

**Activation of Nerve Growth Factor-Induced α by
Methylene-Substituted Diindolylmethanes in Bladder Cancer Cells
Induces Apoptosis and Inhibits Tumor Growth**

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Abbreviations:

C-DIM, methylene-substituted diindolylmethane; DIM-C-pPhOCH₃, 1,1-bis(3'-indolyl)-1-(*p*-methoxyphenyl)-methane; DIM-C-pPh, 1,1-bis(3'-indolyl)-1-(*p*-phenyl)methane; FBS, fetal bovine serum; iNur77, small inhibitory RNA for Nur77; iScr, nonspecific scrambled oligonucleotide; NAG-1, nonsteroidal anti-inflammatory drug activated gene; NGFI-B, nerve growth factor-induced B; RT-PCR, reverse transcriptase-polymerase chain reaction; TRAIL, tumor necrosis factor-related apoptosis-inducing ligand; TUNEL, terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling assay

ABSTRACT

Nerve growth factor-induced B (NGFI-B) genes are orphan nuclear receptors, and NGFI-B α (Nur77, TR3) is overexpressed in bladder tumors and bladder cancer cells compared to non-tumor bladder tissue. 1,1-Bis(3'-indolyl)-1-(*p*-methoxyphenyl)-methane (DIM-C-pPhOCH₃) and 1,1-bis(3'-indolyl)-1-(*p*-phenyl)methane (DIM-C-pPh) have previously been identified as activators of Nur77, and both compounds inhibited growth and induced apoptosis of UC-5 and KU7 bladder cancer cells. The proapoptotic effects of C-DIMs were unaffected by cotreatment with leptomycin B and dependent on nuclear Nur77, and RNA interference with a small inhibitory RNA for Nur77 (iNur77) demonstrated that C-DIM-induced activation of apoptosis was Nur77-dependent. Microarray analysis of DIM-C-pPhOCH₃-induced genes in UC-5 bladder cancer cells showed that this compound induced multiple Nur77-dependent proapoptotic or growth inhibitory genes including tumor necrosis factor-related apoptosis-inducing ligand (TRAIL), cystathionase, p21, p8 and sestrin-2. DIM-C-pPhOCH₃ (25 mg/kg/d) also induced apoptosis and inhibited tumor growth in athymic nude mice bearing KU7 cells as xenografts, demonstrating that Nur77-active C-DIMs exhibit potential for bladder cancer chemotherapy by targeting Nur77 which is overexpressed in this tumor type.

INTRODUCTION

The nuclear receptor family of transcription factors includes the steroid and thyroid hormone, vitamin D, retinoid and ecdysone receptors, ligand-activated orphan receptors, and orphan receptors with no known ligands (Mangelsdorf et al., 1995; Milbrandt, 1988). Nuclear receptors influence diverse aspects of normal physiology in multiple tissues, and several receptors are drug targets for treating several diseases including cancer. Nerve growth factor-induced B (NGFI-B) is part of a subfamily of orphan nuclear receptors and, members of this subfamily include Nur77 (NGFI-B α , TR3), Nurr1 (NGFI-B β), and Nor1 (NGFI-B γ). Nur77 is expressed in multiple tissues; Nurr1 has been detected in thymus osteoblasts, liver and pituitary gland, and Nor1 is highly expressed in the pituitary gland with low expression in other tissues (Bandoh et al., 1997; Maruyama et al., 1997; Milbrandt, 1988). The physiological roles for NGFI-B proteins are not fully understood, however, gene targeting knockout experiments demonstrate several important functions for these proteins. For example, Nurr1 knockout mice have severe impairments in mid-brain neuronal development and dopamine expression, and these animals die soon after birth (DeYoung et al., 2003; Saucedo-Cardenas et al., 1998; Zetterstrom et al., 1997); Nor1 knockout animals die at gestation day 5 (DeYoung et al., 2003), whereas Nur77 knockout mice do not exhibit a specific phenotype (Lee et al., 1995), and this may be related to coexpression of both Nur77/Nor1 which exhibit some overlapping functions.

Several studies suggest that in cancer cells, Nur77 plays a role in cell death pathways activated by apoptosis inducing agents (Holmes et al., 2003a; Holmes et al., 2003b; Li et al., 2000; Lin et al., 2004; Mu and Chang, 2003; Wilson et al., 2003a; Wu et

al., 2002). Li and coworkers (Li et al., 2000) reported that treatment of LNCaP prostate cancer cells with agents such as retinoids, TPA and TNF- α resulted in induction of Nur77 gene expression. Surprisingly, induction of apoptosis and cytochrome c release from the mitochondria was independent of the DNA binding domain of Nur77 and treatment with leptomycin B (a blocker of nuclear export) inhibited induction of Nur77-dependent apoptosis. Induction of apoptosis was accompanied by translocation of Nur77 from the nucleus to the mitochondria, and Nur77 specifically interacted with Bcl-2 and converted Bcl-2 into a proapoptotic factor in HEK293T and HCT116 cells (Lin et al., 2004). However, a study in colon cancer cells reported that butyrate-induced apoptosis was associated with nuclear to cytoplasmic translocation of Nur77 which was not accompanied by subsequent mitochondrial interactions (Wilson et al., 2003a).

