Development of a selective and high affinity radioligand, 

$[^3]H]VU6013720$, for the M₄ muscarinic receptor

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Abbreviations:

- $B_{\text{max}}$: maximal binding
- BCA: bicinchoninic acid
- CHO-K1: Chinese Hamster Ovary K1
- CO$_2$: carbon dioxide
- CNS: central nervous system
- FBS: fetal bovine serum
- HEPES: 4-(2-hydroxyethyl)-1-piperazine ethanesulfonic acid
- IC$_{50}$: inhibitory concentration 50
- $K_d$: equilibrium dissociation constant
- $K_i$: equilibrium inhibitory constant
- $K_{\text{off}}$: dissociation off rate
- $K_{\text{on}}$: association on rate
- nM: nanomolar
- NMS: N-methyl scopolamine
- PAM: positive allosteric modulator
- PET: Positron Emission Tomography
Abstract

M4 muscarinic receptors are highly expressed in the striatum and cortex, brain regions that are involved in diseases such as Parkinson’s disease, schizophrenia, and dystonia. Despite potential therapeutic advantages of specifically targeting the M4 receptor, it has been historically challenging to develop highly selective ligands, resulting in undesired off-target activity at other members of the muscarinic receptor family. Recently, we have reported first-in-class, potent, and selective M4 receptor antagonists. As an extension of that work, we now report the development and characterization of a radiolabeled M4 receptor antagonist, [3H]VU6013720, with high affinity (pKd of 9.5 ± 0.2 at rat M4, 9.7 at mouse M4, and 10 ± 0.1 at human M4 with atropine to define nonspecific binding) and selectivity (80-fold over M2 and >1,000 fold over M1, M3 and M5) for the M4 receptor versus the other muscarinic subtypes. Binding assays using this radioligand in rodent brain tissues demonstrate loss of specific binding in Chrm4 knockout animals. Dissociation kinetics experiments with various muscarinic ligands show differential effects on the dissociation of [3H]VU6013720 from M4 receptors, suggesting a binding site that is overlapping but may be distinct from the orthosteric site. Overall, these results demonstrate that [3H]VU6013720 is the first highly selective antagonist radioligand for the M4 receptor, representing a useful tool for studying the basic biology of M4 as well for the support of M4 receptor-based drug discovery.
Significance statement: This manuscript describes the development and characterization of a novel muscarinic acetylcholine subtype 4 receptor antagonist radioligand, [3H]VU6013720. This ligand binds to or overlaps with the acetylcholine binding site, providing a highly selective radioligand for the M₄ receptor that can be used to quantify M₄ protein expression in vivo and probe the selective interactions of acetylcholine with M₄ versus the other members of the muscarinic receptor family.
Introduction

The five subtypes of muscarinic (M) acetylcholine receptors mediate a wide variety of physiological functions and pathophysiological processes. The M₁, M₃, and M₅ subtypes are preferentially coupled to G₉/₁₁ G proteins and activate phospholipase C, which catalyzes the phosphatidylinositol trisphosphate cascade and leads to activation of protein kinase C and intracellular Ca²⁺ mobilization. In contrast, the M₂ and M₄ subtypes are coupled to Gᵢₒ and inhibit adenylyl cyclase activity and reduce cyclic AMP production upon activation; additionally, signaling downstream of these receptors also modulates ion channels (Ashkenazi et al., 1988, Peralta et al., 1988, Offermanns et al., 1994, Ishii et al., 2006, Lebois et al., 2018). Muscarinic acetylcholine receptor subtypes are distributed in both the central nervous system (CNS) and periphery. Although non-selective muscarinic receptor antagonists have shown therapeutic potential for the treatment of certain neurological disorders, usage has been limited due to intolerable side effects resulting from their broad inhibition of all the muscarinic receptor subtypes (Fahn et al., 1990, Cloud et al., 2010). This has led to the development of subtype-selective ligands to improve efficacy while reducing side effects.

Of the muscarinic receptors, the M₄ subtype is most highly expressed in striatum, cortex, and some subcortical regions such as the thalamus (Brann et al., 1988, Buckley et al., 1988, Weiner et al., 1990). It has been reported that M₄ is the primary muscarinic receptor subtype responsible for regulating dopamine signaling and release in the basal ganglia (Gomeza et al., 1999, Jeon et al., 2010, Foster et al., 2016, Moehle et al., 2017, Moehle et al., 2019) and M₄ has been implicated in several neurological disorders such
as Parkinson’s disease, schizophrenia, and dystonia (Katzenschlager et al., 2003, Moehle et al., 2019, Moran et al., 2019).

