents* 13:149-166.

Stroud RM and Walter P (1999) Signal Sequence Recognition and Protein Targeting. *Curr Opin Struct Biol* 9:754-759.

Voigt S, Jungnickel B, Hartmann E and Rapoport T A (1996) Signal Sequence-Dependent Function of the TRAM Protein During Early Phases of Protein Transport Across the Endoplasmic Reticulum Membrane. *J Cell Biol* 134:25-35.

Woolhead CA, McCormick P J and Johnson A E (2004) Nascent Membrane and Secretory Proteins Differ in FRET-Detected Folding Far Inside the Ribosome and in Their Exposure to Ribosomal Proteins. *Cell* 116:725-736.

### Figure Legends

#### **Figure 1. CAM741 inhibits cotranslational translocation of VEGF SP-SEAP. Panel A.**

Sequence and schematic representation of the construct used. In vitro translocation of truncated VEGF SP-SEAP NCs (SP + 146 amino-acid residues SEAP mature domain) in the absence or presence of CAM741 (1  $\mu$ M), either untreated or treated with proteinase K (left panel); deglycosylation of sedimented VEGF SP-SEAP NCs with endoglycosidase F (Endo F; right panel). **Panel B.** Schematic representation of the construct used. In vitro translocation of truncated 131 amino-acid residues VEGF NCs in the absence of microsomes (left panel), in the presence of microsomes (middle panel), or in the presence of microsomes and 1  $\mu$ M CAM741 (right panel), either untreated, or treated with proteinase K, or proteinase K and 1% Triton X-100. *Open circle*, glycosylated NCs with the SP attached; *arrow*, glycosylated NCs without SP; *star*, non-processed NCs; *arrowhead*, NCs with SP cleaved off.

**Figure 2. CAM741 inhibits release of endogenously expressed VEGF<sub>165</sub>.** HaCaT cells were treated with TGF- $\alpha$  in the presence of increasing concentrations of CAM741. The concentration of released VEGF<sub>165</sub> (pg/ml) was determined by ELISA.

**Figure 3. Differential sensitivity of VEGF SP mutants to inhibition by CAM741.** Schematic representation of the constructs used. In vitro translocation of fusion constructs of VEGF SP mutants and the 146 amino-acid residues SEAP mature domain in the absence of microsomal membranes, or in the presence of microsomes and increasing concentrations of CAM741. *Arrow*, glycosylated NCs; *star*, unprocessed NCs.

**Figure 4. CAM741 alters positioning of the VEGF NCs at the translocon. Panel A.**

Schematic representation of the construct used. In vitro targeting and chemical crosslinking with either MBS (left panels, labeled MBS) or BMH (right panels, labeled BMH) of truncated 81 amino-acid residues VEGF NCs and immunoprecipitation with a Sec61 $\alpha$  or Sec61 $\beta$  antiserum. *Closed diamonds*, Sec61 $\alpha$  crosslink; *closed circle*, Sec61 $\beta$  crosslink; *star*, non-processed NCs; *open arrowheads*, additional unspecified crosslinks; *open circle*, residual peptidyl-tRNA-NCs.

**Panel B.** Schematic representation of the constructs used. In vitro targeting and chemical crosslinking with BMH of truncated VEGF SP mutants fused to 54 amino-acid residues SEAP mature domain containing a cysteine at position 30 (V30C; fourth position of the SEAP mature domain; left panels) and immunoprecipitation with a Sec61 $\beta$  antiserum (right panels). *Closed circle*, Sec61 $\beta$  crosslink; *star*, NCs.

**Figure 5. CAM741 inhibits N-terminal translocation of a 17 amino-acid residues tag fused to the VEGF SP.**

Sequences and schematic representation of the constructs. The N-terminal tag is indicated in italic letters, the glycosylation site is underlined. In vitro targeting of truncated 81 amino-acid residues VEGF NCs (left upper panel), or 81 amino-acid residues VEGF NCs containing the 17 amino-acid residues N-terminal tag in the absence of microsomes, or in the presence of microsomes with or without 1  $\mu$ M CAM741 (middle upper panel). Deglycosylation with endoglycosidase F (Endo F; right upper panel). In vitro targeting of 81 amino-acid residues VEGF NCs containing the 17 amino-acid residues N-terminal tag in the presence of increasing concentrations of CAM741 (lower panel). *Arrow*, glycosylated NCs; *star*, unprocessed NCs; *arrowhead*, NCs with SP cleaved off.