Studies in this laboratory have characterized a series of methylene-substituted diindolylmethane (C-DIM) analogs as activators of orphan receptors (Chintharlapalli et al., 2005a; Chintharlapalli et al., 2004; Cho et al., 2007; Inamoto et al., 2008; Kassouf et al., 2006; Qin et al., 2004). Two of these compounds, namely 1,1-bis(3'-indolyl)-1-(*p*-methoxyphenyl)methane (DIM-C-pPhOCH₃) and 1,1-bis(3'-indolyl)-1-(*p*-phenyl)methane (DIM-C-pPh), activate Nur77 in colon and pancreatic cancer cells (Chintharlapalli et al., 2005a; Cho et al., 2007), and 1,1-bis(3'-indolyl)-1-(*p*-chlorophenyl)methane (DIM-C-pPhCl) activates NGFI-B β (Nurr1) orphan receptor in bladder cancer cells (Inamoto et al., 2008). Nur77 and Nurr1 are widely expressed in bladder cancer cells (25) and we now show overexpression of Nur77 in bladder tumors. Nur77-active C-DIMs, such as DIM-C-pPhOCH₃, inhibit bladder cancer cell growth and induce apoptosis. DIM-C-pPhOCH₃ activates nuclear Nur77, and results of RNA interference studies show that

induction of apoptosis is due to induction of several proapoptotic genes and proteins including tumor necrosis factor-related apoptosis inducing ligand (TRAIL). DIM-C-pPhOCH₃ inhibited bladder tumor growth in a xenograft mouse model, suggesting that C-DIM-dependent activation of Nur77 is a promising new mechanism-based pathway for developing new agents for bladder cancer chemotherapy.

MATERIALS AND METHODS

Cells, biochemicals and antibodies. Human bladder cancer cell lines KU7 and UC-5 were provided by Dr. Ashish Kamat (M.D. Anderson Cancer Center, Houston, TX). Cells were maintained in DMEM/Ham's F-12 (Sigma, St. Louis, MO) without phenol red supplemented with 0.22% sodium bicarbonate, 0.011% sodium pyruvate, 5% fetal bovine serum (FBS), and 10 ml/L of 100X antibiotics antimycotic solution (Sigma-Aldrich). Cells were maintained at 37°C in the presence of 5% CO₂. Antibodies for cleaved PARP and cleaved caspase 8 were purchased from Cell Signaling (Danvers, MA). Antibodies for Nur77, TRAIL, Sp1, IgG and β -tubulin were purchased from Santa Cruz Biotechnology (Santa Cruz, CA). Nur77 antibodies from Imgenex (San Diego, CA) were also used for immunostaining and western blots, and results were similar to Nur77 antibodies from Santa Cruz. Caspase inhibitor (z-VAD) was obtained from Alexis Biochemicals (Lausen, Switzerland). Western Lightning chemiluminescence reagent was from Perkin-Elmer Life Sciences (Boston, MA). For RNA interference assays, we used a nonspecific scrambled (iScr) oligonucleotide as described (Abdelrahim et al., 2002). The small inhibitory RNA for Nur77 (iNur77) was identical to the reported oligonucleotide (Lin et al., 2004), and these were purchased from Dharmacon Research (Lafayette, CO). Leptomycin B was purchased from Sigma. The C-DIMs were synthesized in this laboratory as previously described (Qin et al., 2004).

Cell proliferation assay. KU-7 and UC-5 cells were seeded in 12-well plates and were treated with DMSO and different concentrations of test compounds dissolved in DMSO as indicated in DMEM/Ham's F-12 containing 2.5% charcoal-stripped FBS for 24 and 48 hr until plates reached 50-60% confluence which was usually observed 24 hr

after seeding. KU7 and UC-5 cells were counted to evaluate the effect of C-DIMs and DMSO (solvent control) on viable cell number using Z1 Coulter Particle Counter (Beckman Coulter, Fullerton, CA). Each experiment was carried out in triplicate and results are expressed as means \pm SD for each treatment group.

Western blot analysis. Cells were treated with the C-DIM compounds, and caspase inhibitor and leptomycin B were added simultaneously. For RNA interference studies, the oligonucleotides (iScr or iNur77) were transfected into bladder cancer cells and after 36 hr cells were treated with DMSO or C-DIM compounds as described (Cho et al., 2007). Cells were collected and cell lysates were prepared using lysis buffer [20 mM Tris-HCl, pH 7.4, 150 mM NaCl, 1 mM ethylenediaminetetraacetic acid (EDTA), 1% Triton X-100, 0.1% sodium dodecyl sulfate (SDS), 1 mM sodium orthovanadate; 1 mM phenylmethylsulfonyl fluoride (PMSF), 1 μ M leupeptin and 1 μ g/ml aprotinin]. After centrifugation of lysates at 15,000 *g* for 20 min, the supernatants were recovered, and protein was quantified by the Bradford protein assay using reagent kit from Bio-Rad (Hercules, CA). Protein samples (20-60 μ g) were size-separated by electrophoresis on sodium dodecylsulfate-polyacrylamide gels under non-reducing conditions. Separated proteins were electroblotted onto nitrocellulose membranes. The blot was blocked by incubating in blocking buffer (5% skim milk, 10 mM Tris, pH 7.5, 10 mM NaCl and 0.1% Tween 20) for 1 hr at 20°C, and was incubated with the primary antibody overnight at 4°C. Incubation with a horseradish peroxidase-conjugated anti-mouse or rabbit secondary antibody was then carried out at 37°C for 1 hr. Antibody-bound proteins were detected by the ECL western blotting analysis system.