Despite these biological observations regarding the therapeutic potential of M4 modulation, development of highly selective M4 receptor orthosteric and allosteric modulators has been challenging, and, by extension, the development of radioligands specifically targeting M4 has historically been difficult. Recently, two M4 receptor positive allosteric modulators (PAMs), \[^{11}C\]-VU0467485/AZ13713945 (Deng et al., 2019) and \[^{18}F\]-M4R-1911 (Deng et al., 2020) have been radiolabeled and demonstrated to specifically bind to M4 receptors. Another M4 PAM radioligand, \[^{18}F\]\_12, has also been shown to display species’ differences in terms of its binding specificity and selectivity (Haider et al., 2023). Encouragingly, the M4 receptor PAM radioligand \[^{11}C\]MK-6884 has been recently reported to specifically bind to the M4 receptor and penetrate into the brain and has been used clinically as a Positron Emission Tomography (PET) ligand (Tong et al., 2020, Li et al., 2022). In this report, we complement these selective M4 PAM radioligands with \[^{3}H\]VU6013720, a highly selective M4 antagonist radioligand.

Recently, our group successfully synthesized and characterized several highly selective M4 receptor antagonists, VU6013720, VU6021302, and VU6021625 (Moehle et al., 2021), VU6028418 (Spock et al., 2021), and VU6015241 (Bender et al., 2022). Of these compounds, VU6013720 exhibited the best in vitro functional potency (rat M4 IC\(_{50}\) = 20 nM, human M4 IC\(_{50}\) = 0.6 nM from calcium mobilization assays) as well as a good selectivity profile among the muscarinic receptor family, with IC\(_{50}\) values of >10,000 nM at rM3 and rM5. At rM1, VU6013720 exhibited a functional IC\(_{50}\) of 1700 nM, displaying ~85-fold selectivity, while VU6013720 had an IC\(_{50}\) of 670 nM at rM2, displaying ~34-fold
selectivity. Additionally, the compound exhibited unbound percentages of 24% and 31% in rat and mouse plasma, as well as 5% in both rat and mouse brain tissue.

In the current report, we generated a tritiated form of this compound, \(^{3}\text{H}\)VU6013720. As shown here, \(^{3}\text{H}\)VU6013720 selectively binds to \(M_4\) receptors with little to no binding to the \(M_1\), \(M_2\), \(M_3\), and \(M_5\) receptor subtypes. Binding assays with brain tissue homogenates reveal that \(^{3}\text{H}\)VU6013720 exhibits specific binding in extracts from cortex and striatum. As other existing radioligands bind to allosteric sites, it is anticipated that a selective antagonist radioligand that interacts with the acetylcholine binding site should provide a valuable tool to explore selective binding of acetylcholine to the \(M_4\) receptor in native tissues, \textit{in vivo}, in physiological and pathophysiological disease states and for selective \(M_4\) receptor drug development.

**Materials and Methods**

**Cell culture and membrane preparation.** Chinese Hamster Ovary (CHO-K1) cells were purchased from the American Tissue Culture Collection (ATCC, Manassas, Virginia). CHO-K1 cells stably express rat, human, or mouse muscarinic receptors (\(M_1\)–\(M_5\)); for \(M_2\) and \(M_4\), cells were co-transfected with \(G_{q15}\). Cells were cultured in F12 medium containing 10% fetal bovine serum, 20 mM HEPES, and 1X Antibiotic/Antimycotic (contains 10,000 units/mL of penicillin, 10,000 \(\mu\)g/mL of streptomycin, and 25 \(\mu\)g/mL of Gibco Amphotericin B; Thermo Fisher Scientific, Waltham, MA) at 37°C in the presence of 5% CO\(_2\). Cells were monitored by periodical PCR detection using a LookOut Mycoplasma PCR Detection Kit (Sigma-Aldrich, St
Louis, MO) to eliminate potential mycoplasma infection and cells were not mycoplasma positive for the studies described here.

Cell membranes were made as previously reported (O’Brien et al., 2018, Moehle et al., 2021). CHO-K1 cells stably expressing different muscarinic receptor subtype were seeded in 150-mm dishes and allowed to grow to approximately 80%–90% confluence. Cells were washed once with ice-cold phosphate-buffered saline, scraped from the dishes, and collected using a 5-minute centrifugation at 1000g. The supernatant was removed, and the cell pellet was stored at -80°C. To prepare membrane protein, cell pellets were thawed, re-suspended in membrane binding buffer (20 mM HEPES with 10 mM MgCl₂ and 10 mM NaCl, pH 7.4), and homogenized using two 15-second bursts from a TR-10 polytron (Tekmar, Vernon, BC, Canada). Cell lysates were then centrifuged at 1000g for 10 minutes at 4°C, and the pellet was discarded. The resultant supernatant was centrifuged at 20,000g for 30 minutes at 4°C and the supernatant was discarded. The remaining pellet was re-suspended in membrane binding buffer. The protein concentration was determined by BCA assay (Thermo Scientific, Rockford, IL). Membranes were then stored at -80°C until use.

Radiochemistry. The synthesis of VU6013720 has been reported previously (Moehle et al., 2021). Cold VU6013720 compound was radiolabeled by RC TriTec (Teufen, Switzerland) with photocatalytic H/T exchange (Figure 1), with the [³H] label most likely in the alpha position of the tertiary amine. The final radioligand, [³H]VU6013720, had a purity > 98% with a specific activity 23.6 Ci/mmol (233.8 GBq/mmol) and a concentration of 1 mCi/ml (37 MBq/ml). [³H]NMS was manufactured
on 03/08/2018, Specific activity: 80.0 Ci/mmol, Dilution: 1mCi/ml (PerkinElmer, Boston, MA).