**Figure 6. Structures of CAM741 and NFI028**



Table 1.

Different sensitivity of SP-SEAP fusion constructs to inhibition by CAM741

signal peptide	sequence	CAM741 IC <sub>50</sub> (nM)	inhibition at 10 $\mu$ M
CCL22 + 10	MARLQTALLVVLVLLAVALQATEA- <u>GPYGANMEDS</u>	>10000	
CCL2 + 10	MKVSAALLCLLLIAATFIPOGLA- <u>QPDAINAPVT</u>	>10000	23%
IL-13 SP + 10	MALLTTVIALTCLGGFASP- <u>GPVPPSTALR</u>	>10000	27%
IFN- $\gamma$ + 5	MKYTSYILAFQLCIVLGSGLCYC- <u>QDPYV</u>	>10000	45%
CCR7 SP + 10	MDLGKPMKSVLVVALLVIFQVCLC- <u>QDEVTTDDYIG</u>	>10000	49%
E-selectin	MIASQFLSALTLVLLIKESGA-	9753 $\pm$ 1791	
IL-12p40 + 3	MCHQQLVISWFSLVFLASPLVA- <u>IWE</u>	4313 $\pm$ 1381	
CXCL8 + 7	MTSKLAVALLAFLISAALC- <u>EGAVLPR</u>	1473 $\pm$ 764	
ICAM-1 + 5	MAPSSRPALPALLVLLGALFPGPGNA- <u>QTSVS</u>	687 $\pm$ 220	
VEGF	MNFLLSWVHWSLALLLYLHHAKWSQA-	125 $\pm$ 82	

HEK293 cells were transfected with different SP-SEAP fusion constructs (in some cases SP + additional residues of the mature domain) and incubated with increasing concentrations of CAM741. Twenty-four hours post-transfection, supernatants were harvested and analysed for alkaline phosphatase activity. Results shown are IC<sub>50</sub> values from at least three independent experiments performed in triplicates. The cleavage site is underlined, amino-acid residues of the mature region are boxed.

Table 2.

Modulation of the sensitivity to CAM741 by mutations in the VEGF SP

VEGF signal peptide	sequence	CAM741 IC50 (nM)
	n-region      h-region      c-region	
	<u>MN</u> FLLSWVHWS <u>L</u> LALLLYL <u>HHAKWSQA</u>	
<b>n-region</b>		
wt	MN <u>FLLS</u> WVHWSLALLLYLHHAKWSQA	125 ± 82
Δ2-5	MSWVHWSLALLLYLHHAKWSQA	1247 ± 345
L4G, L5G	MNF <u>GG</u> SWVHWSLALLLYLHHAKWSQA	312 ± 108
L4Q, L5Q	MNF <u>QQ</u> SWVHWSLALLLYLHHAKWSQA	1007 ± 408
<b>h-region</b>		
W7H	MN <u>FLLS</u> HVHWSLALLLYLHHAKWSQA	58 ± 38
H9F	MN <u>FLLS</u> WV <u>F</u> WSLALLLYLHHAKWSQA	93 ± 76
W10G	MN <u>FLLS</u> WVH <u>G</u> SLALLLYLHHAKWSQA	133 ± 89
L12G	MN <u>FLLS</u> WVHWS <u>G</u> ALLLYLHHAKWSQA	36 ± 21
A13G	MN <u>FLLS</u> WVHWSL <u>G</u> LLLYLHHAKWSQA	82 ± 53
L12G, A13G	MN <u>FLLS</u> WVHWS <u>G</u> GLLLYLHHAKWSQA	1.7 ± 1.0
L12A	MN <u>FLLS</u> WVHWS <u>A</u> ALLLYLHHAKWSQA	32 ± 8
L12V	MN <u>FLLS</u> WVHWS <u>V</u> ALLLYLHHAKWSQA	295 ± 110
A13V	MN <u>FLLS</u> WVHWSL <u>V</u> LLLYLHHAKWSQA	340 ± 103
L12V, A13V	MN <u>FLLS</u> WVHWS <u>VV</u> LLLYLHHAKWSQA	1148 ± 559
A13L	MN <u>FLLS</u> WVHWSL <u>L</u> LLLYLHHAKWSQA	2670 ± 757
L12I	MN <u>FLLS</u> WVHWS <u>I</u> ALLLYLHHAKWSQA	883 ± 297
A13I	MN <u>FLLS</u> WVHWSL <u>I</u> LLLYLHHAKWSQA	1484 ± 220