Immunostaining. Cells were fixed immediately in 4% paraformaldehyde, added with 0.3% Triton X-100 (Roche Molecular Biochemicals) for 10 min, and pre-incubated for 1 hr with 10% normal goat serum (Vector laboratories). Cells were incubated with anti-Nur77 antibody (1:100) or anti-IgG (1:100) and were incubated with FITC-conjugated secondary antibody (1:200, Vector Laboratories, Burlingame, CA). The 4-well chambers were mounted with mounting medium (Vector Laboratories) to be viewed on a fluorescence microscope (Olympus).

Reverse transcriptase-polymerase chain reaction (RT-PCR). Total RNA was extracted using RNeasy Mini Kit (Qiagen Inc., Valencia, CA), and one microgram of RNA was used to synthesize cDNA using Reverse Transcription System (Promega). The PCR conditions were as follows: initial denaturation at 94°C (2 min) followed by 28 cycles (NAG-1 and CSE), 30 cycles (p8 and Sest-in-2) or 26 cycles (GAPDH) of denaturation for 1 min at 94°C, annealing for 1 min at 58°C (NAG-1), 61°C (CSE, p8, Sest-in-2 and GAPDH) and extension at 72°C for 1 min, and a final extension step at 72°C for 5 min. The mRNA levels were normalized using GAPDH as an internal housekeeping gene. Primers obtained from IDT (Coralville, IA) and used for amplification were as follows: NAG-1 sense 5'-GTG CTC ATT CAA AAG ACC GAC ACC G-3', antisense 5'-ATA CAC AGT TCC ATC AGA CCA GCC CC-3'; p8 sense 5'-ATG GCC ACC TTC CCA CCA GCA-3', antisense 5'-TCA GCG CCG TGC CCC TCG CT-3'; CSE sense 5'-GGC GAT CCA TGT GGG CCA GGA-3', antisense 5'-ATG TCT CCA TGC TTA TGG ACA AT-3'; sest-in-2 sense 5'-GAC TCC GAG TGC CGC GCA GAG-3', antisense 5'-ATG GCG GGC GGC AGC CAT GAT-3'; and GAPDH sense 5'-ACG GAT TTG GTC GTA TTG GGC G-3', antisense 5'-CTC CTG GAA GAT GGT GAT

GG-3'. PCR products were electrophoresed on 1% agarose gels containing ethidium bromide and visualized under UV transillumination.

Quantitative real-time PCR. cDNA was prepared from the total RNA of cells using Reverse Transcription System (Promega). Each PCR was carried out in triplicate in a 20- μ L volume using SYBR Green Mastermix (Applied Biosystems, Foster City, CA) for 15 min at 95°C for initial denaturing, followed by 40 cycles of 95°C for 30 s and 60°C for 1 min in the Applied Biosystems 7900HT Fast Real-time PCR System. The ABI Dissociation Curves software was used following a brief thermal protocol (95°C for 15 s and 60°C for 15 s, followed by a slow ramp to 95°C) to control for multiple species in each PCR amplification. Values for each gene were normalized to expression levels of TATA-binding protein. The sequences of the primers used for real-time PCR were as follows: p21 sense 5'-GGC AGA CCA GCA TGA CAG ATT TC-3', antisense 5'-CGG ATT AGG GCT TCC TCT TGG-3'; p8 sense 5'-CTA TAG CCT GGC CCA TTC CT-3', antisense 5'-TCT CTC TTG GTG CGA CCT TT-3'; sestrin-2 sense 5'-CAA GCT CGG AAT TAA TGT GCC-3', antisense 5'-CTC ACA CCA TTA AGC ATG GAG-3'; and TATA-binding protein sense 5'-TGC ACA GGA GCC AAG ATG GAA-3', antisense 5'-CAC ATC ACA GCT CCC CAC CA-3'. The PCR primers for CSE, NAG1, and Nur77 were purchased from Qiagen.

Microarray experiments. Microarray studies focused on early-induced genes, and UC-5 cells were treated with DMSO or 15 μ M DIM-C-pPhOCH₃ for 2 and 6 hr. RNA was isolated as described for the RT-PCR experiment and analyzed for gene expression using the Codelink Whole Genome Bioarrays (300026), and three replicates were determined for each time point and the DMSO control. The microarray data were

analyzed using GeneSpring software version 7.2 (Agilent, Palo Alto, CA). The data were normalized in two steps. First, for each array, the expression value of each gene was divided by the median of all the values in that array. Second, for each gene, the expression value in each array was divided by the median value of that gene across all arrays. Genes with low quality signals were excluded for statistical analysis. One-way ANOVA (assume equal variances) was carried out to identify differentially expressed genes. A gene was said to be differentially expressed if the Benjamini and Hochberg adjusted p-values were less than 0.05.