**[^3]H]VU6013720 saturation binding assays.** Saturation binding experiments were performed to determine the specificity of[^3]H]VU6013720 binding to the muscarinic receptor subtypes. Briefly, 10 µg CHO-K1 cell membranes expressing each receptor were incubated with[^3]H]VU6013720 in deep 96-well plates (Corning, New York) on a shaker at room temperature for 3 hours in a reaction volume of 500 μL. Saturation binding experiments were performed in triplicate or duplicate, and nonspecific binding was assessed by addition of 10 μM atropine or VU6013719 (VU6013720 structural analogue, Supp. Fig. 1). The reactions were terminated by vacuum filtration onto Unifilter-96 GF/B plate through a 96-well harvester. The filters were washed three times with cold membrane harvesting buffer (50 mM Tris-HCl and 0.9% NaCl, pH 7.4). Plates were then dried overnight and 40 µl of MicroScint 20 (PerkinElmer) was added to each well. The membrane-bound radioactivity was determined using a TopCount NXT microplate scintillation and luminescence counter (PerkinElmer). Calculation of specific binding versus total counts revealed that radioligand depletion was not a factor in these studies using this assay protocol.

**Competition binding assays with[^3]H]NMS and[^3]H]VU6013720.** Radioligand competition binding assays were performed as previously described (Moehle et al., 2021) using concentrations of radioligand that were within two-fold of the calculated K_d. Briefly, M_4 antagonists were 1:3 serially diluted into membrane binding buffer and added to each well of a 96-well plate, along with 10 µg/well cell membrane and approximately 100 pM[^3]H-N-methylscopolamine ([^3]H]NMS, PerkinElmer, Boston, MA)
or 100 - 400 pM $[^3H]VU6013720$ in a reaction volume of 500 μL. Following a 3-hour incubation period on a shaker at room temperature, the membrane-bound ligand was separated from free ligand by filtration through glass fiber 96-well filter plates (Unifilter-96, GF/B; PerkinElmer, Boston, MA) using a 96-well Brandel harvester. The filters were washed three times with cold membrane harvesting buffer (50 mM Tris-HCl and 0.9% NaCl, pH 7.4). Plates were then dried overnight and 40 µl of MicroScint 20 (PerkinElmer) was added to each well. The membrane-bound radioactivity was determined using a TopCount NXT microplate scintillation and luminescence counter (PerkinElmer). Nonspecific binding was determined using 10 μM atropine.

**Kinetic studies.** Kinetic studies of $[^3H]VU6013720$ association/dissociation from M₄ receptors were performed at room temperature. For association assays, membranes were incubated with $[^3H]VU6013720$ at concentrations 2-fold over the $K_d$ value for 0, 2, 5, 10, 20, 30, 45, 60, 90, 120, 150, and 180 minutes. For dissociation assays, membranes and $[^3H]VU6013720$ were allowed to equilibrate for 120 minutes and then 10 μM of cold competitor was added at 120, 90, 60, 45, 30, 20, 15, 10, 5, 2, and 1 minute before termination of the assay. Following these pretreatment times, membranes were immediately harvested by quick filtration to a GF/B plate using ice-cold harvesting buffer and a 96-well Brandel harvester. Buffer and temperature conditions for all binding studies were the same.

**Animals.** Male and female wildtype (2 male and 2 female) or Chrm4 knockout (2 male and 2 female) mice (bred in-house) and male Sprague Dawley (n=15) rats (Envigo, Indianapolis, IN) were used for this study. They were housed in the animal care facility certified by the American Association for the Accreditation of Laboratory Animal
Care (AALAC) under a 12-hour light/dark cycle (lights on: 7 a.m.; lights off: 7 p.m.). The experimental protocols performed during the light cycle were approved by the Institutional Animals Care and Use Committee of Vanderbilt University and conformed to the guidelines established by the National Research Council Guide for the Care and Use of Laboratory Animals.

**Brain tissue membrane preparation and in vitro saturation binding.** Rats and mice (wild-type and Chrm4 knockout) were anesthetized by continuous isoflurane (5%) and killed by decapitation, and striatum and cortex were dissected on ice. The dissected brain regions were weighed and homogenized at a concentration of 1 mg of tissue per 25 volumes of membrane binding buffer with a polytron for two 20-second bursts. The homogenates were centrifuged at 20,000g for 20 minutes at 4°C, and the supernatant was discarded. The pellet was homogenized in membrane binding buffer and centrifuged under the same conditions as above. The final homogenate was re-suspended in membrane binding buffer and incubated at 37°C for 10 minutes and centrifuged as above. The resultant pellet was re-suspended in membrane binding buffer, and the protein concentration was determined by BCA assay. The brain homogenate was stored at -80°C until use. To determine the specific binding of [³H]VU6013720 using brain tissue membranes, saturation assays were performed as above with CHO cell membranes but using 150 µg of brain homogenate protein per well.