L12I, A13I	MNFLLSWVHWSIILLLLYLHHAKWSQA	2133 ± 741
A13P	MNFLLSWVHWSLPLLLLYLHHAKWSQA	6.5 ± 3.1
L14A, L15A	MNFLLSWVHWSLAAALYLHHAKWSQA	56 ± 18
L14G, L15G	MNFLLSWVHWSLAGGLYLHHAKWSQA	145 ± 69
L16V, L18V	MNFLLSWVHWSLALLVYVHHAKWSQA	648 ± 204
Y17P	MNFLLSWVHWSLALLLPLHHAKWSQA	12 ± 11
Δ2-5 Y17P	MSWVHWSLALLLPLHHAKWSQA	312 ± 162
<b>c-region</b>		
H19A	MNFLLSWVHWSLALLLYLAHAKWSQA	580 ± 211
H20A	MNFLLSWVHWSLALLLYLHAAKWSQA	766 ± 295
H19A, H20A	MNFLLSWVHWSLALLLYLAAAKWSQA	473 ± 162
K22E	MNFLLSWVHWSLALLLYLHHA <sup>E</sup> WSQA	378 ± 176
W23V	MNFLLSWVHWSLALLLYLHHAK <sup>V</sup> WSQA	475 ± 188
V30C	MNFLLSWVHWSLALLLYLHHAKWSQA - IIPC	177 ± 133
Δ2-5, V30C	MSWVHWSLALLLYLHHAKWSQA - IIPC	3803 ± 476
L12G, A13G, V30C	MNFLLSWVHWSGGLLLYLHHAKWSQA - IIPC	7.7 ± 5.8
L12I, A13I, V30C	MNFLLSWVHWSIILLLLYLHHAKWSQA - IIPC	4707 ± 1219

HEK293 cells were transfected with different VEGF SP-SEAP fusion constructs and incubated with increasing concentrations of CAM741. Twenty-four hours post-transfection, supernatants were harvested and analysed for alkaline phosphatase activity. Results shown are IC<sub>50</sub> values from at least three independent experiments performed in triplicates. Mutations are indicated by grey boxes, the cleavage site is underlined, amino-acid residues of the SEAP mature region are boxed.

Table 3.

Sensitivity of the VCAM1 ( $\Delta$ 2-10) SP and VEGF SP mutants to inhibition by NFI028

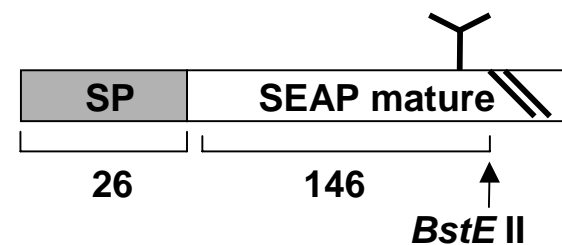
signal peptide	sequence	NFI028 IC <sub>50</sub> (nM)
VCAM ( $\Delta$ 2-10)	MASNILWIMFAASQA- <span style="border: 1px solid black; padding: 0 2px;">FKIE</span>	14.3 $\pm$ 2.3
VEGF	MNFLLSWVHWSLALLL <u>YLHHAKWSQA</u>	>10000
VEGF (Y17P)	MNFLLSWVHWSLALLL <span style="background-color: #cccccc;">P</span> LHHAKWSQA	3234 $\pm$ 1446
VEGF (A13P)	MNFLLSWVHWSL <span style="background-color: #cccccc;">P</span> LLLYLHHAKWSQA	4984 $\pm$ 2343
VEGF (L12G, A13G)	MNFLLSWVHWS <span style="background-color: #cccccc;">GG</span> LLLYLHHAKWSQA	464 $\pm$ 53

HEK293 cells were transfected with different SP-SEAP fusion constructs and incubated with increasing concentrations of NFI028. Twenty-four hours post-transfection, supernatants were harvested and analysed for alkaline phosphatase activity. Results shown are IC<sub>50</sub> values from at least three independent experiments performed in triplicates. Mutations are indicated by grey boxes, the cleavage site is underlined, amino-acid residues of the VCAM1 mature region are boxed.