Xenograft studies in athymic mice. Male athymic nude mice (*Foxn1*^{nu}, ages 7-8 weeks) were purchased from Harlan (Indianapolis, IN). The mice were housed and maintained in laminar flow cabinets under specific pathogen-free conditions. A xenograft was established by s.c. injection of *in vitro* cultured KU-7 cells (1×10^7 cells/150 μ L) into the flanks of individual mice. Tumors were allowed to grow for 7 days until tumors were palpable. Mice were then randomized into two groups of 5 mice per group and dosed by oral gavage with either corn oil or 25 mg/kg/day DIM-C-pPHOCH₃ for 17 days. The mice were weighed and tumor size was measured twice a week with calipers to permit calculation of tumor volumes, $V=L*W^2/2$, where L and W were length and width. Final body, organ and tumor weights were determined at the end of the dosing regimen, and both organ and tumor blocks were obtained for hematoxylin and eosin-staining and histopathological analysis.

Terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling (TUNEL) assay. For the TUNEL assay, tumor tissue was fixed in formalin and embedded in paraffin, TUNEL staining was carried out using DeadEnd Colorimetric

TUNEL System (Promega). Paraffin-embedded sections (4-6 μm thick) were processed per manufacturer's protocol. Briefly, sections were deparaffinized in xylene and then treated with a graded series of alcohol [100%, 95%, 85%, 70%, and 50% ethanol (v/v) in double-distilled water] and rehydrated in PBS (pH 7.5). Tissues were then treated with proteinase K solution for permeabilization and then refixed with 4% paraformaldehyde solution. Slides were then treated with recombinant terminal deoxynucleotidyl transferase reaction mix and incubated at 37°C for 1 hr. Reaction was terminated by immersing the slides in 2X SSC solutions for 15 min at room temperature. After blocking the endogenous peroxidases activity (by 0.3% hydrogen peroxide), slides were washed with PBS and then incubated with streptavidin horseradish peroxidase solution for 30 min at room temperature. After washing, slides were incubated with 3,3'-diaminobenzidine (substrate) solution until a light brown background appeared (10 min) and then rinsed several times in deionized water. After mounting, slides were observed by light microscope.

Statistical analysis. Statistical significance was assessed using Student's t-test. A value of $P < 0.05$ compared with solvent control was considered statistically significant.

RESULTS

The potential application of Nur77-active C-DIMs for bladder cancer chemotherapy is dependent on expression of this orphan receptor in bladder tumor tissue. Figure 1 summarizes immunostaining of Nur77 in bladder tumors from patients. Nur77 staining was observed in low grade non-invasive tumors (Fig. 1A), in high grade stage T2 bladder cancer (Fig. 1B), and in high grade stage T3 bladder cancer (Fig. 1C) growing in adipose tissue outside the muscle of the bladder. This pattern of Nur77 staining was observed in multiple bladder tumor samples, whereas minimal to non-detectable staining of Nur77 was detected in non-tumor tissue. In addition, expression of Nur77 was also investigated in low grade RT4 and high grade invasive UC-14 tumors established in nude mice (Fig. 1D). Sections from the low grade well-differentiated RT4 tumors growing in the wall of the mouse bladder expressed Nur77 throughout the tumor, whereas staining for this protein in non-tumor tissue was not detected. Nur77 staining was also detected in the UC-14 cells which are shown invading the wall of the mouse bladder, whereas Nur77 was not detected in non-tumor tissue. Previous studies show that western blot analysis of whole cell lysates from several bladder cancer cell lines, including KU7 and UC-5 cells, show that all of these cell lines express Nur77 protein (Inamoto et al., 2008).

Figures 2A and 2B illustrate the concentration-dependent effects of DIM-C-pPhOCH₃ and DIM-C-pPh (5, 10, and 15 μ M) on proliferation of UC-5 and KU7 bladder cancer cell lines, respectively. Both compounds inhibited growth of UC-5 and KU7 cells within 24 hr after treatment, and the magnitude of growth inhibition was higher after 48 hr. IC₅₀ (48 hr) values for DIM-C-pPhOCH₃ in UC-5 and KU7 cells were 12.3 μ M and

9.3 μM , and the corresponding values for DIM-C-pPh were 11.8 μM and 11.2 μM , respectively.

We further investigate the subcellular location of this receptor in the absence or presence of DIM-C-pPhOCH₃. Figure 3A illustrates the expression of Nur77, PARP, and Sp1 proteins in cytosolic (C) or nuclear (N) fractions in untreated cells and cells treated with 15 μM DIM-C-pPhOCH₃, 0.05 ng/ml leptomycin B, or their combination for 24 hr. In all treatment groups, Nur77, PARP (full length or cleaved) and Sp1 proteins were observed only in the nuclear fraction and this is consistent with the subcellular nuclear location of these proteins. Moreover, treatment with DIM-C-pPhOCH₃ did not change the subcellular location of these proteins. A non-specific band was observed in both cytosolic and nuclear fractions. DIM-C-pPhOCH₃ also induced caspase-dependent PARP cleavage and this was observed in the presence or absence of leptomycin B which inhibits nuclear export, suggesting that induction of caspase-dependent PARP cleavage by DIM-C-pPhOCH₃ was associated with nuclear Nur77 and did not require nuclear export of the receptor. The nuclear location of Nur77 in UC-5 cells treated with DMSO or 15 μM DIM-C-pPhOCH₃ was also confirmed by immunostaining for Nur77 which was located in the nucleus of these cells (Fig. 3B). Both DIM-C-pPhOCH₃ and DIM-C-pPh induced PARP and caspase-8 cleavage in UC-5 and KU7 cells after treatment for 24 hr and the decreased apoptosis using 15 μM DIM-C-Ph in KU7 cells was due to cytotoxicity in this cell line (Fig. 3C). In addition, DIM-C-pPhOCH₃- and DIM-C-pPh-induced PARP cleavage in UC-5 cells was inhibited after cotreatment with the pancaspase inhibitor Z-VAD-fmk (Fig. 3D).