**Data analysis.** All experiments were carried out in duplicate or triplicate and were performed at least three times (unless stated otherwise), which aligns with our historical and standard assay protocol. Studies presented in this manuscript are
exploratory in nature and were performed to characterize a new radioligand of general interest to scientists interested in the biology of the muscarinic receptors. Data were analyzed using GraphPad Prism (GraphPad Software, San Diego, CA) with the following equations and were not constrained at the top or bottom.

**Saturation binding**: nonspecific binding at each radioligand concentration was subtracted from total binding and the data were fit to the following equation:

\[
\text{Specific Binding} = \frac{(B_{\text{max}}*[L])}{(K_d+[L])}, \quad \text{where } [L] \text{ is the concentration of free radioligand in nM.}
\]

**Competition binding**: Data were analyzed using the Cheng Prusoff equation:

\[
K_i = \frac{IC_{50}}{1 + [\text{radioligand}]/K_d}.
\]

**Association kinetics**: \(Y = Y_0 + (\text{Plateau}-Y_0) \times (1-\exp(-K \times x)), \) where \(Y_0\) is the \(Y\) value when time \((X)\) is 0, expressed as \% \([^3H]VU6013720\) binding; plateau is the \(Y\) value at infinite time, expressed as \% \([^3H]VU6013720\) binding; \(K\) is the rate constant, expressed as inverse minutes; \(\text{Tau}\) is the time constant, expressed in minutes; half-time is calculated as \(\ln(2)/K\), expressed as minutes, and span is the difference between \(Y_0\) and Plateau, expressed as \% \([^3H]VU6013720\) binding.

**Dissociation kinetics**: One-phase or two-phase exponential decay

One phase: \(Y = (Y_0-\text{NS}) \times \exp(-K \times X) + \text{NS}, \) where \(Y_0\) is the binding at time zero, expressed as \% \([^3H]VU6013720\) binding; \(\text{NS}\) is nonspecific binding at infinite time,
expressed as % $[^3]H$VU6013720 binding; $K$ is the rate constant in inverse minutes; half-time equals $\ln(2)$ divided by $K$.

Two phase:

$$\text{Span}_{\text{Fast}} = (Y_0 - \text{Plateau}) \times \text{Percent}_{\text{Fast}} \times 0.01$$

$$\text{Span}_{\text{Slow}} = (Y_0 - \text{Plateau}) \times (100 - \text{Percent}_{\text{Fast}}) \times 0.01$$

$$Y = \text{Plateau} + \text{Span}_{\text{Fast}} \times \exp(-K_{\text{Fast}} X) + \text{Span}_{\text{Slow}} \times \exp(-K_{\text{Slow}} X)$$

where $Y_0$ is the $Y$ value when $X$ (time) is zero, expressed as % $[^3]H$VU6013720 binding; plateau is the $Y$ value at infinite time, expressed in minutes; $K_{\text{Fast}}$ and $K_{\text{Slow}}$ are the rate constants, expressed as inverse minutes; time units; $\text{Tau}_{\text{Fast}}$ and $\text{Tau}_{\text{Slow}}$ are the two time constants, expressed as time; Half-life (fast) and Half-life (slow) are computed as $\ln(2)/K$ and expressed as minutes; PercentFast is the fraction of the span (from $Y_0$ to Plateau) accounted for by the faster of the two components.

Data from individual experiments were fit independently and then presented as the $pIC_{50}$ or $pK_i/pK_d$ as Mean ± Standard Deviation. Data shown in the figures represents either combined data from multiple experiments or, in some cases, as a data set from a typical experiment as noted in the various figure legends.
Results

**VU6013720 is a selective M₄ antagonist.** VU6013720 (Figure 1) binding affinities and subtype selectivity at the five rat muscarinic receptor subtypes were assessed using binding competition against the non-selective orthosteric radioligand [³H]N-methylscopolamine (NMS) in membranes made from CHO-K1 cells expressing each receptor subtype (Table 1 and Figure 2). VU6013720 fully displaced [³H]NMS binding from rat M₄ receptors with a pKᵢ of 8.8 ± 0.1 (n=4), while pKᵢ values were 6.9 ± 0.2 (n=4) for M₂ and affinity values were above a micromolar for M₁, M₃, and M₅. The ratio of the nanomolar Kᵢ value at the receptor with the closest binding affinity, M₂, versus M₄ was 80-fold. These competition binding data, together with IC₅₀ values from calcium mobilization assays (Moehle et al., 2021), demonstrate that VU6013720 is a highly selective M₄ antagonist with single-digit nanomolar affinity, and possesses the characteristics needed to move to further characterization as a radioligand.