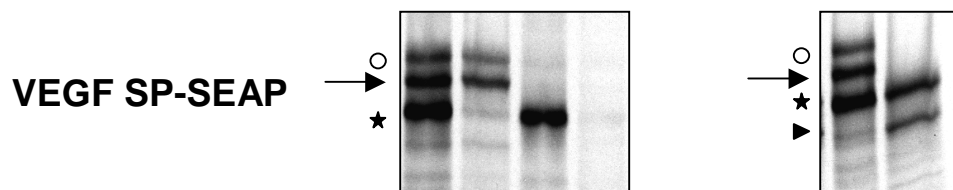
**Figure 1**

**A**

VEGF SP-SEAP      MNFLLSWVHWSLALLLYLHHAKWSQA - SEAP



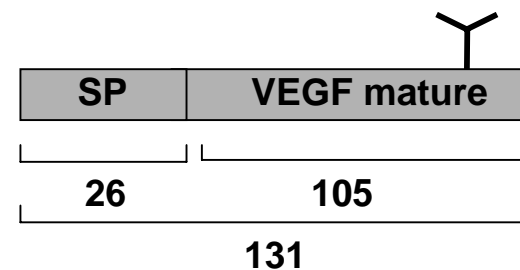
**High-resolution gel**



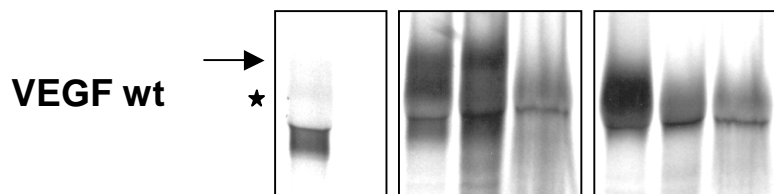
microsomes	+	+	+	+	Endo F	-	+
CAM741	-	-	+	+			
Proteinase K	-	+	-	+			

**B**

VEGF wt      MNFLLSWVHWSLALLLYLHHAKWSQA - mature



**High-resolution gel**



microsomes	-	-	+	+	+	+	+	+
CAM741	-	-	-	-	-	+	+	+
Proteinase K	-	+	-	+	+	-	+	+
Triton X-100	-	-	-	-	+	-	-	+