Previous studies in pancreatic and colon cancer cells showed that DIM-C-pPhOCH₃ induced TRAIL and the extrinsic pathway of apoptosis (Chintharlapalli et al., 2005a; Cho et al., 2007). Since Nur77-active C-DIMs activated caspase-8, we also investigated activation of TRAIL in UC-5 cells, and treatment with 15 μ M DIM-C-pPhOCH₃ or DIM-C-pPh for 48 hr induced TRAIL protein (Fig. 4A). Similar results were observed for TRAIL mRNA levels (data not shown). The role of Nur77 in mediating the induction of TRAIL and apoptosis in UC-5 cells by DIM-C-pPhOCH₃ was investigated by transient transfection of a small inhibitory RNA for Nur77 (iNur77) as previously described (Cho et al., 2007). Cells were transfected with a non-specific oligonucleotide (iScr) or iNur77 and treated with DMSO or 15 μ M DIM-C-pPhOCH₃, and whole cell lysates were analyzed by western blot analysis. RNA interference decreased Nur77 expression in the DMSO and DIM-C-pPhOCH₃ treatment groups by approximately 60% in whole cell lysates (Fig. 4B), indicating that both transfection efficiencies and knockdown of Nur77 by iNur77 were high. In a parallel experiment, RNA interference in UC-5 cells transfected with iNur77 and treated with DMSO or 15 μ M DIM-C-pPhOCH₃ also decreased TRAIL expression and PARP cleavage by 45-50% (Fig. 4C), confirming that these responses were also Nur77-dependent.

Although DIM-C-pPhOCH₃ induced apoptosis through Nur77-dependent activation of TRAIL in UC-5 cells, this compound activates Nur77-dependent and Nur77-independent growth inhibition and proapoptotic genes in other cancer cell lines (Chintharlapalli et al., 2007; Cho et al., 2008; Lei et al., 2006). Therefore, we treated UC-5 bladder cancer cells for 2 or 6 hr with DMSO or 12.5 μ M DIM-C-pPhOCH₃ and performed microarray analysis (3 arrays for each treatment group using the CodeLink

Human Whole Genome Bioarrays). Results for DIM-C-pPhOCH₃ were compared to a DMSO control at each time point, and the microarray data were analyzed using GeneSpring software version 7.2 (Agilent, Palo Alto, CA). Several growth inhibitory and proapoptotic genes were induced by DIM-C-pPhOCH₃ after treatment for 2 or 6 hr, and these include the following genes (fold induction after 2 or 6 hr indicated in brackets): nonsteroidal anti-inflammatory drug activated gene (NAG-1) (5.58 and 22.70); p8 (1.50 and 5.88); cystathionase (0.97 and 5.02); sestrin-2 (1.65 and 4.59); and p21 (2.01 and 2.28) (Supplemental Table 1). In addition, we also observed activation of genes (by \geq 2.5-fold) associated with metabolism and homeostasis, signal transduction, transcription, protein folding and modification, and genes with several other functions (Supplemental Table 1). Results in Figure 5A illustrate that DIM-C-pPhOCH₃ significantly induced NAG-1, p8, CSE, and sestrin-2 mRNA levels in UC-5 cells as determined by semiquantitative RT-PCR, and replicates of these experiments confirm the significance of these induction responses (Fig. 5B). DIM-C-pPhOCH₃ and other C-DIMs induce receptor-independent responses in cancer cells (Chintharlapalli et al., 2007; Cho et al., 2008; Lei et al., 2006) and therefore we investigated the Nur77-dependent effects of DIM-C-pPhOCH₃ on induction of several genes in UC-5 cells transfected with iScr or iNur77. DIM-C-pPhOCH₃ significantly induced CSE, NAG-1, p21, p8, and sestrin-2 and, in the presence of iNur77, there was a significant decrease in CSE, p21, p8, and sestrin-2 mRNA levels, whereas NAG-1 mRNA levels were not affected by Nur77 knockdown. Levels of Nur77 mRNA were not affected by DIM-C-pPhOCH₃; however, in UC-5 cells transfected with iNur77, there was decreased expression of Nur77 mRNA in

the control and DIM-C-pPhOCH₃ treated groups. This study identifies CSE, p21, p8, and sestrin-2 as Nur77-regulated genes in bladder cancer cells.

The *in vivo* antitumorigenic activity of DIM-C-pPhOCH₃ was determined in athymic nude mice bearing KU7 cells as xenografts. At doses of 25 mg/kg, DIM-C-pPhOCH₃ significantly inhibited tumor volumes and weights compared to the corn oil treated solvent controls (Figs. 6A and 6B). Histopathological examination of the treated animals did not reveal any significant pathology and changes in body or organ weights were not observed (data not shown). In addition, the TUNEL assay for apoptosis showed enhanced staining (Fig. 6C), and extraction of proteins from tumor tissue obtained from corn oil and DIM-C-pPhOCH₃-treated mice showed that in the latter treatment group there was a significant increase in PARP cleavage (Fig. 6D). These results demonstrate that Nur77-active C-DIMs inhibit bladder cancer cell and tumor growth and induce apoptosis in both *in vitro* and *in vivo* models.