[³H]VU6013720 binds specifically to M₄ receptors with high affinity. We radiolabeled VU6013720 and determined the interaction of this novel radioligand across the muscarinic receptor family using saturation binding assays performed with cell membranes expressing the various receptor subtypes. Nonspecific binding was determined in the presence of a structurally related antagonist, VU6013719 (Supp. Fig. 1; synthesis provided in Supp. Methods), or the nonselective muscarinic antagonist, atropine. [³H]VU6013720 bound specifically to the rat M₄ receptor with a pKᵢ of 9.7 ± 0.2 (Figure 3A and Table 2) and a Bᵢₘₐₓ of 4800 ± 500 fmol/mg using VU6013719 as the nonspecific binding control (n=4) and a pKᵢ of 9.5 ± 0.2 and a Bᵢₘₐₓ of 4100 ± 500 fmol/mg using atropine (Figure 3B and Table 2, n=3). These values were slightly different when compared to the pKᵢ determined with [³H]NMS (8.8 versus 9.7/9.5), which
may suggest differential interaction of \[^{3}H\]VU6013720 and \[^{3}H\]NMS with the receptor. In contrast, \[^{3}H\]VU6013720 exhibited no specific binding at the rat M\(_{1}\), M\(_{2}\), M\(_{3}\), and M\(_{5}\) receptor subtypes (Figure 3C-F, n=3 per receptor). These data suggest that \[^{3}H\]VU6013720 binds specifically to the M\(_{4}\) receptor with high affinity. We performed a comparison of the pK\(_{i}\) value of atropine with both \[^{3}H\]NMS and \[^{3}H\]VU6013720 and found the pK\(_{i}\) to be statistically different between the two radioligands (8.7 ± 0.1 with \[^{3}H\]NMS and 9.0 ± 0.1 with \[^{3}H\]VU6013720, Mean ± S.D., **p=0.009). As these two radioligands are of different structure, this would suggest that the binding sites of the two, or the binding pose adopted by atropine in the presence of each radioligand, are not exactly the same. We also compared B\(_{\text{max}}\) values for saturation binding studies of \[^{3}H\]NMS and \[^{3}H\]VU6013720 and determined B\(_{\text{max}}\) values of 4500 ± 700 and 4100 ± 500 fmol/mg (Supp. Fig. 2, paired Student’s t-test, p=0.219, n=3), respectively, suggesting that the two radioligands label a similar number of binding sites on the receptor. To assess cross-species binding of \[^{3}H\]VU6013720, saturation binding was also assayed with membranes expressing the human and mouse M\(_{4}\) receptors. Similar to the rat M\(_{4}\) receptor, \[^{3}H\]VU6013720 bound to these receptors with high affinity, with pK\(_{d}\) values of 10 ± 0.1 (n=3) at the human M\(_{4}\) receptor and 9.7 at the mouse M\(_{4}\) receptor, respectively (n=1, Supp. Fig. 3 and Table 2).

**Kinetic evaluation of \[^{3}H\]VU6013720.** To fully characterize this novel radioligand, kinetic binding studies examining association and dissociation were next conducted using a concentration of radioligand calculated to be within two-fold of the K\(_{d}\) in each experiment. Association experiments demonstrated that \[^{3}H\]VU6013720 binding reached equilibrium within 10 minutes (Figure 4A) with an association on-rate (K\(_{on}\)) of
1.7 ± 0.1 nM⁻¹min⁻¹ (n=3). [³H]VU6013720 dissociated completely by adding 10 µM VU6013719 or atropine, although with distinct time courses (Figure 4B; Table 3, n=3). The data for both of these compounds was better fit to a two-site decay model versus a single site. We observed faster dissociation with the structural analog VU6013719 than atropine (Table 3), suggesting that [³H]VU6013720 and VU6013719 may share a binding site that is distinct or overlapping that of atropine. We also examined dissociation kinetics induced by the positive allosteric modulator, VU0467154 (Bubser et al., 2014). VU0467154 induced partial but saturable displacement of [³H]VU6013720, similar to what we had previously observed with [³H]NMS (Bubser et al., 2014). This suggests that VU0467154 is binding to an allosteric site but induces negative cooperativity with [³H]VU6013720 by partially displacing the ligand.

[³H]VU6013720 specifically binds to M₄ in the rodent brain. One of the main advantages of a highly specific radioligand is sensitivity for detection of receptors in native tissues. To determine the utility of [³H]VU6013720 for detecting M₄ receptors in brain tissue, membranes were prepared from the striatum and cortex of rats. As shown in Figure 5, saturation binding assays detected specific binding of [³H]VU6013720 to homogenates from rat striatum and cortex with high affinities (pKᵋ = 9.0 ± 0.2 in cortex, (n=3, A)) and 9.3 ± 0.2 in striatum, (n=3, B)). B_max values of 260 ± 110 (cortex) and 350 ± 60 (striatum) fmol/mg protein were also determined (n=3). We would note that, when the pKᵋ values for rat M₄ cells, cortex, and striatum were compared using a one-way ANOVA, there was a significant difference (p<0.05) between rat M₄ cells and the cortex, but not the striatum. This may reflect distinct microenvironments between an in vitro cell system versus the brain.
To further confirm that these binding sites were indeed M₄ receptors, saturation binding was also performed with membranes made from both wildtype (WT) and Chrm4 knockout mouse cortex. As shown in Figure 5C and D, [³H]VU6013720 specifically bound to WT mouse cortex and exhibited essentially no binding to cortical membranes from Chrm4 knockout mice. These data demonstrate that [³H]VU6013720 is a useful tool for quantification of M₄ receptors in brain tissue.