Figure 2

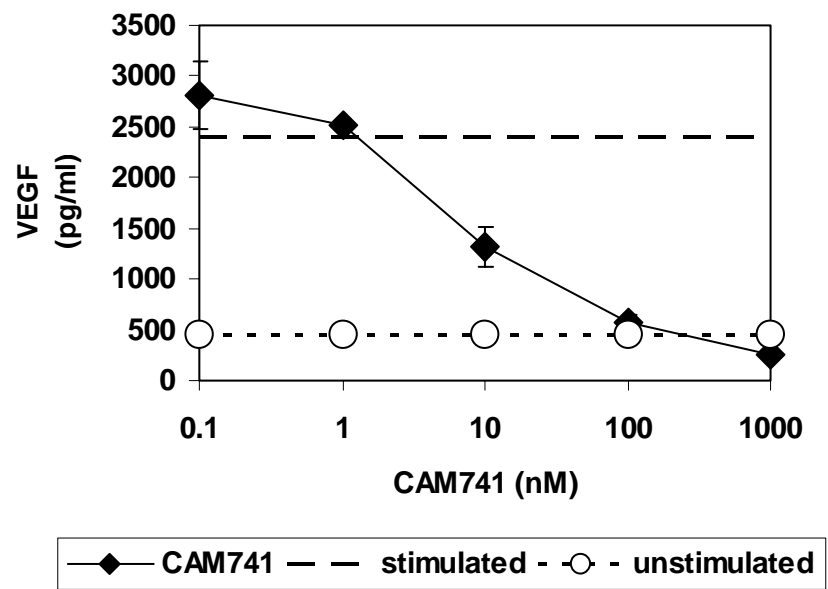


Figure 3

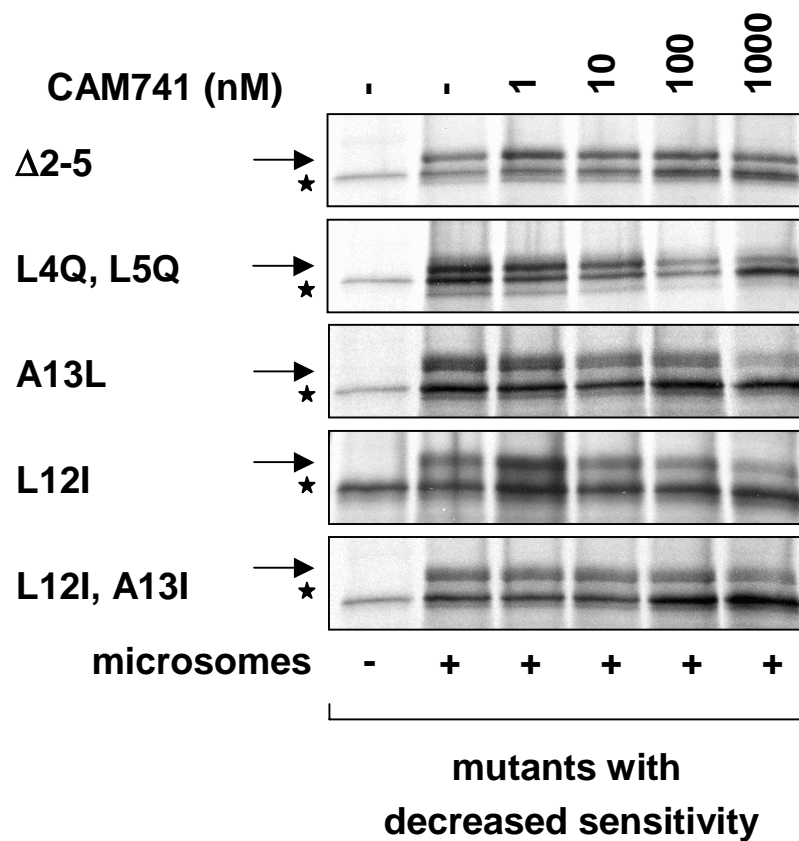
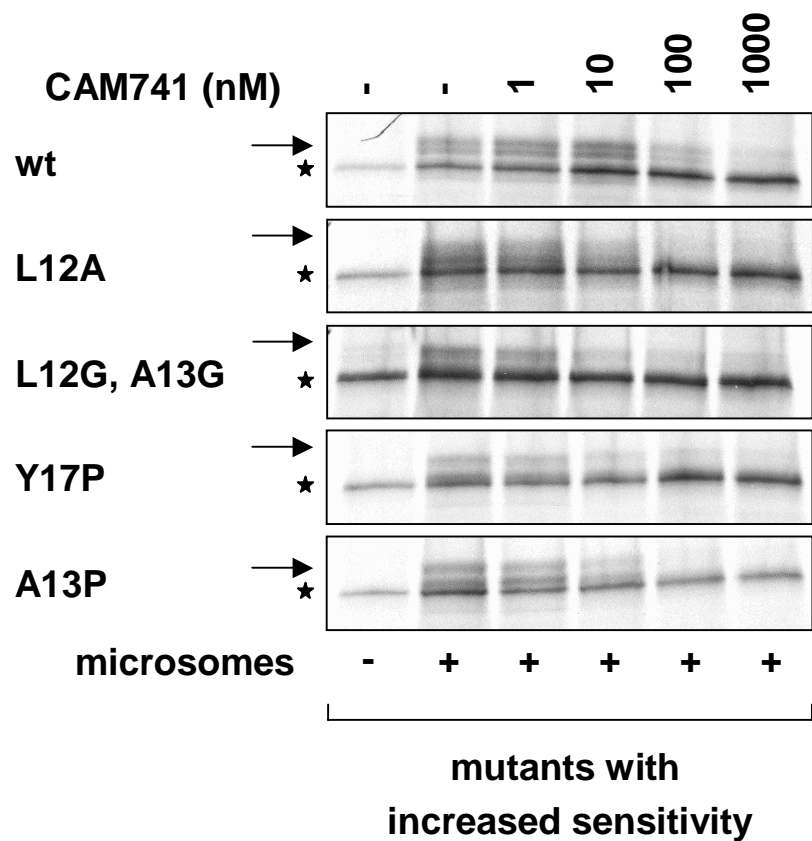
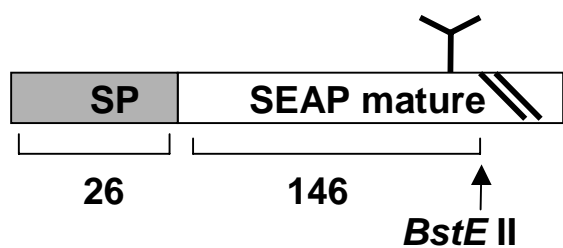