DISCUSSION

Nur77 is an orphan nuclear receptor that plays a unique role in mediating the proapoptotic activity of a wide variety of agents (Chintharlapalli et al., 2005a; Cho et al., 2007; Holmes et al., 2003a; Holmes et al., 2003b; Li et al., 2000; Lin et al., 2004; Mu and Chang, 2003; Wilson et al., 2003a; Wu et al., 2002). There are differences in the proposed mechanisms of action of various apoptosis inducers in mediating induction of Nur77-dependent cell death pathways which may involve interactions of this receptor with mitochondrial or cytosolic proteins. For example, in different cell lines treated with retinoids or phorbol ester, there is evidence that Nur77 translocates from the nucleus to the mitochondria and forms a proapoptotic complex with bcl-2 (Li et al., 2000; Lin et al., 2004). In all cases where Nur77-dependent apoptosis requires extranuclear transport, the effects of the apoptosis-inducing agents are inhibited after cotreatment with leptomycin B, a nuclear export inhibitor. It has also been reported that phorbol ester induced expression of nuclear Nur77 in LNCaP prostate cancer cells and this resulted in enhanced expression of E2F1 which exhibits proapoptotic activity (Mu and Chang, 2003). In contrast, the Nur77-active C-DIMs used in this study did not induce E2F1 in bladder cancer cells (data not shown) or in pancreatic or colon cancer cells (Chintharlapalli et al., 2005a; Cho et al., 2007). Both DIM-C-pPhOCH₃ and DIM-C-pPh represent a novel class of Nur77-active proapoptotic compounds. The Nur77-active C-DIMs decrease bladder cancer cell proliferation (Fig. 2A) and induce apoptosis in these cells through nuclear Nur77-dependent pathways that are not inhibited by leptomycin B (Fig. 3A).

Previous studies on Nur77-active C-DIMs in pancreatic and colon cancer cells show that DIM-C-pPhOCH₃ induces both receptor-dependent and -independent apoptosis (Chintharlapalli et al., 2005a; Cho et al., 2008; Cho et al., 2007), and the latter pathways are due to several factors including induction of other proapoptotic genes such as NAG-1. DIM-C-pPhOCH₃ and other C-DIMs also induce endoplasmic reticulum stress which results in activation of death receptor 5 and this response is also receptor-independent (Abdelrahim et al., 2006; Chintharlapalli et al., 2005a; Lei et al., 2008a; Lei et al., 2008b). Results illustrated in Figure 4 demonstrate that DIM-C-pPhOCH₃ induces the death receptor ligand TRAIL (Fig. 4A) as previously reported in colon and pancreatic cancer cells (Chintharlapalli et al., 2005a; Cho et al., 2007). Nur77 knockdown in UC-5 cells was determined by transient transfection of iNur77, and the results obtained using whole cell lysates suggest that transfection efficiencies were high and resulted in significant knockdown of Nur77 protein. Based on the comparison of the relative effects of iNur77 on Nur77, PARP cleavage, and decreased TRAIL protein expression (Figs. 4B-4C), the results suggest that induction of apoptosis by DIM-C-pPhOCH₃ in UC-5 cells was primarily Nur77-dependent. Previous studies in pancreatic and colon cancer cell lines suggest that the contribution of Nur77-activated apoptosis by C-DIMs is variable and cell context dependent (Chintharlapalli et al., 2005a; Cho et al., 2007), and current studies are investigating this variability in other bladder cancer cell lines.

We also have used microarrays to investigate induction of gene expression by DIM-C-pPhOCH₃ in bladder cancer cells, and we observed induction of several growth inhibitory and proapoptotic genes. A comparison of the effects of DIM-C-pPhOCH₃ on patterns of gene expression in bladder, colon, pancreatic, and prostate cancer cells and

among the several hundred genes induced in these cell lines, there are only 19 genes induced in all 4 cell lines (data not shown). Two of these genes, NAG-1 and sestrin-2, are growth inhibitory/proapoptotic and are among the more highly induced in UC-5 cells (Fig. 5 and Supplemental Table 1). Among the 5 genes that we have examined by real-time PCR (Fig. 5C), results of RNA interference studies demonstrate that induction of NAG-1 by DIM-C-pPhOCH₃ was Nur77-independent and these results are consistent with previous studies showing that NAG-1 is induced by structurally-diverse compounds including some nuclear receptor agonists, and induction of this gene is receptor-independent (Baek et al., 2004a; Baek et al., 2004b; Baek et al., 2001; Baek et al., 2002; Chintharlapalli et al., 2005b; Lee et al., 2005; Wilson et al., 2003b). However, we also observed that genes involved in cell cycle regulation and arrest (sestrin-2 and p21), stress and apoptosis (p8), and growth inhibition via hydrogen sulfide release (cystathionase) (Budanov and Karin, 2008; Carracedo et al., 2006; Gartel and Tyner, 1999; Yang et al., 2004) are induced by DIM-C-pPhOCH₃ in a Nur77-dependent manner (Fig. 5C) and thereby represent novel genes induced by nuclear Nur77. In pancreatic cancer cells, we have shown that Nur77-dependent induction of p21 involves interactions with specificity proteins bound to proximal GC-rich regions of the p21 promoter (Lee et al., 2009). The molecular mechanisms of transactivation and interactions of Nur77 with *cis* elements in gene promoters are currently being investigated.