**Discussion**

M₄ muscarinic receptors are highly expressed in the brain (e.g., striatum, cortex, thalamus) and have been demonstrated to be involved in several CNS diseases (Pancani et al., 2015, Felder et al., 2018, Moehle et al., 2019, Teal et al., 2019, Dean et al., 2020, Foster et al., 2021, Moehle et al., 2021). Lack of selective muscarinic receptor antagonists has prevented the utilization of anti-muscarinic therapy due to intolerable adverse effects, and tools that assist with drug discovery efforts, such as highly specific radioligands, may open new avenues for exploration. Here, we demonstrate that [³H]VU6013720 is a novel, high-affinity, highly specific antagonist radioligand for the M₄ receptor. [³H]VU6013720 selectively binds to the M₄ receptor with pKᵢ values of 9.5 at rat, 10 at human, and 9.7 at mouse M₄ receptors. Based on this profile, we used this high-affinity radioligand to quantify M₄ receptors in brain tissue by saturation binding with brain tissue homogenates.

There are some interesting, subtle findings from the current studies. While the majority of our data point to an orthosteric or overlapping binding site for VU6013720, the pKᵢ for cold VU6013720 is 8.8 at rat M₄ using [³H]NMS binding while the pKᵢ was
calculated to be 9.5 with atropine and 9.7 with VU6013719. Additionally, competition binding studies using VU6013720 itself revealed a pKᵢ of 9.5 (data not shown), which is close to the Kᵩ value. When considered in the context of the significant shift of the pKᵢ of atropine between [³H]NMS and [³H]VU6013720, it suggests that these two radioligands might make distinct contacts within similar, but not identical, binding sites. This is further supported by the dissociation kinetics experiments in which the dissociation of [³H]VU6013720 differs in the presence of atropine versus VU6013719. Other factors may impact these results, such as differences in residence time. The discrepancy between pKᵢs for atropine with the two radioligands at M₄ also suggested the possibility that, due to the high sequence conservation, a similar leftward affinity shift might exist for M₂ which might have resulted in specific binding at M₂; however, we did not observe any saturable binding to this receptor subtype.

We previously reported a 30-fold selectivity difference for the human and rat M₄ receptors for VU6013720 in a chimeric G protein assay that allows the M₄ receptor to couple to calcium mobilization (Moehle et al., 2021). In contrast, our data here show that there is only a three-fold difference in affinity between the human and rat M₄ receptors. We currently do not have an explanation for this difference, although there are several possibilities. First, the functional assays include acetylcholine whereas the binding assays do not. If VU6013720 is truly orthosteric, this should not be a confound; however, if ACh/NMS and VU6013720 have subtly different binding sites and these binding sites are slightly different between the two species, this could contribute to these discrepancies. Additionally, if VU6013720 is actually an inverse agonist, its functional potency may be dissociated from its binding affinity due to G protein
interactions or other protein/protein interactions that are induced in the cells when the receptor is functionally assessed. We also do not currently know if there is a species difference in terms of contact with the chimeric G protein used in these studies, Gqi5, and whether Gqi5 impacts receptor conformation in binding studies. Future experiments around this issue could include assessment of VU6013720 effects in an assay in which constitutive activity of the receptor might be observed, such as inositol phosphate accumulation or cAMP generation, to determine if there is a distinction in inverse agonist activity, and further exploration of the effects of various G proteins or signaling partners.

To our knowledge, this is the first development of a broadly useful, highly specific antagonist radioligand with high affinity for M₄ receptors, and the development of [³H]VU6013720 is anticipated to be a powerful tool for M₄ receptor studies. Previous approaches to investigate the tissue distribution of the M₄ receptor include RNA detection techniques, or by antibody, with confirmation using knockout tissue (Brann et al., 1988, Buckley et al., 1988, Weiner et al., 1990, Levey et al., 1991, Levey 1993, Vilaro et al., 1993, Wolfe et al., 1995, Tice et al., 1996, Goméza et al., 1999). Although these studies provide important insight into relative distribution, detected RNA levels are not always consistent with protein expression levels, and antibodies do not provide the same level of quantitation that is possible using specific radioligand binding. As we have shown in this report, our studies using [³H]VU6013720 were able to confirm and quantify the M₄ receptor protein expression pattern and receptor density levels in native brain tissues. For this reason, we anticipate that [³H]VU6013720 will be an important tool for investigating the M₄ receptor distribution in native tissues and may also be
useful for interrogating potential differences that may occur in different disease states or models.