Figure 4

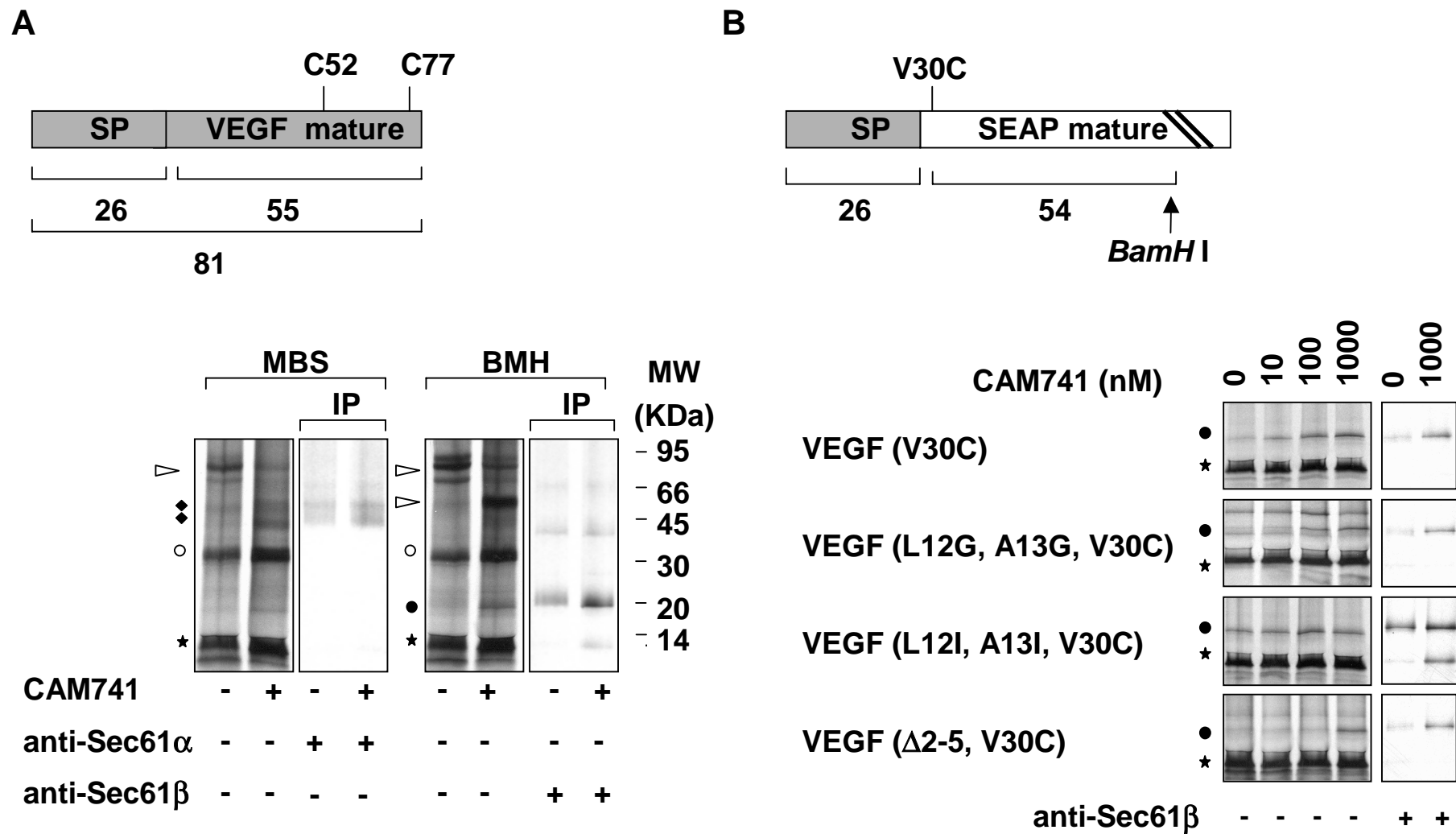




Figure 5

VEGF

MNFLLSWVHWSLALLLYLHHAKWSQA-

N-tag VEGF

MMNESSTLADSSATQAN-MNFLLSWVHWSLALLLYLHHAKWSQA-