It is clear from results of *in vivo* studies that DIM-C-pPhOCH₃ also inhibits bladder tumor growth in a mouse xenograft model with KU7 cells (Fig. 6). Analysis of the tumor tissue demonstrates that DIM-C-pPhOCH₃ induces apoptosis in tumors and

this complements *in vivo* studies in cell culture (Fig. 3). These data, coupled with expression of Nur77 in bladder cancer cells (Inamoto et al., 2008) and human tumors (Fig. 1) suggests that this receptor is a potential target for bladder cancer chemotherapy. Ongoing research is focused on development of additional C-DIM analogs that activate Nur77 and other NGFI-B receptors and their application for treatment of different stages of bladder cancer.

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FOOTNOTES

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FIGURE LEGENDS

Fig. 1. Nur77 expression in human bladder tumors and bladder cancer cell lines.

Expression of Nur77 in low grade (A), high grade stage 2 (B), and high grade stage 3 (C) bladder tumors, and in RT4 and UC-14 (D) bladder cancer cell lines were determined by immunostaining with Nur77 antibodies as outlined in the Materials and Methods.

Fig. 2. Nur77-active C-DIMs decrease cell proliferation in UC-5 and KU7 cells. UC-5

cells (A) and KU7 cells (B) were treated with DMSO or different concentrations of C-DIMs (5, 10 and 15 μ M) for 24 and 48 hr, and the percentages of cells in the treatment groups compared with the solvent control (DMSO; set at 100%) were determined. Columns, mean for three replicate experiments for each treatment group; bars SD. *, $p < 0.05$ compared to DMSO treatment group.

Fig. 3. Nur77-active C-DIMs induce apoptosis in UC-5 and KU7 cells and also act

through nuclear Nur77. (A) Effects of leptomycin B on PARP cleavage. UC-5 cells were treated for 24 hr with DIM-C-pPhOCH₃ alone or in the presence of leptomycin B (0.05 ng/ml), and western blot analysis of nuclear and cytosolic cell lysates were used to detect Nur77, PARP cleavage, Sp1 (loading control for nuclear fraction), N.S. (non-specific band for loading control). (B) Immunostaining for Nur77. UC-5 cells were treated with DMSO or 15 μ M of DIM-C-pPhOCH₃ for 24 hr and immunostained with IgG or Nur77 antibodies. (C) Dose-dependent induction of apoptosis by C-DIMs. UC-5 and KU7 cells were treated with DMSO or different concentrations of DIM-C-pPhOCH₃ or DIM-C-Ph for 48 hr. (D) Inhibition of Nur77-active C-DIMs-induced apoptosis by Z-VAD-fmk.

UC-5 cells were treated with Nur77-active C-DIMs alone or in combination with 10 μ M of Z-VAD-fmk, and whole-cell lysates were analyzed by western blot analysis.

Fig. 4. Nur77-dependent induction of TRAIL and apoptosis in UC-5 cells. (A) Induction of TRAIL by DIM-C-pPhOCH₃ and DIM-C-Ph. Cells were treated with Nur77-active C-DIMs (15 μ M) for 24 hr, and whole-cell lysates were analyzed for TRAIL or β -tubulin. (B) and (C) RNA interference with iNur77. UC-5 cells were transfected with iScr (nonspecific) or iNur77 and treated with DMSO or 15 μ M of DIM-C-pPhOCH₃, and whole-cell lysates were analyzed by western blot analysis for Nur77, PARP cleavage, TRAIL and β -tubulin proteins. Columns, mean for three replicate experiments for each treatment group; bar, SD. Protein levels were normalized to β -tubulin. (B) *, $P < 0.05$, significant inhibition of Nur77 expression. (C) *, significant induction of TRAIL and PARP cleavage; **, decreased expression of these proteins by iNur77.

Fig. 5. Induction of gene expression by DIM-C-pPhOCH₃. (A) and (B) Induction of NAG-1, p8, CSE, and sestrin-2 in UC-5 cells. Cells were treated with DMSO or 15 μ M of DIM-C-pPhOCH₃ for 6 hr, and NAG-1, p8, CSE, sestrin-2 and GAPDH (reference mRNA) were determined by RT-PCR. Induction responses were determined in three separate experiments. *, $P < 0.05$, significantly induced mRNA expression. (C) Effects of iNur77 on induced gene expression. UC-5 cells were transfected with iScr or iNur77, treated with DMSO or 15 μ M of DIM-C-pPhOCH₃ for 6 hr and analyzed by real-time PCR. Columns, mean for three replicate determinations for each treatment group; bars, SD. *, $P < 0.05$,

significant induction by DIM-C-pPhOCH₃; #, significant decrease after transfection with iNur77.