In the past several years, several M4 receptor radioligands have been reported, the M2/M4 PAM [3H]LY2119620 (Schober et al., 2014), and the selective M4 PAMs [11C]-VU0467485/AZ13713945 (Deng et al., 2019), [18F]-M4R-1911 (Deng et al., 2020), [11C]MK-6884 (Tong et al., 2020), and [18F]12 (Haider et al., 2023). However, these radioligands have been developed from M4 positive allosteric modulator (PAM) scaffolds and bind to allosteric sites on the M4 receptor. [3H]VU6013720 is the first reported selective radiolabeled M4 receptor antagonist. Therefore, it is a unique tool that could be paired with PAM radioligands to study compound binding sites as well as for the development of future M4 antagonists and modulators. Additionally, the compound could potentially be used to map acetylcholine levels in the brain relevant for M4 function and be useful for in vitro screening of M4 antagonists for therapeutic treatment of movement disorders and may serve as a starting point for development of antagonists with higher affinity and favorable drug metabolism and pharmacokinetic properties to eventually lead to PET ligands.

In conclusion, we have developed the first highly selective M4 receptor antagonist radioligand with high affinity for the rodent and human M4 receptors, [3H]VU6013720. A reliable tool will lead to a better understanding of the physiology and pathophysiology associated with this important signaling protein and facilitate the support of drug development for neurological diseases for which M4 modulators are predicted to exhibit efficacy, such as Parkinson’s disease, essential tremor, and schizophrenia.
Acknowledgments

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Data Availability Statement

The authors declare that all processed data supporting the findings of this study are available within the paper and its Supplemental Data. Raw data are available on request from the corresponding author.

Author Contributions

Participated in research design: Qi, Rodriguez, Dickerson, Bender, Moehle, Lindsley, Rook, Niswender

Conducted experiments: Qi, Kling, Billard, Peng, Dickerson, Bender

Contributed new reagents or analytic tools: Bender, Engers, Lindsley

Performed data analysis: Qi, Rodriguez, Dickerson, Rook, Niswender

Wrote or contributed to the writing of the manuscript: Qi, Rodriguez, Niswender with input from all authors
References


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Footnotes

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Figure Legends

Figure 1. Structure of VU6013720. Cold VU6013720 was labeled using nonspecific hydrogen/tritium exchange with 99\% tritium gas (RC Tritec, Switzerland) with a specific activity of 23.6 Ci/mmol.

Figure 2. Binding selectivity of VU6013720 at muscarinic acetylcholine receptor subtypes using [\(^{3}\text{H}\)]NMS. VU6013720 and atropine competition binding curves at various muscarinic acetylcholine receptor subtypes were performed using the orthosteric muscarinic radioligand, [\(^{3}\text{H}\)]-N-methylscopolamine. Data are the Mean ± SD from a representative of four independent experiments performed in triplicate.

Figure 3. Among the five muscarinic receptors, [\(^{3}\text{H}\)]VU6013720 binds specifically to the rat M\(_{4}\) receptor. Total and specific binding for [\(^{3}\text{H}\)]VU6013720 was determined using cells expressing each of the rat muscarinic receptors. [\(^{3}\text{H}\)]VU6013720 bound specifically to the rat M\(_{4}\) receptor in a saturable manner using either VU6013719 (A) or atropine (B) as nonspecific binding controls, whereas no detectable binding was observed at rat M\(_{1}\), M\(_{2}\), M\(_{3}\), or M\(_{5}\) using atropine to determine nonspecific binding (C-F). Data shown are the Mean ± SD of a representative of at least three independent experiments performed in triplicate.

Figure 4. Kinetic characterization of the [\(^{3}\text{H}\)]VU6013720 radioligand. Time course experiments defining association and dissociation of [\(^{3}\text{H}\)]VU6013720 at rat M\(_{4}\) membranes were performed at room temperature. A. Association was initiated by addition of [\(^{3}\text{H}\)]VU6013720 to membranes at the indicated time points before filtration. B. Dissociation experiments were performed by allowing [\(^{3}\text{H}\)]VU6013720 to equilibrate with membranes for 2 hours; at this point, a 10 μM final concentration of VU6013719
(white), atropine (black), or VU0467154 (red) was added at designated times before terminating the reaction by filtration. Data are the Mean ± SD of three independent experiments performed in triplicate.

**Figure 5.** $[^{3}H]VU6013720$ binds specifically to rat brain cortical and striatal tissue as well as to cortical membranes from WT but not from *Chrm4* knockout mice. Rat cortical (A) and striatal (B) homogenates were incubated with $[^{3}H]VU6013720$ in the presence and absence of 10 μM atropine to determine total and nonspecific binding. $[^{3}H]VU6013720$ bound to rat cortical membranes with a pK$_d$ of 9.0 ± 0.2 and a B$_{max}$ of 260 ± 110 fmol/mg of protein and to rat striatal membranes with a pK$_d$ of 9.3 ± 0.2 and a B$_{max}$ of 350 ± 60 fmol/mg of protein. Data are Mean ± SD and a representative of three independent experiments performed in triplicate. Cortical homogenates from WT (C) and *Chrm4* knockout mice (D) were incubated with $[^{3}H]VU6013720$ in the presence and absence of 10 μM atropine to determine total and nonspecific binding. $[^{3}H]VU6013720$ binds to cortical membranes from WT mice with a pK$_d$ of 9.0 ± 0.4 and a B$_{max}$ of 240 ± 90 fmol/mg of protein, whereas little specific $[^{3}H]VU6013720$ binding was detected in cortical homogenates from *Chrm4* knockout mice. Data are the Mean ± SD and a representative of three independent experiments performed in duplicate or triplicate.
Tables