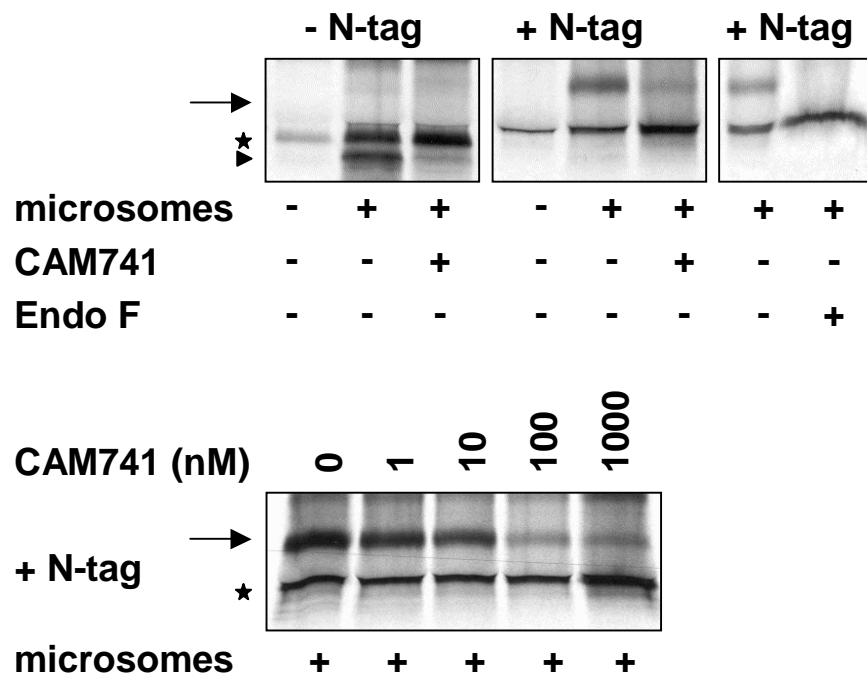
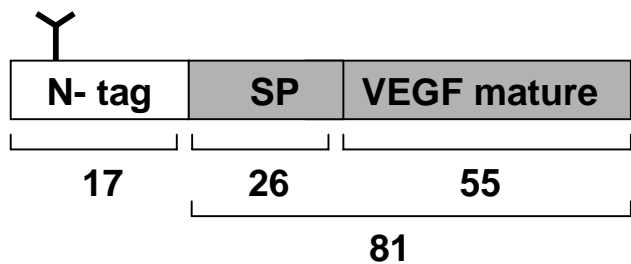
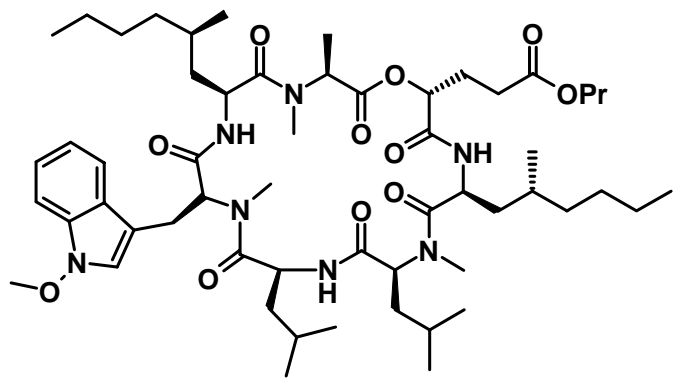
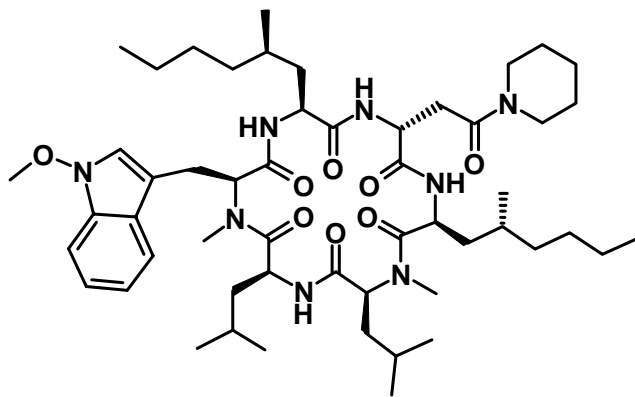


Figure 6



CAM741



NFI028