Fig. 6. Induction of tumor growth by DIM-C-pPhOCH₃. Male athymic nude mice bearing KU7 cell xenografts were treated with DIM-C-pPhOCH₃, and tumor volumes (A) and weight (B) were determined. The compound was given daily (25 mg/kg/day) in corn oil by oral gavage, and corn oil served as a solvent control. (C) Immunostaining for apoptosis. Tumor tissue from animals treated with solvent control or DIM-C-pPhOCH₃ were stained, and a TUNEL assay was used to detect apoptosis. (D) Whole-cell lysates from tumor tissues was analyzed by western blot analysis using PARP antibody to detect apoptosis. Columns, mean for five replicate determinations for each group; bars SE. *, P<0.05, significant induction by DIM-C-pPhOCH₃.

Figure 1

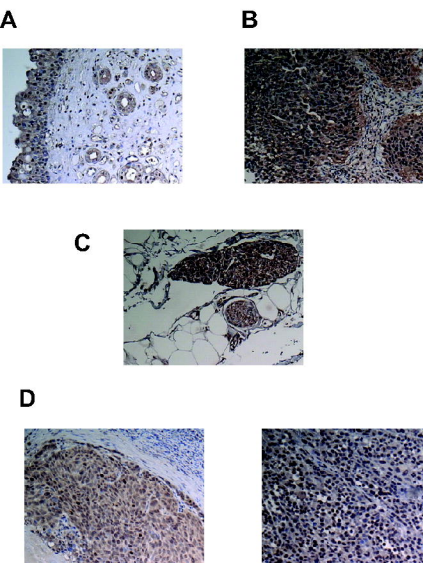
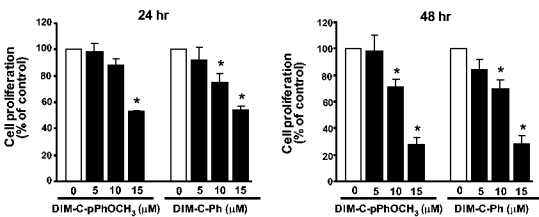


Figure 2

A

UC -5 cells



B

KU -7 cells

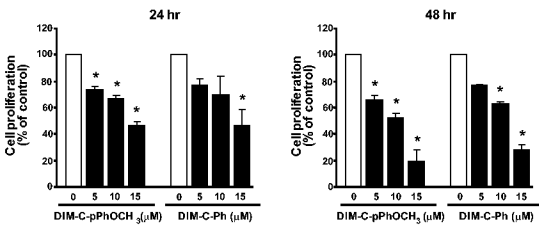
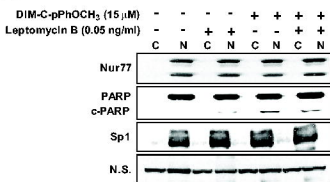
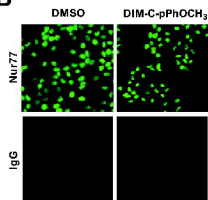


Figure 3

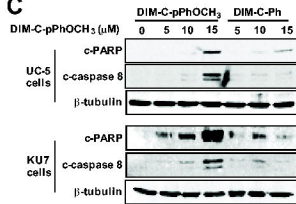
A



B



C



D

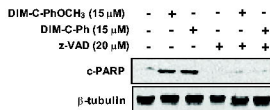
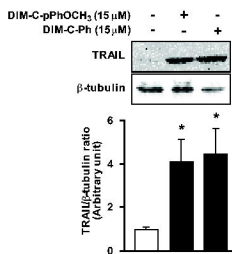
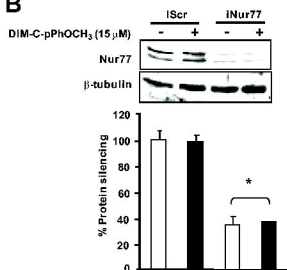


Figure 4

A



B



C

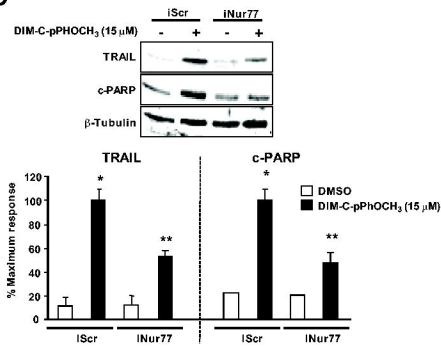


Figure 5

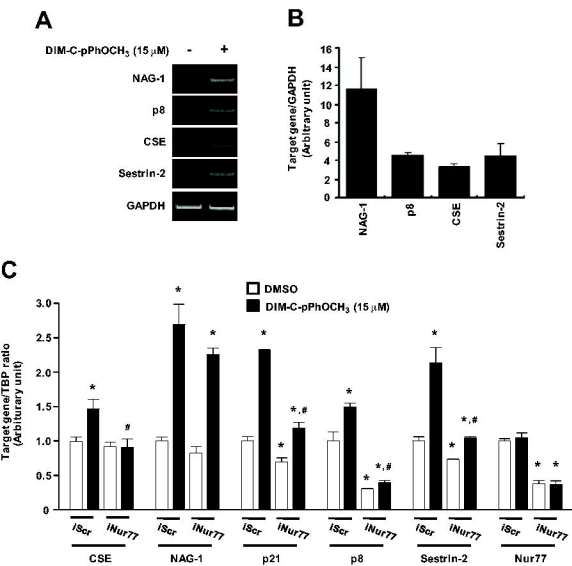


Figure 6