<table>
<thead>
<tr>
<th></th>
<th>pKi (Mean ± SD)</th>
<th>Ki (nM)</th>
<th>Selectivity (fold over M4 Ki)</th>
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<tbody>
<tr>
<td>M4</td>
<td>8.8±0.0</td>
<td>1.6</td>
<td></td>
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<tr>
<td>M1</td>
<td>5.7±0.1</td>
<td>2000</td>
<td>1300</td>
</tr>
<tr>
<td>M2</td>
<td>6.9±0.2</td>
<td>130</td>
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<tr>
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<td>M5</td>
<td>5.5±0.1</td>
<td>3100</td>
<td>2000</td>
</tr>
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Table 1. Characterization of the binding specificity of VU6013720 using the orthosteric muscarinic antagonist radioligand [3H]-N-methylscopolamine. pKi values were obtained from [3H]-NMS competition binding assays with CHO-K1 cell membranes expressing different rat muscarinic receptor subtypes. Data represent the Mean ± SD of four independent experiments performed in triplicate and refer to the curve fits in Figure 2.

<table>
<thead>
<tr>
<th>[3H]-VU6013720</th>
<th>Human M4[#]</th>
<th>Mouse M4[#]</th>
<th>Rat M4[#]</th>
<th>Rat M4[@]</th>
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<tbody>
<tr>
<td>pKi</td>
<td>10±0.1</td>
<td>9.7</td>
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<tr>
<td>Bmax (fmol/mg)</td>
<td>1200±40</td>
<td>1400</td>
<td>4800±500</td>
<td>4100±500</td>
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</table>

Table 2. Summary of binding properties of [3H]VU6013720 at multiple species of M4 receptor. Data represent Mean ± SD from at least 4 experiments performed in quadruplicate (human and rat) and 1 experiment (mouse) performed in triplicate. # and @ denote the use of VU6013719 and atropine, respectively, for non-specific binding. Data refer to saturation binding curves shown in Figure 3 and Supplemental Figure 3.
Table 3. Dissociation kinetics of [³H]VU6013720 induced by multiple ligands.

Dissociation was performed by adding 10 μM of atropine, VU6013719, or VU0467154 at designated times. Data were fit with both one- and two-phase decay curves. For atropine and VU6013719, two-site fits were the preferred model; for VU0467154, a one-site model was preferred. *p<0.05, **p<0.01, ***p<0.001 between atropine and VU6013719.

<table>
<thead>
<tr>
<th></th>
<th>% Fast (min⁻¹)</th>
<th>K_{off} fast (min⁻¹)</th>
<th>Half-life fast (min)</th>
<th>Half-life slow (min)</th>
<th>Plateau (min)</th>
<th>R²</th>
</tr>
</thead>
<tbody>
<tr>
<td>Atropine</td>
<td>51±5</td>
<td>0.56±0.39</td>
<td>0.05±0.01</td>
<td>1.6±0.9</td>
<td>15±3</td>
<td>1.0±0.7</td>
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<tr>
<td>VU6013719</td>
<td>44±8</td>
<td>2.0±0.5*</td>
<td>0.17±0.02***</td>
<td>0.40±0.10</td>
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<td>0.60±0.18</td>
<td></td>
<td>1.2±0.3</td>
<td>58±5</td>
<td>0.84±0.05</td>
</tr>
</tbody>
</table>
Figure 1

VU6013720

[Chemical structure diagram]

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Figure 3

(A) rM4

(B) rM4

(C) rM1

(D) rM2

(E) rM3

(F) rM5

[Graphs showing binding (fmol/mg) of [3H]VU6013720 at different concentrations for each receptor subtype with TB and Atropine (NSB) as controls.]
Figure 4

A) Association

B) Dissociation

%[^3]H]VU6013720 Binding (%)

Time (min)

%[^3]H]VU6013720 Binding (%)

Time (min)

Atropine
VU6013719
VU0467154

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Figure 5

A B

Cortex Striatum

Specific Binding (fmol/mg) Specific Binding (fmol/mg)

0.0 0.5 1.0 1.5 2.0 2.5 3.0 3.5 0.0 0.5 1.0 1.5 2.0 2.5 3.0 3.5


0 100 200 300 400 500 0 100 200 300 400 500

B C

Wildtype Chrm4 Knockout

Specific Binding (fmol/mg) Specific Binding (fmol/mg)

0 3 6 9 12 15 0 3 6 9 12 15


0 200 400 600 800 1000 0 200 400 600 800 1000

D

Chrm4 Knockout

TB Atropine (NSB)

Specific Binding (fmol/mg) Binding (fmol/mg)

0 3 6 9 12 15 0 3 6 9 12 15